

## Antibacterial Evaluation of the Methanolic and Chloroform Extract of *Glycine max* (Soybean)

Megala K.<sup>1</sup>, N. Subramanian<sup>2</sup> and N. Ramadass\*<sup>3</sup>

Received: 10 June 2020 | Revised accepted: 26 Dec 2020 | Published online: 05 Jan 2021  
© CARAS (Centre for Advanced Research in Agricultural Sciences) 2021

### ABSTRACT

*Glycine max* is one of the most popular and nutritious foods with high antimicrobial effects. The present work was done to study the antibacterial activity of Methanolic and chloroform extracts of *Glycine max* against gram negative bacteria such as *Escherichia coli*, *Salmonella typhi* and *Pseudomonas aeruginosa*, the gram-positive bacteria such as *Bacillus subtilis* and *Staphylococcus aureus*. The concentrations of the seed used were 500 µg/ml, 750 µg/ml and 1000 µg/ml respectively. At these concentrations the extract inhibited the growth of *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella thphi*, *Bacillus subtilis* and *Staphylococcus aureus* and produced zone of inhibition ranging between 7 mm to 10 mm in Methanolic extract and 6 mm to 14 mm in chloroform extract. The antibacterial activity reported by the Soybean extract may be due to the presence of the phytochemical compounds present in the seeds. Highest zone of inhibition 10 mm was noted in *Staphylococcus aureus* at 1000 µg/ml and 750 µg/ml concentration and lowest zone of inhibition 7 mm was found in *Escherichia coli* at 750 µg/ml and 500 µg/ml, *Bacillus subtilis* at 500 µg/ml, and *Pseudomonas aeruginosa* at 750 µg/ml and 500 µg/ml respectively.

**Key words:** *Glycine max*, Methanolic, Chloroform, Antibacterial, Zone of inhibition

Medicinal plants have continued to attract attention in the global search for effective antibacterial agents that can combat resistant pathogens that have been rendering many conventional drugs absolute in the treatment of infections. The antibacterial compounds produced by plants are active against plant and human pathogenic microorganisms [1]. The active principles isolated from plants appear to be one of the important alternatives when compared with many substandard orthodox synthetic medicines because of their less or no side effect and better bioavailability. Plant extracts have been studied against pathogens for years for assays to detect new and previously undiscovered antibacterial from plant sources [2].

In spite of the considerable progress of medicine, treatment of infectious diseases faced with different unfavorable problems. Occurrence of severe levels of the antibiotic resistance is the main issue facing medical practitioners [3]. Antimicrobial resistance threatens the effective prevention and treatment of an ever-increasing range of infections caused by bacteria [4]. It is an increasingly serious threat with global public health impact that requires action across all government sectors and society [5]. Resistant bacteria caused more severe clinical diseases for longer period of time which causes abundant economic losses [6]. Documented data revealed that pathogenic bacteria and especially *Staphylococcus aureus* (*S. aureus*), *Escherichia coli* (*E. coli*), *Bacillus cereus* (*B. cereus*), *Pseudomonas*

*aeruginosa* (*P. aeruginosa*), *Listeria monocytogenes* (*L. monocytogenes*), *Klebsiella pneumoniae* (*K. pneumoniae*) and *Salmonella typhi* (*S. typhi*) exhibited the high levels of resistance against aminoglycosides, tetracyclines, lincosamides, macrolides, beta-lactams, quinolones, fluoroquinolone and cepheems groups of antibiotics [7]. Therefore, therapeutic and pharmacological factories tried to use from novel sources for antimicrobial agents to produce strong antibiotic drugs. Application of medicinal plants for producing of antimicrobial agents had an ancient history [8]. Soybean (*Glycine max*) belongs to a large botanical family Leguminosae which classically grows in tropical, subtropical and temperate climatic regions like Iran [9]. Soybean is an oilseed and consists of 20% oil content. It is considered as the most significant crop for the production of edible oil. Soybean plant holds great importance in today's world mainly because of its high protein and lipid content and other major constituents including vitamins, minerals, fatty acids and other essential nutritional factors [10].

Besides. Soybean holds much importance from medicinal perspective. Recorded data revealed the high antimicrobial and antioxidant content of the *G. max*. Soybean seeds are rich in proteins, isoflavones and phytoestrogens, while Genistein, a soy isoflavone, has also been reported to possess anti-cancerous, antioxidant, anti-inflammatory and anti-osteoporosis effects and is considered as potential compound for metabolic disorders' treatment. Many infectious diseases have been known to be treated with herbal remedies throughout the history of mankind. Natural products, either as pure compounds or as standardized plant extracts, provide unlimited opportunities for new drug leads because of the unmatched availability of chemical diversity [11]. There is a

\*N. Ramadass  
nramadass1974@gmail.com

<sup>1-3</sup>P.G and Research Department of Zoology, Arignar Anna Government Arts College, Cheyyar - 604 407, Tamil Nadu, India

continuous and urgent need to discover new antimicrobial compounds with diverse chemical structures and novel mechanisms of action for new and re-emerging infectious disease. Therefore, researchers are increasingly turning their attention to folk medicine, looking for new leads to develop better drugs against microbial infections.

In addition to the high content of protein and fibres, legumes contain saponins, which are bioactive compounds. Saponins are a group of surface-active glycosides, which distinguishes them from other glycosides [12]. The highest concentration of saponins occurs in soybeans ( $6500 \text{ mg kg}^{-1}$ ) [13]. The physiological role of saponins in plants is not yet completely cleared up. Saponins are secondary metabolites and play a role in the protection of plants against microorganism. Many saponins show strong antibacterial activities. As saponins are probably a part of plants' defence systems, they have been included in a group of protective molecules in plants called phytoprotectant [14]. Soybean is an oilseed and consists of 20% oil content. Because of this characteristic, it is considered as the most important crop for the production of edible oil. The top five countries with maximum soybean production at present are USA, Brazil, Argentina, China and India, and these contribute to 92% of the world's soybean production. Soy contains significant amounts of all the essential amino acids for humans, and so is a good source of protein, which is generally used to prepare extracts or powders for medicinal use. The current investigation aims to explore scientifically the antibacterial potential of crude and pure saponin seed extract of *Glycine max* plant.

## MATERIALS AND METHODS

### Plant materials

*Glycine max* L., subjected to Magnoliophyta phylum, Magnoliopsida class, Fabales order, Fabaceae family [15]. Soybean seeds were purchased from the local market in Kanchipuram district of Tamil Nadu and identified at P.G. and Research Department of Botany, Arignar Anna Government Arts College, Cheyyar, Thiruvannamalai district of Tamil Nadu, India.

### Preparation of extracts

Soybean seeds were grinded to powder using an electrical blender two different solvent were used chloroform and methanol to obtain different extracts.

### Methanol and chloroform extracts

Plant materials (seeds) were extracted using a Soxhlet extractor with solvents beginning with methanol than chloroform, each extraction was carried out by 8-10 hours continuously until the used solvents turned pure and colorless. The solvents were removed using a rotary vacuum evaporator at  $40^\circ\text{C}$  to give concentrated extracts which were frozen and freeze dried until use.

### Tested bacterial strains

The microorganism used induced two-gram positive bacteria *Staphylococcus aureus*, *Bacillus subtilis* and three-gram negative bacteria *Pseudomonas aeruginosa*, *Escherichia coli* and *Salmonella typhi* along with standard Amphotericin (20  $\mu\text{l}/\text{disc}$ ). Stock cultures were maintained at  $4^\circ\text{C}$  on nutrient agar slant. Active cultures or experiments were prepared by transferring a loop full of culture from the stock cultures into the test tubes containing nutrient broth, that were incubated at 24 hrs at  $37^\circ\text{C}$ . The assay was performed by agar disc diffusion method.

### Agar disc diffusion method (NCCLS, 1993)

Antibacterial of sample was determined by disc diffusion method on Muller Hinton agar (MHA) medium. Muller Hinton agar (MHA) medium is poured into the Petri plate. After the medium was solidified, the inoculums were spread on the solid plates with sterile swab moistened with the bacterial suspension. The disc was placed in MHA plates and add 20  $\mu\text{l}$  of sample (concentration 1000  $\mu\text{g}$ , 750  $\mu\text{g}$ , and 500  $\mu\text{g}$ ) were placed in the disc. The plates were incubated at  $37^\circ\text{C}$  for 24 hrs. Then the antimicrobial activity was determined by measuring the diameter of zone of inhibition.

## RESULTS AND DISCUSSION

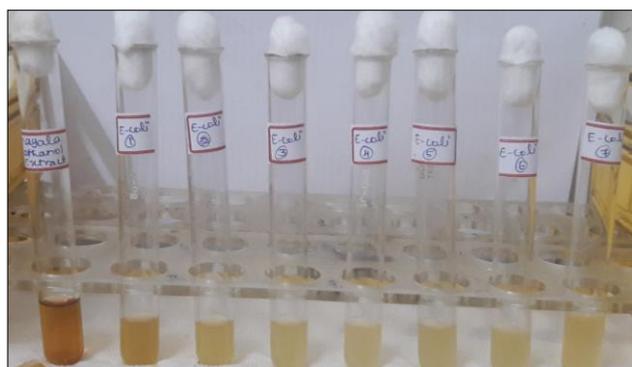
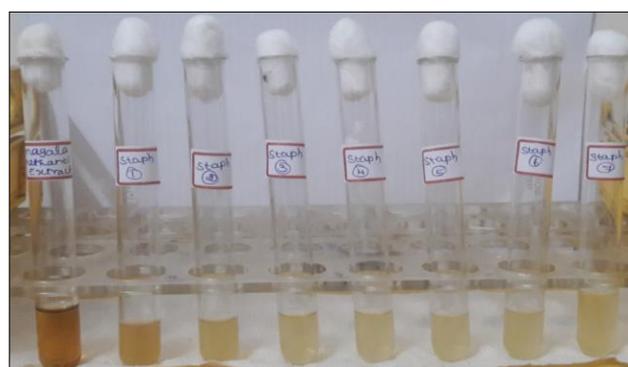
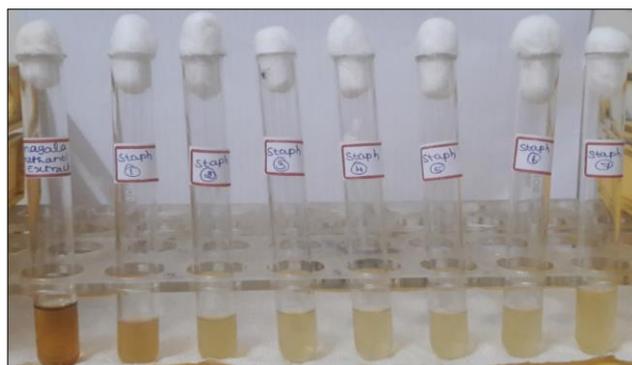
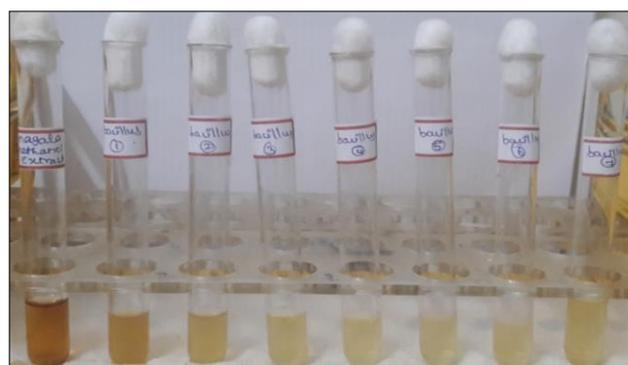
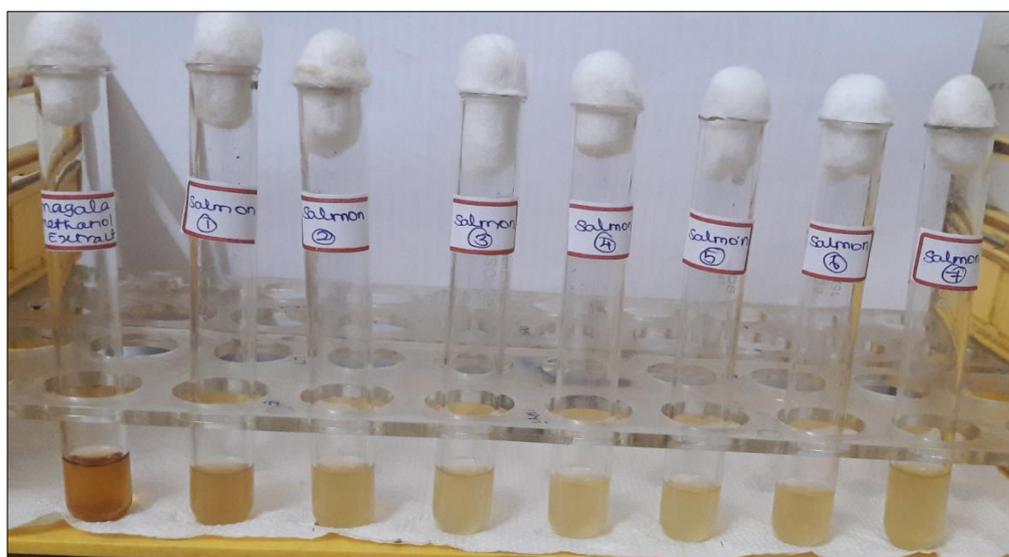
Study results showed various inhibitory effects of different extracts from *Glycine max* seeds against tested bacteria via agar-well diffusion method (Table 1-2, Fig 1-2) revealed the methanol. Extract was active against *Staphylococcus aureus* with zone of inhibition 10 mm in diameter at 1000  $\mu\text{g}/\text{ml}$  and 750  $\mu\text{g}/\text{ml}$  respectively. *E. coli* showed 9 mm, 7 mm and 7 mm at 1000  $\mu\text{g}/\text{ml}$ , 750  $\mu\text{g}/\text{ml}$  and 500  $\mu\text{g}/\text{ml}$ . the *Bacillus* showed 9 mm, 9 mm and 7 mm at 1000  $\mu\text{g}/\text{ml}$ , 750  $\mu\text{g}/\text{ml}$  and 500  $\mu\text{g}/\text{ml}$ , *Staphylococcus aureus* showed 8 mm at 500  $\mu\text{g}/\text{ml}$ , *Salmonella* showed 9 mm, 9 mm and 8 mm at 1000, 750, 500  $\mu\text{g}/\text{ml}$  and *Pseudomonas aurtginea* showed 9 mm, 7 mm and 7 mm at 1000, 750, 500  $\mu\text{g}/\text{ml}$  respectively (Table 1, Fig 1).

Table 1 Antibacterial activity of soybean methanol extract on bacterial strains

| Organisms                     | Zone of inhibition (mm)      |                             |                             | Antibiotic (1mg/ml) |
|-------------------------------|------------------------------|-----------------------------|-----------------------------|---------------------|
|                               | 1000 $\mu\text{g}/\text{ml}$ | 750 $\mu\text{g}/\text{ml}$ | 500 $\mu\text{g}/\text{ml}$ |                     |
| <i>Escherichia coli</i>       | 9                            | 7                           | 7                           | 12                  |
| <i>Bacillus subtilis</i>      | 9                            | 9                           | 7                           | 14                  |
| <i>Staphylococcus aureus</i>  | 10                           | 10                          | 8                           | 11                  |
| <i>Salmonella typhi</i>       | 9                            | 9                           | 8                           | 16                  |
| <i>Pseudomonas aeruginase</i> | 9                            | 7                           | 7                           | 12                  |

Table 2 Antibacterial activity of *Glycine max* (soybean) chloroform extract on bacterial strains

| Organisms                     | Zone of inhibition (mm)      |                             |                             | Antibiotic (1mg/ml) |
|-------------------------------|------------------------------|-----------------------------|-----------------------------|---------------------|
|                               | 1000 $\mu\text{g}/\text{ml}$ | 750 $\mu\text{g}/\text{ml}$ | 500 $\mu\text{g}/\text{ml}$ |                     |
| <i>Escherichia coli</i>       | 10                           | 9                           | 6                           | 14                  |
| <i>Bacillus subtilis</i>      | 9                            | 9                           | 7                           | 14                  |
| <i>Staphylococcus aureus</i>  | 8                            | 7                           | 6                           | 11                  |
| <i>Salmonella typhi</i>       | 9                            | 7                           | 7                           | 17                  |
| <i>Pseudomonas aeruginase</i> | 14                           | 13                          | 10                          | 18                  |

*Escherichia coli**Staphylococcus aureus**Bacillus subtilis**Pseudomonas aeruginosa**Salmonella spp.*

The chloroform extract showed highest zone of inhibition 10 mm at 1000  $\mu\text{g/ml}$  in *Pseudomonas aeruginosa* followed by 13 mm and 10 mm at 750 and 500  $\mu\text{g/ml}$  in the same bacteria. The *E. coli* bacteria showed 10, 9, 6 mm *Bacillus subtilis* showed 9, 9, 7 mm, *Staphylococcus aureus* showed 8, 7, 6 mm and *Salmonella typhi* showed 9, 7, 7 mm at 1000, 750 and 500  $\mu\text{g/ml}$  concentrating the chloroform extract respectively (Table 2, Fig 2). From the (Table 2) it can be seen that chloroform extracts from soybean seeds indicated good inhibitory activity against all tested bacteria with inhibition zones between 7 to 14 mm in diameter.

The present work revealed that methanolic extract of *G. max* had significant antimicrobial effects on tested bacteria and especially *Staphylococcus aureus*, *Escherichia coli*, *Bacillus subtilis*, *Salmonella typhi* and *Pseudomonas aeruginosa* at concentration of 1000, 750 and 500  $\mu\text{g/ml}$

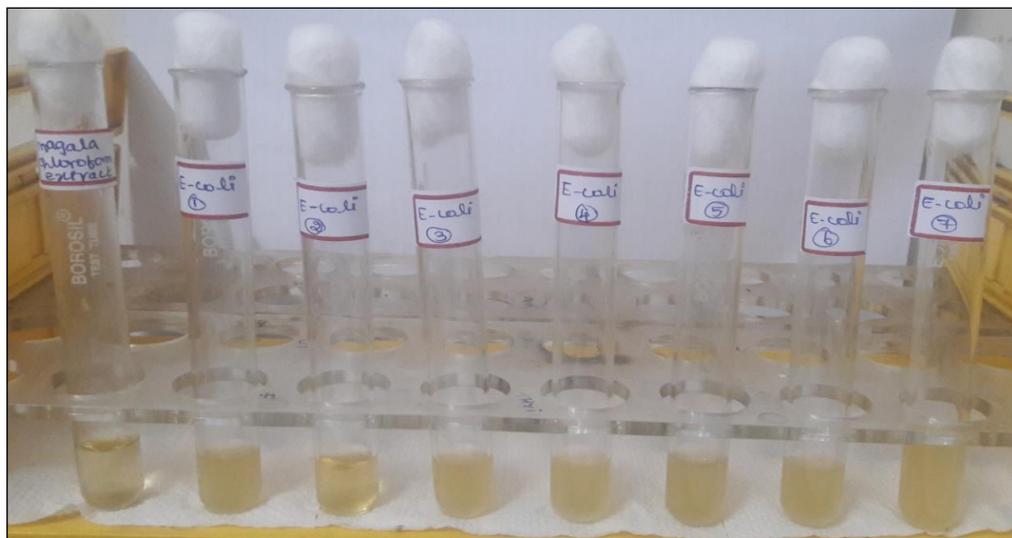
showed 10, 9, 8, 7 mm zone of inhibition [16]. They reported that *Glycine max* is a rich source of Phenols, Saponins, Micronutrients, Flavonoids and Poly saccharides which may be responsible for its high antimicrobial effects. Phenolic compounds are well known to have a negative effect on the growth of bacteria through inhibitory their nucleic acid synthesis, enzymatic activity, cytoplasmic membrane function and energy metabolism [17]. The chloroform extracts of *G. max* showed (6-14mm) 14, 13, 10, 9, 8, 7, 6 mm zone of inhibition at 1000, 750 and 500  $\mu\text{g/ml}$  of concentration. The highest zone of inhibition 14 mm was found in *Pseudomonas aeruginosa* at 1000  $\mu\text{g/ml}$  of concentration [18]. They showed that soybean isoflavones inhibited the nucleic acid synthesis of *S. aureus*, in addition phenolic acids were found at significant concentrations and their antimicrobial activity has been widely investigated. The synergic, coumaric, ferulic and vanilic acids

which are formed in the *Glycine max* extracts exhibited significant antimicrobial activity against gram-positive and gram-negative bacteria [19].

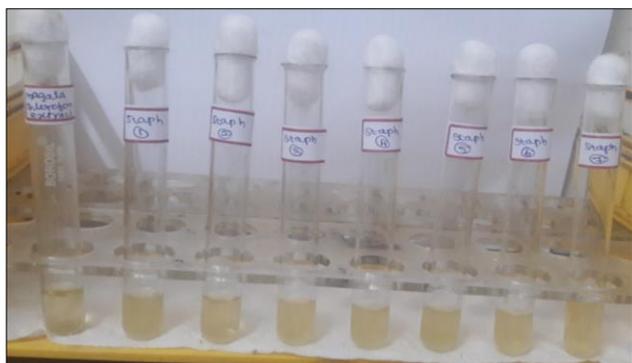
In the present study the gram-positive bacteria were more susceptible against *G. max* than gram-negative bacteria and the chloroform extracts showed high antibacterial activity than methanol extracts [20]. They reported that among tested bacteria the highest antimicrobial activity was observed against *L. monocytogenes*, *B. cereus* and *E. faecalis* with values around 50% inhibition even at the lowest concentration [21]. This is may be due to the assumption that gram-negative

bacteria are more resistant to antibacterial compounds due to their outer lopopoly saccharide membrane.

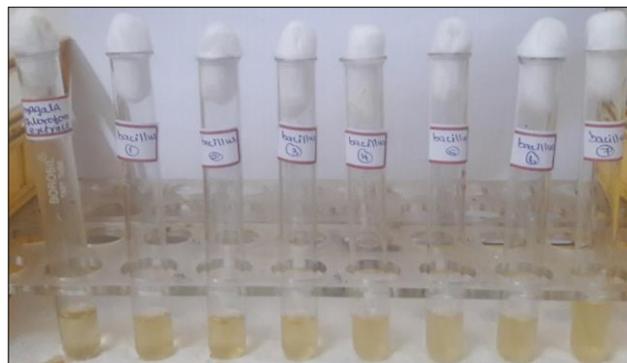
| MIC values of chloroform extract |  |
|----------------------------------|--|
| Name of the organism             | Minimum inhibitory concentration (µg/ml) |
| <i>Escherichia coli</i>          | 500                                      |
| <i>Staphylococcus aureus</i>     | 250                                      |
| <i>Bacillus subtilis</i>         | 250                                      |
| <i>Pseudomonas aeruginosa</i>    | 500                                      |
| <i>Salmonella spp.</i>           | 500                                      |



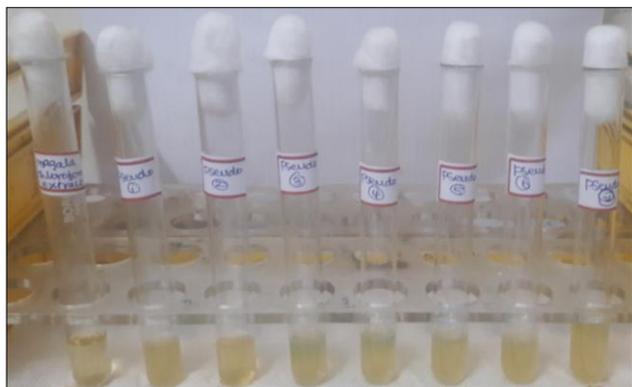
*Escherichia coli*



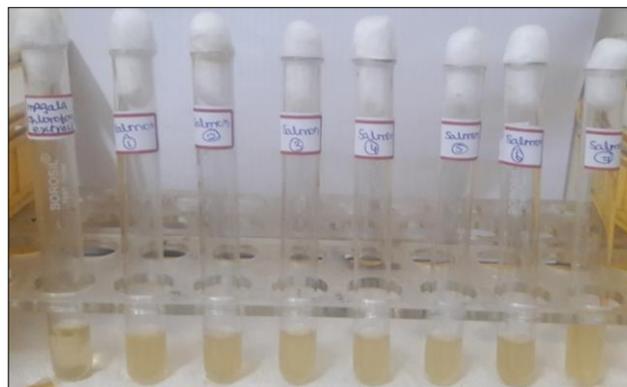
*Staphylococcus aureus*



*Bacillus subtilis*



*Pseudomonas aeruginosa*



*Salmonella spp.*

### CONCLUSIONS

The present study indicated that the Methonolic and Chloroform extracts of *Glycine max* (Soybean) showed high

antibacterial activity against both gram negative and positive bacteria such as *Escherichia coli*, *Salmonella typhi* and *Pseudomonas aeruginosa*, the gram-positive bacteria such as *Basillus subtilis* and *Staphylococcus aureus*. This antibacterial

activity may be due to presences of various phyto-constituents present in the plant seed extracts. So, from the study we concluded that the seed extracts of *Glycine max* could be used as alternative to anticancer, antibiotics, vaccines, and chemotherapeutic agents.

#### Acknowledgments

The authors are grateful to the Head, P.G and Research Department of Zoology and Principal, Arignar Anna Government Arts College, Cheyyar, for providing the laboratory facilities.

### LITERATURE CITED

1. Lee CK, Kin HKH, Shun K H. 1998. Screening and isolation of antibiotic resistance inhibitors from herb materials resistance inhibition of volatile components of Korean aromatic herbs. *Archives of Pharmaceutical Research* 21(1): 62-66.
2. Scazzocchio F, Cometa MF, Tomassini LM. 2001. Antibacterial activity of Hydrastiscanadensis extract and its major isolated alkaloids. *Planta Med.* 67: 561-564.
3. Dehkordi FS, Yazdani F, Mozafari J. 2014. Virulence factors, serogroups and antimicrobial resistance properties of *Escherichia coli* strains in fermented dairy products. *BMC Res Notes* 7: 217-220.
4. Dormanesh B, Safarpour Dehkordi F, Hosseini S. 2014. Virulence factors and o-serogroups profiles of uropathogenic *Escherichia coli* isolated from Iranian pediatric patients. *Iran Red Crescent Med Journal* 16: e14627.
5. Ranjbar R, Masoudimanesh M, Safarpour DF. 2017. Shiga (Vero)-toxin producing *Escherichia coli* isolated from the hospital foods; virulence factors, o-serogroups and antimicrobial resistance properties. *Antimicrob Resist Infect Control* 20: 6-4.
6. Momtaz H, Safarpour DF, Rahimi E. 2013. Virulence genes and antimicrobial resistance profiles of *Staphylococcus aureus* isolated from chicken meat in Isfahan province, Iran. *Journal of Appl Poult Research* 22: 913-921.
7. Owusu-Kwarteng J, Wuni A, Akabanda F. 2017. Prevalence, virulence factor genes and antibiotic resistance of *Bacillus cereus* sensu lato isolated from dairy farms and traditional dairy products. *BMC Microbiology* 17: 65.
8. Mirkamandar E, Shakibaie MR, Adeli S. 2012. *In-vitro* antimicrobial activity of *Salvadora persica* extract on *Helicobacter pylori* strains isolated from duodenal ulcer biopsies. *Microbiol Research* 3(e9): 38-41.
9. Carroll BJ, McNeil DL, Gresshoff PM. 1985 Isolation and properties of soybean [*Glycine max* (L.) Merr.] mutants that nodulate in the presence of high nitrate concentrations. *Proc Natl Acad Sci USA* 82: 4162-4166.
10. Villalobos Mdel C, Serradilla MJ, Martín A. 2016. Antioxidant and antimicrobial activity of natural phenolic extract from defatted soybean flour byproduct for stone fruit postharvest application. *Journal of Sci Food Agric* 96: 2116-2124.
11. Harbone JB. 1973. Phytochemical methods: A guide to modern techniques of plants analysis. Chapman and Hall Ltd. London. pp 49-88.
12. Sparg SG, Light ME, Van Staden J. 2004. Biological activities and distribution of plant saponins. *Journal of Ethnopharmacology* 94: 219-243.
13. Sagratini G, Zuo Y, Caprioli G, Cristalli G, Giardinà D, Maggi F, Molin L, Ricciutelli M, Traldi P, Vittori S. 2009. Quantification of soyasaponins I and βg in Italian lentil seeds by solidphase extraction (SPE) and high-performance liquid chromatography-mass spectrometry (HPLC-MS). *Journal of Agric Food Chem* 57: 11226-11233.
14. Francis G, Kerem Z, Makkar HPS, Becker K. 2002. The biological action of saponins in animal systems: a review. *Brit Journal of Nutrition* 88: 587-605.
15. Blackman SA, Obendorf RL, Leopold AC. 1992. Maturation proteins and sugars in desiccation tolerance of developing soybean seeds. *Plant Physiology* 100: 225-230.
16. Abdel-Hady H, Abdel-Wareth MTA, El-Wakil EA, Helmy EA. 2016. Identification and evaluation of antimicrobial and cytotoxic activities of *Penicilliumislandicum* and *Aspergillustamariethyle* acetate extracts. *World Jr. Pharm. Pharm. Sci.* 5(9): 2021-2039.
17. Tim TP Cushnie, Andrew JL. 2005. Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agent* 26(5): 343-356.
18. Wang S, Parsek MR, Wozniak DJ, Ma LZ. 2013. A spider web strategy of type IV pili-mediated migration to build a fibre-like Psl polysaccharide matrix in *Pseudomonas aeruginosa* biofilms. *Environ. Microbiol.* 15: 2238-2253.
19. Alves MJ, Ferreira ICFR, Froufe HJC, Abreu RMV, Martins A, Pintado M. 2013. *Antimicrobial activity of phenolic compounds identified in wild mushrooms, SAR analysis and docking studies.* *Journal of Applied Microbiology* 115(2): 346-357.
20. Saavedra MJ, Borges A, Dias C, Aires A, Bennett RN, Rosa ES, Simoes M. 2010. Antimicrobial activity of phenolics and glucosinolate hydrolysis products and their synergy with streptomycin against pathogenic bacteria. *Med Chem (Los Angeles)* 6: 174-183.
21. Kubmarawa D, Khan ME, Puna AM, Hassan M. 2008. Phytochemical Screening and antibacterial activity of extracts from *Pakia Clapperotnianakeay* against human pathogenic bacteria. *Jr. Med. Plants. Res.* 2(12): 352-355.