



*Prevalence of Aflatoxin in Maize Grains of  
Madhubani District - North Bihar*

Diksha Sultania, Gajendra Prasad and Lucky Rani

Research Journal of Agricultural Sciences  
An International Journal

P- ISSN: 0976-1675

E- ISSN: 2249-4538

Volume: 13

Issue: 04

*Res. Jr. of Agril. Sci.* (2022) 13: 1311–1313



C A R A S



# Prevalence of Aflatoxin in Maize Grains of Madhubani District - North Bihar

Diksha Sultania\*<sup>1</sup>, Gajendra Prasad<sup>2</sup> and Lucky Rani<sup>3</sup>

Received: 19 Jun 2022 | Revised accepted: 09 Aug 2022 | Published online: 20 Aug 2022  
© CARAS (Centre for Advanced Research in Agricultural Sciences) 2022

## ABSTRACT

Mycotoxin contamination is a serious problem of cereal crops in tropical regions like India. Aflatoxin (AF) is one of the most serious mycotoxin contaminants as aflatoxin B<sub>1</sub> is classified as group I human carcinogen. It has mutagenic, tetraoxygenic and carcinogenic effects. Maize samples from three villages near Kosi river, of Madhubani district were collected and examined for associated mycoflora and aflatoxin producing potential of *Aspergillus flavus*. It was found that *Aspergillus flavus* was of dominant occurrence followed by *Aspergillus niger* and species of *Fusarium*, *Penicillium* and *Rhizopus*. All fifteen samples, five of each village viz Ghoghardiha, Madhepur and Hatni were found contaminated with aflatoxin, samples collected from Madhepur and Hatni had higher value of AFB<sub>1</sub> contamination (1-250 µg/kg) than of Ghoghardiha village (1-80 µg/kg). 41% of the total *Aspergillus flavus* isolates were toxigenic, producing aflatoxin B<sub>1</sub>, B<sub>2</sub> and G<sub>1</sub>. The high level of aflatoxin is probably due to favorable environmental conditions, backward agricultural practices, poor storage conditions of grains and due to yearly flood problems in this area.

**Key words:** Maize seeds, Aflatoxin, Mycoflora, Madhubani district

Maize (*Zea mays* L) is one of the most important cereal crops from the point of agriculture and growing demand of food, as it has wide climatic tolerance, high genetic yield potential and rich nutrient content. It is boosted with antioxidants that help protect consumers from various degenerative diseases [1]. It contributes to 36% global food grain production and is a dominant cereal crop for both livestock feed and human consumption. However agronomic practices and climatic conditions favorable to mould growth severely affect the quality of maize seeds. Main toxigenic fungi associated with maize grains in tropical regions are species of *Aspergillus*, *Fusarium* and *Penicillium* [2-4]. Many mycotoxins are produced by these fungi of which aflatoxin is secondary metabolites produced mainly by *Aspergillus flavus* and *Aspergillus parasiticus* and is group I human carcinogen (ICRA 2002). *Aspergillus* species colonization and aflatoxin contamination is prevalent in tropical regions like India, as growth is favored by high temperature and humidity.

As Bihar is a poor state maize is a staple food and feed of this state. In Bihar, farmers mainly choose rabi (winter) season for maize cultivation because kharif and autumn crop suffer water logging and flood respectively. Madhubani district fall under north-west zone of Bihar and is a flood prone region.

Maize is harvested at 20-30% moisture content and lack of advance drying and storage facilities, due to socio-economic backwardness leads to further mould and toxin production [5]. Study which can put light on quality of maize produced in this region is lacking, thus an attempt is made to enumerate the mycoflora associated and aflatoxin produced by isolates of *Aspergillus flavus* in maize samples collected from farmers of Ghoghardiha, Hatni and Madhepur village of Madhubani district.

## MATERIALS AND METHODS

### Sample collection

Fifteen samples, of eight month old (from main harvest season) maize seeds were collected from Ghoghardiha, Hatni and Madhepur based farmers and kept in sterile bags for further use. Moisture content of each sample was recorded by help of OSAW moisture meter.

### Isolation and identification of Mycoflora

100 kernels of all three-sampling site were surface sterilized in 2% NaOCl and plated on moist blotting paper in sterile petri dishes (ISTA 1966). Enough space was kept between each kernel for proper growth of fungi and avoid cross contamination. Plates were incubated at room temperature for one week followed by macro and microscopic identification [6-8]. Fungal colonies were maintained on PDA media for further use and identification.

\* **Diksha Sultania**

✉ diksha4112@gmail.com

<sup>1-3</sup> Mycotoxin and Pathology Laboratory, University Department of Botany, Lalit Narayan Mithila University, Darbhanga - 846 008, Bihar, India

**Aflatoxin analysis of toxigenic isolates:** *Aspergillus flavus* isolates were allowed to grow on SMKY liquid media [9] for 7-8 days at 30±2°C and aflatoxin extracted by the process of Thomas *et al.* [10]. Then it was extracted with chloroform, dried on water bath and concentrated extracts were kept in screw tight bottles for qualitative and quantitative analysis.

Qualitative analysis of aflatoxin was done using thin layer chromatography (TLC), using toluene-isoamyl alcohol – methanol (90:32:2 v/v) solvent system [11]. For chemical confirmation trifluoroacetic acid [12] or 25% sulphuric acid spray was used.

Quantitative estimation of aflatoxin was done spectrophotometrically [13].

**Mycotoxin analysis of maize samples:** 100gm of maize sample was ground to fine powder and aflatoxin extraction was done by the process of Thomas *et al.* [10]. For qualitative and quantitative analysis same process was followed as that mentioned above for *Aspergillus flavus* isolates.

## RESULTS AND DISCUSSION

Each maize seed plated was carefully examined for fungal growth and associated mycoflora were noted which include *Aspergillus flavus*, *Aspergillus niger*, *Fusarium*

*moniliforme*, *Fusarium sp.*, *Penicillium sp.*, and *Rhizopus stolonifer* (Table 1).

The (Table 2) represents the data of toxigenic isolates of *Aspergillus flavus* obtained from SMKY liquid medium, type of aflatoxin produced and range of aflatoxin B<sub>1</sub> produced by these isolates. Total 197 isolates of *Aspergillus flavus* was screened for aflatoxin producing potential and 81 isolates were found positive. Out of 81 isolates, 57 produce AFB<sub>1</sub>, 18 produces both AFB<sub>1</sub> and AFB<sub>2</sub> and only 6 isolates produce AFB<sub>1</sub>, AFB<sub>2</sub>, and AFG<sub>1</sub> however no isolate was found to be positive for AFG<sub>2</sub>. In the isolates of sample collected from Ghoghardiha amount of aflatoxin B<sub>1</sub> ranged between 0.2-5ppm whereas samples of Madhepur and Hatni village had higher amount of aflatoxin B<sub>1</sub> contamination, in the range of 0.2-10ppm. A very few isolates (13%) produced AFB<sub>1</sub> above 5ppm and remaining isolates were low producers of AFB<sub>1</sub>.

Maize grains were analyzed for the presence of different types of aflatoxin and the quantity of aflatoxin B<sub>1</sub> present, (Table 3) shows that range of AFB<sub>1</sub> in samples collected from Ghoghardiha village is comparatively low (1-80µg/kg) from that collected from Madhepur and Hatni villages (1-250µg/kg). All samples were positive for AFB<sub>1</sub> and AFB<sub>2</sub> and AFG<sub>1</sub> was additionally present in samples of Madhepur and Hatni. Moisture content of samples collected from Ghoghardiha, Madhepur and Hatni villages are 8.8, 10.2, and 9.1 respectively.

Table 1 Mycoflora associated with maize seeds

Mycoflora	Ghoghardiha (N=100)		Madhepur (N=100)		Hatni (N=100)	
	Number of seeds infested	Moisture content	Number of Seeds infested	Moisture content	Number of seeds infested	Moisture content
<i>Aspergillus flavus</i>	60		73		64	
<i>Aspergillus niger</i>	5		8		2	
<i>Fusarium moniliforme</i>	1	8.8	0	10.2	2	9.1
<i>Fusarium sp</i>	4		5		1	
<i>Penicillium sp</i>	0		4		3	
<i>Rhizopus stolonifer</i>	18		14		23	

Table 2 Aflatoxin producing potential of toxigenic isolates of *Aspergillus flavus*

Place of collection	Total <i>Aspergillus flavus</i> isolates	Number of isolates producing aflatoxin	% positive isolates of <i>Aspergillus flavus</i>	Type of Aflatoxin			Range of AFB <sub>1</sub> (ppm)
				B <sub>1</sub>	B <sub>1</sub> B <sub>2</sub>	B <sub>1</sub> B <sub>2</sub> G <sub>1</sub>	
Ghoghardiha	60	26	43	18	8	-	0.2-5
Madhepur	73	32	44	25	4	3	0.2-10
Hatni	64	23	36	14	6	3	0.2-10
Total	197	81	41	57	18	06	

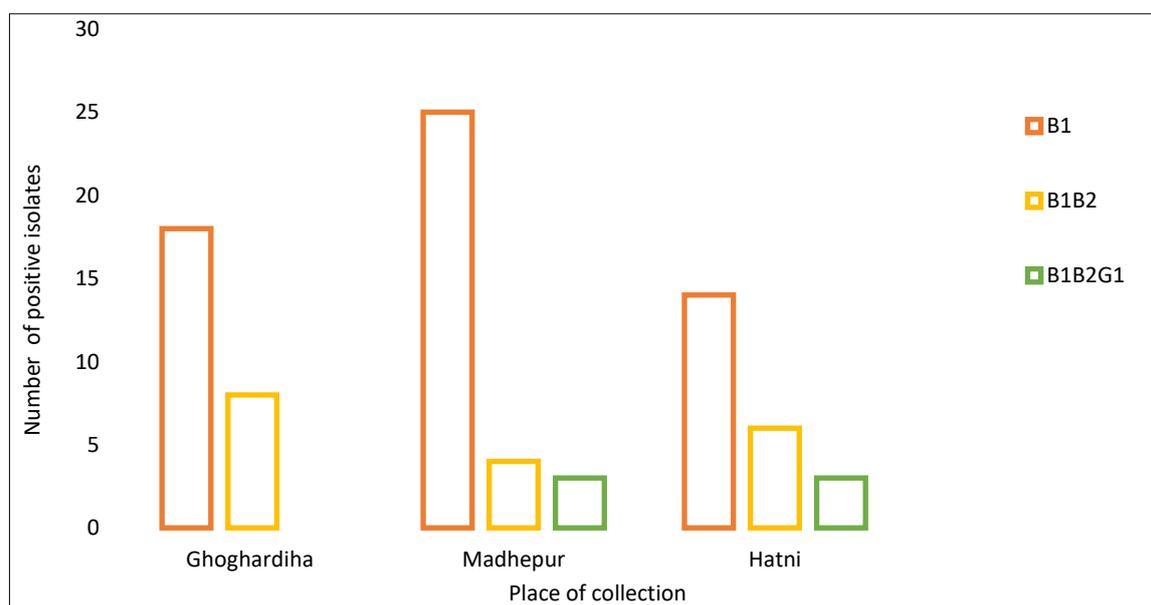


Fig 1 Extent of different aflatoxin produced by toxigenic isolates of *Aspergillus flavus*

Table 3 Aflatoxin contamination in maize grains

Place of collection	Moisture content	Sample positive for Aflatoxin			Range of AFB <sub>1</sub> (µg/kg)
		B <sub>1</sub>	B <sub>2</sub>	G <sub>1</sub>	
Ghoghardiha	8.8	+	+	-	1-80
Madhepur	10.2	+	+	+	1-250
Hatni	9.1	+	+	+	1-250

This study clearly indicates the association of mycotoxin producing fungi in maize seeds, viz., *Aspergillus*, *Penicillium* and *Fusarium*, the dominance of *Aspergillus* concurrence with earlier studies performed in the state of Bihar [14-15]. High aflatoxin level is probably due to favorable environmental conditions, poor storage, lack of use of natural and chemical control methods due to absence of knowledge and poverty.

Most of the maize crop produced by the farmers of this region is used either as food or feed requirement of their own or sold in local markets and dearth of knowledge and proper checks is somehow affecting the health of these people without being noticed, can lead to aflatoxicosis outbreaks, thus there is a need for better management by using techniques like IPM (Integrated Pest Management) which can eliminate or at least

bring down the level of *Aspergillus flavus* and consequently aflatoxin contamination in maize grains produced in Madhubani district of Bihar [16].

## CONCLUSION

All samples tested in this study were found dangerously contaminated with aflatoxin, but no awareness among farmers and common people about the side effects of consumption of such contaminated food is opening ways for contamination of entire food chain with aflatoxin. This brings the need for intense research and application of different control methods of *Aspergillus* contamination in modern agriculture.

## LITERATURE CITED

- Bathla S, Jaidka M, Kaur R. 2019. *Nutritive Value*. In: (Eds) Maize - Production and Use. IntechOpen. <https://doi.org/10.5772/intechopen.88963>
- Khosravi AR, Mansouri M, Bahonar AR, Shokri H. 2007. Mycoflora of maize harvested from Iran and imported maize. *Pakistan Journal of Biological Sciences* 10(24): 4432-4437.
- Chulze SN. 2010. Strategies to reduce mycotoxin levels in maize during storage: A review. *Food Additives and Contaminants* 27(5): 651-657.
- Dudoiu R, Cristea S, Lupu C, Popa D, Oprea M. 2016. Microflora associated with maize grains during storage period. *Agrolife Scientific Journal* 5(1): 63-58.
- Singh SB, Singh SP, Kasana RK. 2018. Status of corn cultivation in Bihar: Opportunities and future challenges. Chandra Shekhar Azad University of Agriculture & Technology, Kanpur (UP) India.
- McClenny N. 2005. Laboratory detection and identification of *Aspergillus* species by microscopic observation and culture: the traditional approach. *Medical Mycology* 43(Supplement-1): S125-S128.
- Gautam AK, Bhadauria R. 2012. Characterization of *Aspergillus* species associated with commercially stored triphala powder. *African Journal of Biotechnology* 11(104): 16814-16823.
- Adame-García J, Rodríguez-Guerra R, Iglesias-Andreu LG, Ramos-Prado JM, Luna-Rodríguez M. 2015. Molecular identification and pathogenic variation of *Fusarium* species isolated from *Vanilla planifolia* in Papantla Mexico. *Botanical Sciences* 93(3): 669-678.
- Diener UL, Davis ND. 1966. Aflatoxin production by isolates of *Aspergillus flavus*. *Phytopathology* 56(12): 1390-1393.
- Thomas F, Eppley RM, Trucksess MW. 1975. Rapid screening method for aflatoxins and zearalenone in corn. *Journal of the Association of Official Analytical Chemists* 58(1): 114-116.
- Reddy TV, Viswanathan L, Venkitasubramanian TA. 1970. Thin-layer chromatography of aflatoxins. *Analytical Biochemistry* 38(2): 568-571.
- Stack ME, Pohland AE. 1975. Collaborative study of a method for chemical confirmation of the identity of aflatoxin. *Journal of the Association of Official Analytical Chemists* 58(1): 110-113.
- Nabney J, Nesbitt BF. 1965. A spectrophotometric method for determining the aflatoxins. *Analyst* 90(1068): 155-160.
- Sinha KK. 1987. Aflatoxin contamination of maize in flooded areas of Bhagalpur, India. *Applied and Environmental Microbiology* 53(6): 1391-1393.
- Sinha KK. 1990. Incidence of mycotoxins in maize grains in Bihar State, India. *Food Additives and Contaminants* 7(1): 55-61.
- Rustom IY. 1997. Aflatoxin in food and feed: occurrence, legislation and inactivation by physical methods. *Food Chemistry* 59(1): 57-67.