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B. Meena, K. Durgadevi and V. Ramamurthy

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Studies on Phytochemical and Antibacterial Potential of *Cymodocea serrulata*

B. Meena¹, K. Durgadevi² and V. Ramamurthy^{*3}

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ABSTRACT

The present study was designed to evaluate the antibacterial and anti-inflammatory potential of three different extract of *Cymodocea serrulata*. The extract of the seagrass was tested against *Streptococcus bovis*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Chlamydia pneumoniae* and *Helicobacter pylori* by agar method. The anti-inflammatory activity of *C. serrulata* was done by protein denaturation method. Phytochemical screening revealed the presence of carbohydrates, reducing sugars, alkaloids, saponins, phenolic compounds and flavonoids in aqueous seagrass extract. The results of the present study conclude that the studied plant possesses broad-spectrum antibacterial and antioxidant properties and may act as a potent antioxidant for biological systems susceptible to free radical-mediated reactions.

Key words: *Cymodocea serrulata*, Seagrass extract, Anti-inflammatory, Antibacterial activity, Phytochemical activity

Natural products have been an important resource for the maintenance of life for ages. Several life-saving drugs have been developed from the plants. The plant kingdom has provided an endless source of medicinal plants first used in their crude forms as herbal teas, syrups, infusions, ointments, liniments and powders. Herbal remedies and alternative medicines are used throughout the world and in the past herbs often represented the original sources of most drugs. Marine species are known to produce a large number of structurally diverse secondary metabolites [1]. Seagrasses, a group of marine flowering plants, inhabit the tidal and sub-tidal zones of shallow and sheltered localities of seas, gulfs, bays, backwaters, lagoons, and estuaries along temperate and tropical coastlines of the world [2-3]. With only about 72 species and 13 genera, seagrasses play key ecological roles in fisheries production, sediment accumulation, and stabilization [4] and have direct value to humanity as food, feed, green manure, and medicine [5-6]. Phytochemical analyses of seagrass species have shown that they are potential sources of antioxidants [7-8], antibacterial, antifungal and anti-inflammatory agents [9-10], and source of anticancer compounds [11]. The present study the anti-inflammatory of important seagrasses (*Cymodocea serrulata*) of Thanjavur, Tamil Nadu, India, along with an estimation of phytochemical and antimicrobial activity.

MATERIALS AND METHODS

Sample collection

Algal samples will be collected from Thanjavur district, East coastal region, Tamil Nadu. The wet algal species were identified by standard according to their morphologies [12-13]. Wet algal species will be first washed with sea water to remove the debris like sand, sea shells, pieces of wood and tiny stones. It will be shade dried for 24 hours and then finally dried in a tray drier at 60°C to remove the water content. Dry algae obtained will be finely chopped into pieces and then ground into fine powder using mortar and pestle. Microwave drying makes the drying process faster without any degradation of cell components.

Preparation of extract

For extraction, different solvents such as methanol, ethanol and chloroform were added to 100 g of powdered leaves separately and placed in Soxhlet apparatus for 24 h. The extracts were filtered with Whatman 40 filter paper and then concentrated using a rotary evaporator to give rise to a semi-solid mass. Each solvent extraction method was repeated thrice for the purpose of accuracy. The residues obtained were stored in refrigerator for further analysis.

Phytochemical screening

Qualitative phytochemical screenings were performed using standard procedures [14-15]. The occurrence of phytochemicals in the crude extracts of *Cymodocea serrulata* was determined.

Screening of antibacterial activity

* V. Ramamurthy

✉ v.ramamoorthy07@gmail.com

¹⁻³ P. G. & Research Department of Biochemistry, Marudupandiyar College, Thanjavur - 613 403, Affiliated to Bharathidasan University, Tiruchirappalli, Tamil Nadu, India

Antibacterial activity of the *Cymodocea serrulata* was tested using the agar diffusion method described by Collin and Lyne [16]. The extracts were tested for the antibacterial activity against the six bacterial species such as *Streptococcus bovis*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Chlamydia pneumoniae* and *Helicobacter pylori*. *Cymodocea serrulata* were prepared and tested using Nutrient agar medium. The plates were incubated at 37°C for 24 hours and the zones of inhibition measured.

Anti-inflammatory activity

Anti-inflammatory activity of the seagrass extract was determined by *in vitro* method such as protein denaturation [17-18].

RESULTS AND DISCUSSION

Preliminary phytochemical screening

The phytochemicals were analyzed qualitatively by using standard protocols in different solvent extract of sea grass.

The protein, reducing sugar, phenol, tannins, amino acid and steroids were found in all the extracts. The flavonoids, anthraquinones and terpenoids were present in methanol and chloroform extracts. Tanins, alkaloids, amino acids, steroids and phenol were present in the ethanol extract of *C. serrulata*. The saponins, resins and glycosides were present only in the methanol extracts of sea grass *C. serrulata*. This is consistent with the findings of Ragupathi *et al.* [19] who had reported the qualitative analysis of the above phytoconstituents in the methanolic extracts of five seagrasses like *Enhalus acoroides*, *Thalassia hemprichii*, *Halodule pinifolia*, *Cymodocea serrulata* and *Cymodocea rotundata* from Chinnappallam coast of Tamil Nadu. Athiperumalsami *et al.* [20] screened four seagrasses such as *Halophila ovalis*, *S. isoetifolium*, *C. serrulata* and *H. pinifolia* and reported 15 phytochemicals from benzene and petroleum ether extract of *S. isoetifolium* collected from Gulf of Mannar. The results of the present study is also in line with the results of Girija *et al.* [21] who reported the presence of ten phytoconstituents in the methanol extracts of *C. serrulata* collected from the study site.

Table 1 Qualitative phytochemical analysis for the extracts of *Cymodocea serrulata*

Phytochemicals	Solvents		
	Methanol	Ethanol	Chloroform
Proteins	+	+	+
Resins	+	-	-
Tannins	+	+	+
Saponins	+	+	+
Flavonoids	+	+	+
Alkaloids	+	+	+
Amino acids	+	+	+
Steroids	+	+	+
Reducing sugar	+	+	+
Glycosides	+	+	+
Anthraquinones	+	-	+
Terpenoids	+	+	+
Phenol	+	+	+

Antibacterial analysis

The zone of inhibition measured for bacteria using well diffusion method. The antibacterial analysis (Table 2) showed a remarkable activity against the bacterial pathogens with different extract of *Cymodocea serrulata*. The maximum activity compared to the control shows the potential of the seagrass and is an indicator for determining the significance of the activity against the pathogens. The overall antibacterial analysis reveals maximum against the *Helicobacter pylori* and minimum activity was noted against the *Escherichia coli*. Overall observation reveals that the seagrass has inhibitory

activity against all the pathogens studied. *Cymodocea serrulata* is a potential source of broad-spectrum antibacterial agents due to the presence of flavonoids, which have been reported to be involved in inhibition of nucleic acid biosynthesis and other metabolic processes [22]. Some of the seagrasses have been used in traditional medicine for example in India for malaria, skin diseases and the early stage of leprosy [23]. Some extracts also have antibacterial activity [24-26]. During the long period of co-evolution, a cooperative relationship has been formed between each endophyte and its host plant. Some endophytes have the ability to produce similar bioactive compounds to those that originate from their terrestrial host plants [27].

Table 2 Antibacterial activity of *C. serrulata* extract against pathogens

Pathogens	Crude extracts (Zone of inhibition-mm)			Standard
	Methanol	Chloroform	Ethanol	
<i>Staphylococcus aureus</i>	17.5 ± 0.22	11.2 ± 0.18	13.5 ± 0.21	17.9 ± 0.62
<i>Streptococcus bovis</i>	15.7 ± 0.38	10.6 ± 0.21	12.2 ± 0.14	16.2 ± 0.18
<i>Escherichia coli</i>	12.9 ± 0.12	09.2 ± 0.12	10.6 ± 0.25	13.8 ± 0.25
<i>Salmonella typhi</i>	17.1 ± 0.32	11.5 ± 0.17	13.9 ± 0.32	15.2 ± 0.15
<i>Chlamydia pneumoniae</i>	15.6 ± 0.25	10.3 ± 0.24	12.1 ± 0.13	16.8 ± 0.22
<i>Helicobacter pylori</i>	19.5 ± 0.18	14.4 ± 0.15	16.9 ± 0.25	21.7 ± 0.29

Each value is the Mean ± SD of three replicates

Anti-inflammatory activity

The percentage of protein denaturation for extract and standard drug diclofenac was done at 100µg/ml, 200µg/ml, 300µg/ml, 400µg/ml and 500µg/ml respectively as given

in (Table 3). The *C. serrulata* exhibits minimum stabilization 21.8% at 100µg/ml and maximum stabilization 35.3% at 500µg/ml which was presented in (Table 3). Percent inhibition of protein denaturation activity was exhibited on the basis of

concentration dependent manner. Sodium diclofenac was used as a standard. During inflammation the lysosomal enzymes is released which produced a variety of disorders and these enzymes is said to be related to acute or chronic inflammation.

The diclofenac drugs act either by inhibiting these lysosomal enzymes or by stabilizing the lysosomal membrane. The seagrass extracts of *C. serrulata* showed biphasic effects on protein denaturation method.

Table 3 Anti-inflammatory activity of *C. serrulata*

Concentration (µg/ml)	Percent inhibition of protein denaturation		
	Methanol	Ethanol	Chloroform
100	20.5 ± 0.03	19.2 ± 0.14	18.3 ± 0.16
200	22.4 ± 0.15	21.7 ± 0.23	20.5 ± 0.32
300	26.1 ± 0.21	24.3 ± 0.12	23.1 ± 0.18
400	30.5 ± 0.18	28.6 ± 0.26	27.2 ± 0.25
500	34.8 ± 0.39	32.4 ± 0.18	30.9 ± 0.12
Standard	74.2 ± 0.16	70.8 ± 0.25	68.5 ± 0.22

Each value is the Mean ± SD of three replicates

In the present study, the results show that methanolic extracts of *C. serrulata* have well anti-inflammatory properties *in vitro* in several models such as the inhibition of denaturation of proteins, 5-LOX, COX and ROS. Inflammation, which is a very complex physiopathological response, involves the production of free radicals derived from neutrophils, NO, ROS, cytokines, and prostaglandins during its process [28]. Protein denaturation is the process by which proteins lose their tertiary structure and secondary structure. Proteins denaturation is a well-documented cause of inflammation [29]. In various inflammatory and allergic disorders, COX and 5-LOX are the main enzymes in the synthesis of prostanoids and eicosanoids from polyunsaturated fatty acids. The effective reduction of chronic inflammatory conditions is important by double inhibition of LOX and COX [30]. Substances capable of producing double inhibition of COX and 5-LOX with consequent substantial reduction in leukotriene and prostaglandin production produce a broad spectrum of anti-inflammatory activity and can be considered to have an excellent profile of pharmacological safety in clinical practice [31]. The *C. serrulata* exhibits minimum stabilization 21.8% at 100µg/ml and maximum stabilization 35.3% at 500µg/ml which

was presented in (Table 3). Percent inhibition of protein denaturation activity was exhibited on the basis of concentration dependent manner. Sodium diclofenac was used as a standard. During inflammation the lysosomal enzymes is released which produced a variety of disorders and these enzymes is said to be related to acute or chronic inflammation. The diclofenac drugs act either by inhibiting these lysosomal enzymes or by stabilizing the lysosomal membrane. These results are justified by the fact that compounds such as betulinic acid isolated from *D. thollonii* possess an *in vitro* inhibition property of cyclooxygenase (COX-1 and COX-2) and leukotriene B4 formation mediated by 5-LOX [32].

CONCLUSION

On the basis of the results obtained in the present study, it is concluded that methanol extract of *C. serrulata* has potent anti-inflammatory and antibacterial activities. Thus the *C. serrulata* extract may be attributed to the presence of phenolic compounds and flavonoids etc., therefore, further investigation is needed to isolate and identify the active compounds present in the plant extract and its efficacy.

LITERATURE CITED

1. Sundaram R, Nanthini Devi K, Ajith Kumar TT, Ajmalkhan M. 2011. Antibacterial activity of seagrass species of *cymodocea serrulata* against chosen bacterial fish pathogens. *Annals of Biological Research* 2(1): 88-93.
2. Green EP, Short FT. 2003. *World Atlas of Seagrasses*. Berkeley: University California Press.
3. Short FT, Coles RG, Pergent-Martini C. 2001. Global seagrass distribution. In: Short FT, Coles RG, editors. *Global Seagrass Research Methods*. Amsterdam: Elsevier Science BV. pp 5-30.
4. Ronnback P, Kautsky N, Pihl L, Troell M, Söderqvist T, Wennhage H. 2007. Ecosystem goods and services from Swedish coastal habitats: Identification, valuation, and implications of ecosystem shifts. *Ambio*. 36: 534-544.
5. Newmaster AF, Berg KJ, Ragupathy S, Palanisamy M, Sambandan K, Newmaster SG. 2011. Local knowledge and conservation of seagrasses in the Tamil Nadu state of India. *Jr. Ethnobiol. Ethnomed*. 7: 37.
5. Ross C, Puglisi MP, Paul VJ. 2008. Antifungal defenses of seagrasses from the Indian River Lagoon, Florida. *Aqua Botany* 88: 134-141.
6. Ragupathi KR, Radjessegarin A, Perumal A. 2013. Seagrasses as potential source of medicinal food ingredients: Nutritional analysis and multivariate approach. *Biomed. Prevent. Nutrition* 3: 375-80.
7. Ragupathi KR, Radjessegarin A, Meenakshi S, Perumal A. 2010. Thin layer chromatography analysis of antioxidant constituents of seagrasses of gulf of mannar biosphere reserve, South India. *Int. Jr. Chem. Tech. Research* 2: 1526-1530.
8. Rengasamy RR, Rajasekaran A, Micheline GD, Perumal A. 2011. Antioxidant activity of seagrasses of the Mandapam coast, India. *Pharm Biol*. 50: 182-187.
9. Puglisi MP, Engel S, Jensen PR, Fenical W. 2007. Antimicrobial activities of extracts from Indo-Pacific marine plants against marine pathogens and saprophytes. *Mar Biol*. 150: 531-540.
10. Yuvaraj N, Kanmani P, Satishkumar R, Paari A, Pattukumar V, Arul V. 2012. Seagrass as a potential source of natural antioxidant and anti-inflammatory agents. *Pharm Biol*. 50: 458-467.
11. Folmer F, Jaspars M, Dicato M, Diederich M. 2000. Photosynthetic marine organisms as a source of anticancer compounds. *Phytochem. Rev*. 9: 557-579.
12. Menez EG, Phillips RC, Calumpong HP. 1983. Seagrasses from the Philippines. *Smithsonian Contrib. Mar Sci*. 21: 1-39.
13. Coles R, McKenzie L, Campbell S, Mellors J, Waycott M. 2004. Queensland, Australia: Brochure of CRC Reef Research Centre.

14. Sofowora A. 1993. *Phytochemical Screening of Medicinal Plants and Traditional Medicine in Africa*. Spectrum Books Ltd, Ibadan, Nigeria. 2, 320.
15. Trease GE, Evans MD. 1989. *A Text Book of Pharmacognosy*. 13th Edition Baillier, Tindal and Caussel, London. pp 144-148.
16. Collins CH, Lyne PM. 1970. *Microbiological Methods*. 3rd Edition. Butterworth and Co. Ltd. pp 414-427.
17. Mizushima Y, Kobayashi M. 1968. Interaction of anti-inflammatory drugs with serum preteins, especially with some biologically active proteins. *Jr. Pharma. Pharmacology* 20: 169-173.
18. Elias G, Rao MNA. 1988. Synthesis and anti-inflammatory activity of substituted (E)-4-phenyl-3-buten-2-ones. *Eur. Jr. Med. Chemistry* 23: 379-380.
19. Ragupathi RKR, Arumugam R, Anantharaman P. 2013. Phytochemical constituents, antioxidant properties and p-coumaric acid analysis in some seagrasses. *Food Res. Inter.* 12: 25-28.
20. Athiperumalsami T, Kumar V, Louis JL. 2008. Survey and phytochemical analysis of seagrasses in the Gulf of Mannar, southeast coast of India. *Bot. Mar.* 51: 269-277.
21. Girija K, Parthiban C, Hemalatha A, Saranya C, Anantharaman P. 2013. Evaluation of antioxidant activities and preliminary phytochemical analysis of seagrass *Halodule pinifolia*, *Halophila ovalis* and *Syringodium isoetifolium*. *The Jr. Phytochem. Photon.* 114: 181-187.
22. Cushnie TPT, Lamb AJ. 2005. Detection of galangin-induced cytoplasmic membrane damage in *Staphylococcus aureus* by measuring potassium loss. *Jr. Ethnopharmacology* 101: 243-248.
23. Kumar CS, Sarada DVL, Gideon TP, Rengasamy R. 2008. Antibacterial activity of three South Indian seagrasses, *Cymodocea serrulata*, *Halophila ovalis* and *Zostera capensis*. *World Jr. Microbiol. Biotechnology* 24: 1989-1992.
26. Engel S, Puglisi MP, Jensen PR, Fenical W. 2006. Antimicrobial activities of extracts from tropical Atlantic marine plants against marine pathogens and saprophytes. *Mar. Biology* 149: 991-1002.
26. Ravikumar S, Alil MS, Ajmalkhan AM, Dhinakaraj M. 2011. Antibacterial activity of *Cymodocea serrulata* root extract against chosen poultry pathogens. *Indian Jr. Sci. Technology* 4: 98-100.
27. Zhao K, Penttinen P, Guan T, Xiao J, Chen Q. 2011. The diversity and anti-microbial activity of endophytic actinomycetes isolated from medicinal plants in Panxi Plateau China. *Curr. Microbiology* 62: 182-190.
28. Mannan MM, Maridass M, Victor B. 2008. A review on the potential uses of ferns. *Ethnobotanical Leaflets* 12(24): 281-285.
29. Anoop MV, Bindu AR. 2015. *In-vitro* anti-inflammatory activity studies on *Syzygium zeylanicum* (L) DC leaves. *Inter. Jr. Pharmacol. Res. and Review* 4(8): 18-27.
30. De Gaetano G, Donati MB, Cerletti C. 2003. Prevention of thrombosis and vascular inflammation: benefits and limitations of selective or combined COX-1, COX-2 and 5-LOX inhibitors. *Tren. Pharmacol. Sci.* 24(5): 245-252.
31. Frey FM, Meyers R. 2010. Antibacterial activity of traditional medicinal plants used by Haudenosaunee peoples of New York State. *BMC Complem. Alter. Med.* 10(1): 64.
32. Wenzig EM, Widowitz U, Kunert O. 2008. Phytochemical composition and *in vitro* pharmacological activity of two rose hip (*Rosa canina* L.) preparations. *Phytomedicine* 15(10): 826-835.