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Cultivation of *Haematococcus pluvialis* and *Nostochopsis* for the Enhancement of Proximate Analysis of *Catla catla* (Hamilton, 1822)

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ABSTRACT

In the present investigation, *H. pluvialis* and *Nostochopsis* were grown in the bold basal medium in controlled airconditioned culture room. *Catla catla* fish weighing 40 g were randomly distributed in 2 rectangular of 20 L capacity at a density of 6 fish per tank. Tank 1 fish were given normal feed, tank 2 fish were given *H. pluvialis* and *Nostochopsis* biomass (100 mg/kg b.w.). After 24 h gut evacuation period, all fish were anaesthetized and weighed. Out of six, three samples were randomly collected and stored at -20°C, until proximate analysis and lipid extraction. In normal feed treated *Catla catla* group, lipid content was found as 82.2±1.3, in *H. pluvialis* and *Nostochopsis* biomass-based feed group, lipid content was observed as 177.6±2.1 However, protein and ash content were reported as 133.6±1.2 and 14.4±2.1 in *H. pluvialis* and *Nostochopsis* biomass-based feed group. The present study reveals that *H. pluvialis* and *Nostochopsis* biomass can be used for the different aquatic feed formulations.

Key words: Haematococcus pluvialis, Astaxanthin, Nostochopsis feed, Formulations

Astaxanthin is a xanthophyll carotenoid [1]. Antioxidant activity of astaxanthin was found more than bcarotene and vitamin E [2]. Haematococcus pluvialis, freshwater green microalga is considered a potent producer of astaxanthin (3, 3'-dihydroxy-b, bcarotene-4, 4'-dione) [3-6]. Astaxanthin is used in shrimp farming, especially for kuruma prawn and tiger prawn, which has in high demand worldwide [7]. Fish, like other animals, cannot synthesize their own coloring pigments de novo, and must obtain these pigments from their diet. Pigmentation of cultured salmonids has been achieved with the inclusion of various synthetic carotenoids (\beta-carotene, canthaxanthin, zeaxanthin, and astaxanthin) and/or natural sources (yeast, bacteria, algae, higher plants, and crustacean meal) [8-9] in their diets. Global population increase is associated with increased demand of food. To overcome this challenge, the World Health Organization has suggested a doubling of food production by 2050. The range of non-conventional biotechnological measures involving improvement in CO₂ fixation efficiency of crop plants can be used to enhance the food productivity per hectare of agricultural land [10-11]. Several factors such as nutrient

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¹⁻³ Algal Biotechnology Laboratory, Department of Post Graduate Studies and Research in Biological Sciences, Rani Durgavati University, Jabalpur - 482 001, Madhya Pradesh, India mining, water limitation, accumulation of noxious xenobiotic compounds in the soil, soil corrosion and climate change have deteriorated the quality and fertility of agricultural land [12]. In our previous study [13-14] *Haematococcus pluvialis* was cultivated under laboratory conditions and astaxanthin was extracted from the green alga *Haematococcus pluvialis*. In present investigation, efforts were made to study the application part of crude astaxanthin.

MATERIALS AND METHODS

Cultivation of Haematococcus pluvialis and Nostochopsis

Haematococcus pluvialis and Nostochopsis were grown in the bold basal medium (BBM) [15]. For the preparation of the inoculum, the cells from the stock culture were centrifuged at 5000 rpm for 5 minutes the supernatant was discarded and the pellet was washed with the sterilized double distilled water thrice. The pellet was homogenized in 1 ml BBM and transferred aseptically in a 250 ml conical flask containing 100 ml of fresh BBM (KH₂PO₄, 17.5; CaCl₂·2H₂O, 2.5; MgSO₄·7H₂0, 7.5; NaNO₃, 25; K₂HPO₄, 7.5; NaCl, 2.5; Na₂ETDA, 10; FeSO₄·7H₂O, 4.98; H₃BO₃, 11.5 g/L, pH 6.8) and incubated under continuous illumination of 35 µmol m⁻² s⁻ at 25±2°C for 4 days. A 4-day old culture was used as an inoculum for the experiment. The experiment was performed in 250 ml conical flasks. 4-day old culture approximately 1×10^6 cells mL⁻¹ was inoculated into 100 ml sterilized fresh medium in 250 ml flasks and incubated in controlled air-



conditioned culture room at 25 ± 2 °C. All the cultures were shaken thrice a day with a rotary flask shaker.

Harvest and drying of Haematococcus pluvialis and Nostochopsis

The *H. pluvialis* and *Nostochopsis* biomass were centrifuged at 5000 rpm for 5 minutes, the supernatant was discarded and the pellet was dried in the oven at 70° C [16] and packed for the further analysis.

Chemicals

All the media chemicals used for the experiments were analytical grade, obtained from companies- Sisco Research Laboratories Pvt. Ltd., Mumbai; HiMedia Laboratories Pvt. Ltd., Mumbai; Loba Chemie Pvt. Ltd., Mumbai. Qualigens Fine Chemicals, Mumbai and Sigma-Aldrich Chemicals, USA. The feed was obtained from the golden feed, New Delhi.

Experimental design for the fish feed treatments and proximate analysis

All experiments were approved by the institutional animal ethics committee (IAEC). Catla catla fish weighing 40 g were used for the study obtained from a local hatchery Jabalpur. The fish were acclimated to the laboratory conditions for 2 weeks by feeding commercially available normal fish feed (Table 1) After this conditioning period, fish (average body weight, 40g) were randomly distributed into 2 rectangular of 20 L capacity at a density of 6 fish per tank. Tank 1 fish were given a normal diet, tank 2 were given H. pluvialis biomass and Nostochopsis (100 mg/kg b.w) (Table 2). Each experimental diet was fed to 3 replicate groups of fish twice per day for 4 weeks. Freshwater was supplied at a flow rate of 1 L/min in the re-circulating system and aeration was continuously provided in each tank. The photoperiod was left at the natural condition, and the average water temperature during the trial was 30±2°C. After a 24 h gut evacuation period all fish were anaesthetized and weighed. Out of six, three samples were randomly collected and stored at -20°C, until proximate analysis and lipid extraction [17].

Table 1 Composition of normal feed for the treatments of Catla catla

Calla Calla					
S. No.	Ingredients				
1	Soybean				
2	Rice Bran				
3	Wheat bran				
4	Wheat flour				
5	Vitamin- Mineral mix*				

*Vitamin D Complex, Vitamin E, Vitamin K, Vitamin B Complex, Mn, Fe, Mg, Zn, Cu, Co, Ca

Table 2 Formulation of <i>H. pluvialis</i> and <i>Nostochopsis</i>					
	based feed for the treatments of Catla catla				
C M	Traatmanta				

5 . NO.	Treatments		
1	Normal fish feed		
2	H. pluvialis and Nostochopsis biomass		

RESULTS AND DISCUSSION

Haematococcus pluvialis and *Nostochopsis* were grown in the bold basal medium for 30 days in incubated in controlled air-conditioned culture room (Fig 1).



Fig 1 Cultivation of *H. pluvialis and Nostochopsis* in controlled airconditioned culture room

Drying of H. pluvialis and Nostochopsis

After the harvest, *H. pluvialis* and *Nostochopsis* biomass was dried in hot air oven for 8 h and packed in polybags.

Effect of Haematococcus pluvialis and Nostochopsis biomass on the proximate analysis of Catla catla fish

The parameters determined for proximate analysis include proteins, lipid and ash content. In normal feed treated *Catla catla* fish group, lipid content was found as 82.2 ± 1.3 . In *Haematococcus pluvialis* and *Nostochopsis* biomass-based feed group, lipid content was observed as 177.6 ± 2.1 . However, protein and ash content were reported as 133.6 ± 1.2 , 98.6 ± 1.4 and 14.4 ± 2.1 , 7.2 ± 2.2 in *Haematococcus pluvialis* biomass-based group and normal feed group respectively (Table 3).

Table 3 Effect of different feed treatments	on the proximate	analysis of the wh	nole body of <i>Catla ca</i>	<i>atla</i> fish (mg/kg)
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S. No.	Treatments	Lipid	Protein	Ash
2	H. pluvialis and Nostochopsis feed	177.6±2.1	133.6±1.2	14.4 ± 2.1
3	Normal feed	82.2±1.3	98.6±1.4	7.2 ± 2.2

In this study, we evaluated that the inclusion of *H. pluvialis* and *Nostochopsis* biomass as a feed for the *Catla catla* at levels of 100mg/kg caused a significant effect on proximate analysis. According to previous studies testing the inclusion of 5% of *Ulva* spp. meal in feed for carnivorous fish like European sea bass (*Dicentrarchus labrax*) [18] and

rainbow trout [19) reported no effect on growth however some algal feed on *Nile tilapia* [20-21), common carp (*Cyprinus carpio*) [22) gilthead sea bream (*Sparus aurata*) [23] reported significant improvement in growth performances, feed efficiency, nutrient utilization and body composition. In our present study consumption of *H. pluvialis* and *Nostochopsis*



increased the lipid as well as protein content of the *Catla catla*. For the point of view of consumers *Catla catla* showed an umpteen attractive pigmentation (data not mentioned). A critical shortcoming of the crop plant proteins commonly used in fish feeds is that they are deficient in certain amino acids such as lysine, methionine, threonine, and tryptophan [24], whereas analyses of the amino acid content of numerous algae have found that although there is significant variation, they generally contain all the essential amino acids. For example, surveys of 19 tropical seaweeds [25] Analyses of microalgae have found similar high contents of essential amino acids, as exemplified by a comprehensive study of 40 species of

microalgae from seven algal classes that found that, all species had a similar amino acid composition, and were rich in the essential amino acids [26].

CONCLUSION

The present study reveals that *H. pluvialis* and *Nostochopsis* biomass can be used for the different aquatic feed formulations.

Conflict of interest

We declare that we have no conflict of interest.

LITERATURE CITED

- 1. Higuera-Ciapara I, Felix-Valenzuela L, Goycoolea FM. 2006. Astaxanthin: A review of its chemistry and applications. *Critical Reviews in Food Science and Nutrition* 46: 185-196.
- 2. Miki W. 1991. Biological functions and activities of animal carotenoids. Pure and Applied Chemistry 63(1): 141-146.
- 3. Borowitzka MA, Huisman JM, Osbo A. 1991. Culture of the astaxanthin-producing green alga *Haematococcus pluvialis*. Effects of nutrients on growth and cell type. *Journal of Applied Phycology* 3: 295-304.
- 4. Boussiba S, Vonshak A. 1991. Astaxanthin accumulation in the green alga H. pluvialis. Plant and Cell Physiology 32: 1077-87.
- 5. Kobayashi M, Kakizono T, Nagai S. 1991. Astaxanthin production by a green algal, *Haematococcus pluvialis* accompanied with morphological changes in acetate media. *Journal of Fermentation Bioengineering* 71(5): 335-339.
- 6. Lee YK, Soh CW. 1991. Accumulation of astaxanthin in H. lacustris (Chlorophyta). Journal of Phycology 27: 575-577.
- 7. Lorenz RT, Cysewski GR. 2000. Commercial potential for *Haematococcus* microalgae as a natural source of astaxanthin. *Trends in Biotechnology* 18: 160-167. doi:10.1016/S0167-7799(00)01433-5.
- 8. Shahidi F, Brown JA. 1998. Carotenoid pigments in seafoods and aquaculture. Critical Rev. Food Sci. and Nutrition 38: 1-67.
- 9. Kalinowski CT, Robaina LE, Fernández-Palacios H, Schuchardt D, Zquierdo MS. 2005. Effect of different carotenoid sources and their dietary levels on red porgy (*Pagrus pagrus*) growth and skin colour. *Aquaculture* 244: 223-231. doi: 10.1016/j.
- Parry MAJ, Hawkesford MJ. 2010. Food security: increasing yield and improving resource use efficiency. *Proc. Nutr. Soc.* 69: 592-600. doi: 10.1017/S0029665110003836
- 11. Parry MAJ, Reynolds M, Salvucci ME, Raines C, Andralojc PJ, Zhu XG. 2011. Raising yield potential of wheat. II. Increasing photosynthetic capacity and efficiency. *Jr. Exp. Botany* 62: 453-467. doi: 10.1093/jxb/erq304
- 12. Singh JS. 2014. Cyanobacteria: a vital bio-agent in eco-restoration of degraded lands and sustainable agriculture. *Climate Change Environ. Sustain.* 2: 133-137.
- 13. Rather AH, Singh S. 2018. Preliminary evaluation of impact of monochromatic light on the biosynthesis of astaxanthin in green alga *Haematococcus pluvialis*. World News of Natural Science 19: 45-50.
- 14. Singh S, Rather AH. 2018. Impact of light and dark (L/D) period on the biosynthesis of astaxanthin in green alga *Haematococcus pluvialis. Journal of Applied Biology and Biotechnology* 6(6): 58-60.
- 15. Kanz T, Bold HC. 1969. In: Physiological Studies. 9. Morphological and Taxonomic Investigations of Nostoc and Anabaena in Culture. Austin Texas, University of Texas, Publication. pp 6924.
- 16. Sarada R, Vidhyavathi R, Usha D, Ravishankar GA. 2006. An efficient method for extraction of astaxanthin from green alga *Haematococcus pluvialis. Journal of Agriculture and Food Chemistry* 4: 7585-7588.
- 17. Folch JM, Lees M, Sloane Stanley GH. 1957. A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry* 226: 497-509.
- 18. Diler I, Tekinay AA, Guroy D, Kut Guroy B, Soyuturk M. 2007. Effects of *Ulva rigida* on the growth, feed intake and body composition of common carp, *Cyprinus carpio* L. *Journal of Biological Science* 7(2): 305-308.
- Valente LMP, Gouveia A, Rema P, Matos J, Gomes EF, Pinto IS. 2006. Evaluation of three seaweeds Gracilaria bursapastoris, Ulva rigida and Gracilaria cornea as dietary ingredients in European sea bass (Dicentrarchus labrax) juveniles. Aquaculture 252(1): 85-91.
- 20. Wassef E, El-sayed AM, Kandee KM. 2005. Evaluation of *pterocladia (Rhodophyta)* and *Ulva (Chlorophyta)* meals as additives to gilthead seabream *Sparus aurata* diets. *Egypt Journal of Aquatic Research* 31: 321-332.
- 21. Güroy D, Güroy B, Merrifield DL, Ergün S, Tekinay AA, Yiğit M. 2011. Effect of dietary Ulva and Spirulina on weight loss and body composition of rainbow trout, Oncorhynchus mykiss (Walbaum), during a starvation period. Journal of Animal Physiology and Animal Nutrition 95(3): 320-7. 10.1111/j.1439-0396.2010.01057.x.
- 22. Ergün S, Soyutürk M, Güroy B, Güroy D, Merrifield D. 2009. Influence of Ulva meal on growth, feed utilization, and body composition of juvenile Nile tilapia (*O. niloticus*) at two levels of dietary lipid. *Aquaculture International* 17(4): 355-361.
- 23. Azaza MS, Mensi F, Ksouri J, Dhraief MN, Brini B, Abdelmouleh A. 2008. Growth of Nile tilapia (*Oreochromis niloticus* L.) fed with diets containing graded levels of green algae ulva meal (*Ulva rigida*) reared in geothermal waters of southern Tunisia. *Journal of Applied Ichthy* 24(2): 202-207.
- 24. Li P, Mai K, Trushenski J, Wu G. 2009. New developments in fish amino acid nutrition: towards functional and environmentally oriented aquafeeds. *Amino Acids* 37: 43-53.
- 25. Lourenço SO, Barbarino E, De-Paula JC, da S, Pereira LO, Lanfer Marquez UM. 2002. Amino acid composition, protein content and calculation of nitrogen-to-protein conversion factors for 19 tropical seaweeds. *Phyl. Research* 50: 233-241.
- 26. Brown MR, Jeffrey SW, Dunstan GA. 1997. Nutritional properties of microalgae for mariculture. Aquaculture 151: 315-331.