

Evaluation of Phytochemicals and Assessment of Antibacterial Activity of *Cinnamomum zeylanicum*, *Syzygium aromaticum* and *Vetiveria zizanioides*

Rajendrabhai Daulatbhai Vasait*¹

¹ Department of Microbiology, Karmveer Abasaheb A. N. M. Sonawane Arts, Commerce and Science College, Satana - 423 301, Maharashtra, India

Abstract

Cinnamomum zeylanicum (Dalchini), *Syzygium aromaticum* (Clove), and *Vetiveria zizanioides* (Khus Khus) are commonly used as spices in the preparation of a variety of foods. The variety of spices includes clove, black pepper, ginger, cardamom, vetiver, cinnamon, turmeric, cumin, chili, and so on. Each of them has characteristic properties; some are added to enhance flavour, some are colouring agents, while others play the role of preservatives and also provide a distinctive aroma. In the phytochemical analysis of crude extracts, the presence of carbohydrates, amino acids, proteins, glycosides, tannins, terpenoids, phenols, alkaloids, and saponins was determined. Clinical isolates of *E. coli*, *Staphylococcus aureus*, *Streptococcus faecalis*, *Salmonella paratyphi B*, and *Pseudomonas aeruginosa* were used as test organisms for antibacterial testing, and all test organisms were found sensitive to crude extracts of the spices *Cinnamomum zeylanicum*, *Syzygium aromaticum*, and *Vetiveria zizanioides*. A methanolic extract of *Cinnamomum zeylanicum* was found to be moderately antibacterial; the higher diameter of the zone of inhibition measured was 29 mm against *E. coli*. The methanolic extract of *Syzygium aromaticum* exhibited a 26-mm-diameter inhibition zone against *E. coli*, whereas an ethanolic extract of *Vetiveria zizanioides* exhibited a 23-mm-diameter inhibition zone against *Salmonella paratyphi B*.

Key words: Phytochemicals, Antibacterial activity, *Cinnamomum zeylanicum*, *Syzygium aromaticum*, *Vetiveria zizanioides*

Spices in foods are used for their properties to augment aroma, flavour, and their varied roles [1-2]. Spices and phytochemicals of the varieties of plants have proven their health benefits as well as their wide prospect in medicine and are used against preventing and treating most infections and diseases [3-6]. Spices have long been used as traditional medicines [2], [7]. Spices are obtained from a variety of sources, including phytoplankton and their parts such as leaves, fruits, seeds, and the bark of trees, as well as metabolic products etc. [4]. Each spice's phytochemicals, or secondary compounds, have a particular flavour and aroma, and they aid plants by protecting them from pathogenic agents such as bacteria, moulds, viruses, pests, and parasites [8-9]. Piperine, Eugenol, Cinnamaldehyde, Lignans, Linalool, Thymoquinones, Curcumin, and Allicin are bioactive compounds obtained from spices and condiments such as Black Pepper, Cloves, and Cinnamon [10-13]. The principal components of these spices are tannins, terpenoids, alkaloids, and flavonoids, as well as lectins, saponins, phenolic compounds, and so on [14-15]. Most spices and their ingredients have important pharmacological

properties such as anti-growth, antiparasitic, analgesic, sedative, antiseptic, anti-diabetic, anti-malarial, blood sugar lowering, and cholesterol-lowering agents [9], [16-21]. Some of them are regarded as potent antioxidants and nurturers to the cells and tissues. The most important characteristic of spices, they do not contain any of those toxic substances for humans [11]. The present research investigation was conducted to evaluate the phytometabolites found in spices such as clove, dalchini, and khus khus and their bactericidal capabilities against clinical isolates for their unseen potential as antimicrobials.

MATERIALS AND METHODS

Collection of plant material

The stem bark of *Cinnamomum zeylanicum* (Dalchini), buds of *Syzygium aromaticum* (Clove), and seeds of *Vetiveria zizanioides* (Khus Khus) were purchased from local markets of Nashik district, Maharashtra, India, and brought to the laboratory. These were cleaned by washing them with distilled

Received: 14 Apr 2023; Revised accepted: 15 June 2023; Published online: 01 Sep 2023

Correspondence to: Rajendrabhai Daulatbhai Vasait, Department of Microbiology, Karmveer Abasaheb A. N. M. Sonawane Arts, Commerce and Science College, Satana - 423 301, Maharashtra, India, Tel: +91 9422780508; E-mail: rd73vas@rediffmail.com

Citation: Vasait RD. 2023. Evaluation of phytochemicals and assessment of antibacterial activity of *Cinnamomum zeylanicum*, *Syzygium aromaticum* and *Vetiveria zizanioides*. Res. Jr. Agril. Sci. 14(5): 1059-1064.

water three times to remove impurities, dust, and other particulate matter. The above substances were shade dried in the laboratory at room temperature for 5 days. The dried parts were ground to make a fine powder. Powdered samples were kept in airtight containers until the investigation.

Extraction of plants material

The plants material extracts were prepared as follows:

1. Preparation of aqueous extracts for volatile components

Dried clove buds were ground into a fine powder and dissolved in sterile distilled water (10% w/v) separately in an Erlenmeyer flask to make an aqueous extract. The flasks were incubated at room temperature on an orbital shaker for 48 hours for the extraction process. A widely accepted method, hydro distillation with some modifications [21], was used for the extraction of the bioactive profile from clove powder. The extract was then evaporated using a distillation unit with a round bottom flask at 40°C. The lower temperature is purposefully used for the probable retention of volatile bioactive compounds. The cooled vapours were condensed and collected in a closed container for the imprisonment of volatile components. After cooling, the final extract was stored in an airtight container and kept at 4°C. Similarly, the aqueous extracts for volatile components of *Cinnamomum zeylanicum* stem bark powder and *Vetiveria zizanioides* were prepared.

2. Preparation of aqueous extracts

Fine powder, each plant material extracted with sterile distilled water. 20 gm of powder was soaked in 200 ml of sterile distilled water [22]. The extraction process was carried out for 72 hours at room temperature on an orbital shaker. The extracts were dried under a reduced pressure at 40°C using a rotary vacuum evaporator. The dried extracts were stored in sterile bottles at 4°C until further use.

3. Extraction in organic solvents

Each sample was ground into a fine powder and extracted with organic solvents (ethanol and methanol). Solvent extraction was performed as available in reported studies with some modifications of 20 gm of powder, each selected plant material was soaked in 200 mL of each solvent [3], [23-25]. The extraction process was carried out for 72 hours at room temperature on an orbital shaker. The extracts were dried under a reduced pressure at 40°C using a rotary vacuum evaporator. The dried extracts were stored in sterile bottles at 4°C until further use.

Preliminary phytochemical analysis of crude extracts

The phytochemical analysis of all crude extracts was conducted for the detection of carbohydrates, amino acids, proteins, glycosides, tannins, terpenoids, phenols, alkaloids, and saponins, as adopted from previous reports [14], [22], [26-28].

Preparation of plant extracts for antibacterial activity

Stock solutions of *Cinnamomum zeylanicum*, *Syzygium aromaticum*, and *Vetiveria zizanioides* extracts were prepared by mixing them with sterile distilled water. 1 mg of each extract was mixed with 10 mL of sterile distilled water and designated as stock solution. Two concentrations, i.e., 50 and 100 µg/mL, were prepared with the sterile distilled water [29]. These concentrations were used for the evaluation of an antibacterial activity.

Antibacterial activity by diffusion assay

The agar diffusion method has been used for the evaluation of the antibacterial potency of crude extracts. The two dilutions of the extracts were prepared, i.e., 50 and 100 µg/mL using sterile distilled water. In this test, extracts were impregnated on paper discs of size 6 mm and used for assaying [25], [30]. The disc diffusion assay has performed the testing of the antibacterial potential of each extract. Clinical isolates previously isolated, such as *E. coli*, *Staphylococcus aureus*, *Streptococcus faecalis*, *Salmonella paratyphi B*, and *Pseudomonas aeruginosa* were used as test organisms for the diffusion assay. Mueller Hinton agar containing Beef extract: 2.0 g/L, Acid hydrolysate of casein: 17.5 g/L, Starch: 1.5 g/L, Agar-agar: 17.0 g/L, and pH: 7.3±0.2 was used as a basal and seed medium for the antibacterial assay. Sterile basal agar plates and seed agar butts were prepared. After the pouring and solidification of seed agar, sterile filter paper discs were impregnated with the respective crude extract dilution and placed on the surface of the medium in each plate. The discs impregnated with sterile distilled water were also placed on an agar surface as a negative control. Standard antibiotic streptomycin was used in the antimicrobial assay study as a positive control. Then all plates were incubated at 37°C for 24 hours. After incubation, the diameters of the zones of inhibition obtained were measured in mm and recorded.

RESULTS AND DISCUSSION

Qualitative detection of phytochemicals such as carbohydrates, amino acids, proteins, glycosides, tannins, terpenoids, phenols, alkaloids, flavonoids, and saponins in *Cinnamomum zeylanicum*, *Syzygium aromaticum*, and *Vetiveria zizanioides* crude extracts was conducted. Phytochemicals or the presence of metabolites in plants suggest their numerous activities, including protection against various pathogens, may be regarded as safeguards, as well as their pharmacological properties [7], [16], [31-32]. The phytochemicals analysed in all three crude extracts for volatile components are summarised in (Table 1).

Table 1 Phytochemical detected in crude extracts for volatile components

Phytochemicals	Aqueous extracts for volatile components		
	<i>Cinnamomum zeylanicum</i>	<i>Syzygium aromaticum</i>	<i>Vetiveria zizanioides</i>
Carbohydrates	-	-	-
Amino acids and peptides	-	-	-
Glycosides	-	-	-
Tannins	+	-	+
Terpenoids	+	+	+
Phenols	+	+	+
Saponins	-	-	+
Alkaloids	+	+	-

Present: +; Absent: -

Phytochemicals detected in crude aqueous, ethanol, and methanol extracts of *Cinnamomum zeylanicum*, *Syzygium aromaticum*, and *Vetiveria zizanioides* are depicted in (Tables 2-4). It was reported that phytochemicals like alkaloids, flavonoids, tannins, saponins, steroids, glycosides, and terpenoids in the extract of clove, dachini, and khus khus, etc. were majorly responsible for their vital role as pharmacological constituents for the preparation of various drugs [2]. In all three extracts of *Cinnamomum zeylanicum*, *Syzygium aromaticum*, and *Vetiveria zizanioides*, the presence of carbohydrates, amino acids, proteins, glycosides, tannins, terpenoids, phenols,

alkaloids, flavonoids, and saponins was detected. In a previous report, it was mentioned that *Vetiveria zizanioides* extracts evaluated the presence of glycosides, tannins, terpenoids, alkaloids, flavonoids, and saponins [33].

Table 2 Phytochemicals detected in crude extracts of *Cinnamomum zeylanicum*

Phytochemicals	<i>Cinnamomum zeylanicum</i> extracts		
	Aqueous	Ethanol	Methanol
Carbohydrates	+	+	+
Amino acids and peptides	-	-	-
Glycosides	-	+	+
Tannins	-	+	+
Terpenoids	+	+	+
Phenols	+	+	+
Saponins	+	+	+
Alkaloids	+	+	+
Flavonoids	-	+	-

Present: +; Absent: -

Table 3 Phytochemicals detected in crude extracts of *Syzygium aromaticum*

Phytochemicals	<i>Syzygium aromaticum</i> extracts		
	Aqueous	Ethanol	Methanol
Carbohydrates	+	+	+
Amino acids and peptides	-	-	+
Glycosides	+	+	+
Tannins	-	+	+
Terpenoids	-	+	+
Phenols	+	+	+
Saponins	-	+	+
Alkaloids	-	+	+
Flavonoids	-	+	+

Present: +; Absent: -

Antibacterial assay

The antibacterial disc diffusion assay was performed using clinical isolates of *E. coli*, *Staphylococcus aureus*, *Streptococcus faecalis*, *Salmonella paratyphi B*, and *Pseudomonas aeruginosa* as test organisms. *Cinnamomum*

zeylanicum oil is reported to have antibacterial as well as antifungal properties and used for a variety of purposes in medicine [34]. Aqueous extract showed less antimicrobial activity as compared to ethanol and methanol extracts. The 100 µg/mL concentration of all extracts found a higher zone of inhibition against all tested organisms (Tables 5-7). Test organisms *E. coli* and *Streptococcus faecalis* were found susceptible to both concentrations of extracts of all three spices. All extracts exhibited antibacterial activity against the test organisms in comparison with the control. In the present study, the antibacterial activity of the three extracts was evaluated against both Gram-positive and Gram-negative organisms. *Cinnamomum zeylanicum* extract exhibited a higher diameter of the inhibition zone, i.e., 29 and 26 mm against *Escherichia coli*, followed by an extract of *Syzygium aromaticum* with 26 and 25 mm.

Table 4 Phytochemicals detected in crude extracts of *Vetiveria zizanioides*

Phytochemicals	<i>Vetiveria zizanioides</i> extracts		
	Aqueous	Ethanol	Methanol
Carbohydrates	+	+	+
Amino acids and peptides	+	+	+
Glycosides	+	+	+
Tannins	-	-	+
Terpenoids	+	+	+
Phenols	+	+	+
Saponins	+	+	+
Alkaloids	+	+	+
Flavonoids	-	+	+

Present: +; Absent: -

Plants that are rich in tannins have antibacterial potential due to their basic character, which may allow them to react with enzymes and proteins and weaken or pervasion of the plasma membrane, thereby being regarded as bactericidal by altering metabolism [35]. The zones of inhibition were obtained against clinical isolates are depicted in (Table 5-7), which suggests that plant extracts may contain antibacterial components, some of which were analysed as above.

Table 5 The antibacterial effect exhibited by extracts of *Cinnamomum zeylanicum*

Test organism	Concentration of extract, µg/ml	Diameter of Zone of Inhibition, mm			Streptomycin, 100 µg/ml
		Aqueous	Ethanol	Methanol	
<i>Staphylococcus aureus</i>	50	9	12	12	18
	100	8	14	17	
<i>Streptococcus faecalis</i>	50	6	13	12	16
	100	8	16	13	
<i>E. coli</i>	50	18	21	26	32
	100	23	24	29	
<i>Salmonella paratyphi B</i>	50	8	16	14	29
	100	11	14	19	
<i>Pseudomonas aeruginosa</i>	50	7	13	11	21
	100	9	13	15	

Cinnamomum zeylanicum proved its antimicrobial, anticancer, anti-inflammatory, and antidiabetic properties [36]. *Cinnamomum zeylanicum* extracts exhibited higher antibacterial activity against *Escherichia coli* at both concentrations of 50 and 100 µg/mL. All test organisms were found to be sensitive to the *Cinnamomum zeylanicum* extracts. Bark oil extracts of *Cinnamomum zeylanicum* have been

reported as antimicrobials against pathogenic organisms such as *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Staphylococcus aureus*, *Peptococcus*, *Enterococcus faecalis*, *Streptococcus pneumoniae*, *Klebsiella pneumoniae*, and *S. mutans* [1], [24-25], [37-38]. Clove extract has been evaluated as highly inhibitory against *Botrytis cinerea* in laboratory investigation [39]. Ethanol and methanol extracts of

Syzygium aromaticum exerted an antibacterial effect on all test organisms, while *Pseudomonas aeruginosa* was found to be the moderately sensitive. Previously, it was reported that *Syzygium aromaticum* organic extracts were found to be antibacterial against Gram-positive as well as Gram-negative test organisms [1-2], [16]. Oils from the bark extract of *Cinnamomum zeylanicum* have been assessed as antimicrobial and antifungal [41]. *Vetiveria zizanioides* extracts were found antibacterial against bacteria such as *Pseudomonas aeruginosa*, *Salmonella*

paratyphi, and *Staphylococcus aureus* [42]. It was reported that the *Vetiveria zizanioides* used in various preparations such as juice or sharbat, cosmetics, and soaps, shampoos and perfumes, as well as in many skin disorders, and other causes, which demonstrates its usefulness for society [43]. An ethanolic extract of *Vetiveria zizanioides* previously exerted an antibacterial effect against *Mycobacterium tuberculosis* [44]. *Vetiveria zizanioides* was exhibited an effectiveness against dermatophytes and *C. albicans* [45].

Table 6 The antibacterial effect exhibited by extracts of *Syzygium aromaticum*

Test organism	Concentration of extract, µg/ml	Diameter of Zone of Inhibition, mm			Streptomycin, 100 µg/ml
		Aqueous	Ethanol	Methanol	
<i>Staphylococcus aureus</i>	50	7	11	14	18
	100	8	13	17	
<i>Streptococcus faecalis</i>	50	5	14	13	23
	100	10	17	19	
<i>E. coli</i>	50	13	21	22	28
	100	19	25	26	
<i>Salmonella paratyphi B</i>	50	9	12	10	22
	100	10	13	12	
<i>Pseudomonas aeruginosa</i>	50	6	18	21	26
	100	7	21	23	

Table 7 The antibacterial effect exhibited by extracts of *Vetiveria zizanioides*

Test organism	Concentration of extract, µg/ml	Diameter of Zone of Inhibition, mm			Streptomycin, 100 µg/ml
		Aqueous	Ethanol	Methanol	
<i>Staphylococcus aureus</i>	50	-	8	8	18
	100	7	9	10	
<i>Streptococcus faecalis</i>	50	12	10	14	22
	100	10	12	16	
<i>E. coli</i>	50	18	18	19	28
	100	19	19	23	
<i>Salmonella paratyphi B</i>	50	12	19	14	24
	100	14	23	15	
<i>Pseudomonas aeruginosa</i>	50	10	11	16	23
	100	10	14	12	

CONCLUSION

The efficient antibacterial potential and phytochemical content of each spice were assessed as part of the present study. All three of these spices, evidently having potential as antibacterial agents, will may help in the preparation of novel plant-originated preparations and drugs in combinations.

Conflict of interest: The authors have no conflicts of interest regarding this investigation.

Acknowledgments: The author is thankful to the Principal and Head of the Departments of Microbiology, Botany, Chemistry, KANM Sonawane ASC College, Satana, for providing laboratory facilities to carry out the of research work and for their inspiring help.

LITERATURE CITED

- Shete HG, Chitanand MP. 2014. Antimicrobial activity of some commonly used Indian Spices. *International Journal of Current Microbiology and Applied Science* 3(8): 765-770.
- Wankhede TB. 2015. Evaluation of antioxidant and antimicrobial activity of the Indian clove *Syzygium aromaticum* L. Merr. and Perr. *International Research Journal of Science and Engineering* 3(4): 166-172.
- Manandhar S, Luitel S, Dahal RK. 2019. In vitro antimicrobial activity of some medicinal plants against human pathogenic bacteria. *Journal of Tropical Medicine* 1895340. doi.org/10.1155/2019/1895340.
- Patel A, Tiwari S, Pandey N, Gupta D, Sheo MP. 2020. Role of spices beyond a flavouring agent: the antioxidant and medicinal properties. *Ethnopharmacological Investigation of Indian Spices*. pp 5-35. doi.org/ 10.4018/978-1-7998-2524-1.ch002
- Thilagavathi T, Kathiravan G. 2017. Phytochemical analysis and antimicrobial activity of Ethanolic leaf extract of *Ficus racemosa* Linn. *Research Journal of Pharmacy Technology* 10(2): 537-540. doi.org/10.5958/0974-360X.2017.00107.X.
- Manju V, Revathi R, Murugesan M. 2011. In vitro antioxidant, antimicrobial, anti-inflammatory, anthelmintic activity and phytochemical analysis of Indian medicinal spices. *Research Journal of Pharmacy Technology* 4(4):596-9.
- Compean KL, Ynalvez RA. 2014. Antimicrobial activity of plant secondary metabolites: a review. *Research Journal of Medicinal Plants* 8(5):204-13. doi.org/ 10.3923/rjmp.2014.204.213.

8. Walker JRL. 1994. Antimicrobial compounds in food plants. In: (Eds) Dillon VM, Board RG. Natural antimicrobial systems and food preservation. CAB International. pp 181-204.
9. Manigandan M, Saranraj P. 2018. Antimicrobial activity of secondary metabolites in an Indian Spices. *Life Science Archives* 4(1):1248-59. doi.org/10.22192/lisa.2018.4.1.2.
10. Senanayake UM, Lee TH, Wills RBH. 1978. Volatile constituents of cinnamon (*Cinnamomum zeylanicum*) oils. *Journal of Agriculture and Food Chemistry* 26(4):822-4. doi.org/ 10.1021/jf60218a031.
11. D'Souza SP, Chavannavar SV. 2017. Kanchanashri B, Niveditha SB. Pharmaceutical perspectives of spices and condiments as alternative antimicrobial remedy. *Journal of Evidence Based Complementary Alternative Medicine* 22(4):1002-10. doi.org/10.1177/2156587217703214.
12. Srinivasan K. 2014. Antioxidant potential of spices and their active constituents. *Critical Reviews in Food Science and Nutrition* 54(3):352-72. doi.org/10.1080/10408398.2011.585525.
13. Gorgani L, Mohammadi M, Ghasem DN, Nikzad M. 2016. Piperine-the bioactive compound of black pepper: from isolation to medicinal formulations. *Comparative Review in Food Science and Food Safety* 16(1):124-40.
14. Shaikh JR, Patil MK. 2020. Qualitative tests for preliminary phytochemical screening: an overview. *International Journal of Chemical Studies* 8(2):603-8. doi.org/ 10.22271/chemi.2020.v8.i2i.8834.
15. Agidew MG. 2022. Phytochemical analysis of some selected traditional medicinal plants in Ethiopia. *Bulletin of National Research Centre* 46(1):87. doi.org/10.1186/s42269-022-00770-8.
16. Abd EAMHM, El-Mesallamy AMD, El-Gerby M, Awad A. 2014. Anti-tumor, antioxidant and antimicrobial and the phenolic constituents of clove flower buds (*Syzygium aromaticum*). *Journal of Microbiology and Biochemical Technology Suppl.* 8(007). doi.org/10.4172/1948-5948.S8-007.
17. Jain SK. 1991. *Dictionary of Indian Folk Medicine and Ethno-Botany*. Deep Publishing.
18. Parham S, Kharazi AZ, Bakhsheshi-Rad HR, Nur H, Ismail AF, Sharif S. 2020. Antioxidant, antimicrobial and antiviral properties of herbal materials. *Antioxidants (Basel)*. 9(12):1309. doi.org/10.3390/antiox9121309.
19. Rao PV, Gan SH. 2014. Cinnamon: A multifaceted medicinal plant. *Evidence Based Complement Alternative Medicine* 642942. doi.org/10.1155/2014/642942.
20. Rao UM, Abdurrazak M, Suryati KM. 2016. Phytochemical screening, total flavonoid and phenolic content assays of various solvent extracts of tepal of *Musa paradisiaca*. *Malaysian Journal of Analytical Science*. 20(5):1181-90. doi.org/10.17576/mjas-2016-2005-25.
21. Rasul MG. 2018. Extraction, isolation and characterization of natural products from medicinal plants. *International Journal of Basic Science and Applied Computer* 2(6):1-6.
22. Pandey S. 2014. Study of preliminary phytochemical screening and antibacterial activity of *Tribulus terrestris* against selected pathogenic microorganisms. *Journal of Bioanalysis and Biomedical Suppl* 12(001). doi.org/10.4172/1948-593X.S12-001.
23. Singh J, Bhatnagar SK, Tomar A. 2019. Study on fungicidal effect of plant extracts on plant pathogenic fungi and the economy of extract prepa-ration and efficacy in comparison to synthetic/chemical fungicides. *Journal of Applied and Natural Science* 11(2):333-7. doi.org/ 10.31018/jans.v11i2.2053.
24. Adarsh A, Chettiyar B, Kanthesh BM, Raghu N. 2020. Phytochemical screening and antimicrobial activity of *Cinnamomum zeylanicum*. *International Journal of Pharmaceutical Research Innovations* 13:22-33.
25. Abdulla EH, Abdoun MA, Mahmoud WS, Alhamdani F. 2019. Antibacterial activity of crude *Cinnamomum zeylanicum* ethanol extract on bacterial isolates from orofacial infections. *Acta Science Dental Science* 3(8):58-63. doi.org/10.31080/ASDS.2019.03.0595.
26. Trease GE and Evans WC. 2005. Pharmacognosy. 15th ed. W B Saunders.
27. Vasait RD. 2017. Detection of phytochemical and pharmacological properties of crude extracts of *Tribulus terrestris* collected from tribal regions of Baglan (M.S.), India. *International Journal of Pharmacognosy and Phytochemical Research* 9(4):508-11. doi.org/10.25258/phyto.v9i2.8122.
28. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. 2011. Phytochemical screening and extraction: a review. *International Pharmaceutical Science* 1(1):98-106.
29. Saliem AH, Abedsalih AN. 2018. Evaluation the antibacterial properties of different extracts of *Cinnamomum zeylanicum* barks. *Advances in Animal Veterinary Science* 6(9):380-3. doi.org/ 10.17582/journal.aavs/2018/6.9.380.383.
30. Ezzatzadeh E, Farjam MH, Rustaiyan A. 2012. Comparative evaluation of antioxidant and antimicrobial activity of crude extract and secondary metabolites isolated from *Artemisia kulbadica*. *Asian Pacific Journal of Tropical Disease* 2: S431-4. doi.org/ 10.1016/S2222-1808(12)60198-4.
31. Singh G, Singh BS, Kumar BRV. 1978. Antimicrobial activity of essential oils against keratinophilic fungi. *Indian Drugs*. 16(2):43-5.
32. Cortés-Rojas DF, de Souza CRF, Oliveira WP. 2014. Clove (*Syzygium aromaticum*): A precious spice. *Asian Pacific Journal of Tropical Biomedicine* 4(2):90-6. doi.org/10.1016/S2221-1691(14)60215-X.
33. Dutta S, Mondal S, Hazra K, Mangal AK. 2022. Bolleddu R, Dahiya J, Kharwar S, Prasad PVV. Pharmacognostical evaluation and phytochemical characterisation of *Vetiveria zizanioides* (L.) Nash roots; *International Journal of Pharmacognosy* 9(5):105-12. doi.org/10.13040/IJPSR.0975-8232.IJP.9(5).105-12.
34. Carmo ES, de Oliveira Lima EDO, De Souza EL, De Sousa FB. 2008. Effect of *Cinnamomum zeylanicum* blume essential oil on the growth and morphogenesis of some potentially pathogenic aspergillus species. *Brazilian Journal of Microbiology* 39(1):91-7. doi.org/10.1590/S1517-838220080001000021.
35. Cano A, Andres M, Chiralt A, González-Martínez C. 2020. Use of tannins to enhance the functional properties of protein-based films. *Food Hydrocolloids* 100. doi.org/ 10.1016/j.foodhyd.2019.105443.
36. Chandula Weerasekera A, Samarasinghe K, Krishantha Sameera de Zoysa H, Chathuranga Bamunuarachchige T, Yashasvi Waisundara V. 2021. *Cinnamomum zeylanicum*: morphology, antioxidant properties and bioactive compounds. *IntechOpen*. doi.org/ 10.5772/intechopen.97492.

37. Alsalam HAA, Shawkat MS, Khwaildy MIA. 2017. Antibacterial activity of *Cinnamomum zeylanicum* bark oil and cinnamaldehyde on some locally isolated pathogenic bacteria. *World Journal of Pharmaceutical Research* 6(2):174-85.
38. Thiagarajan S, John S. 2020. Antimicrobial activity of *Cinnamomum zeylanicum* aqueous extract against bacteria and fungi responsible for urinary tract infection. *International Journal of Health and Allied Sciences*. 9(3):229-32. doi.org/10.4103/ijhas.IJHAS_3_20.
39. Šernaitė L, Rasiukevičiūtė N, Valiuškaitė A. 2020. The Extracts of Cinnamon and Clove as Potential Biofungicides against Strawberry Grey Mould. *Plants (Basel)* 9(5): 613. doi.org/ 10.3390/plants9050613.
40. Salih SS, Faraj NM, Hamarash AM. 2014. Effect of plant extract *Eugenia caryophyllus*, *Cinnamon zeylanicum* on antibiotic resistant from *Staphylococcus aureus*. *Brazilian Journal of Pharmacology and Toxicology* 5(4):125-8. doi.org/10.19026/bjpt.5.5178.
41. Husain SS, Ali M. 2013. Analysis of volatile oil of the stem bark of *Cinnamomum Zeylanicum* and its antimicrobial activity. *International Journal of Research on Pharmaceutical Science* 3(4):40-9.
42. Soni A, Dahiya P. 2015. Screening of phytochemicals and antimicrobial potential of extracts of *Vetiver zizanioides* and *Phragmites karka* against clinical isolates. *International Journal of Applied Pharma* 7(1): 22-24. doi.org/10.13140/RG.2.1.2041.5203.
43. Bharti V, Mehta A, Mourya GK, Ayoub Z. *Vetiveria zizanioides* (Khas-Khas): A Medicinal Grass. *Med Plants India Importance Cultivation Inbook*. 2020: 424-35.
44. Saikia D, Parveen S, Gupta VK, Luqman S. 2012. Anti-tuberculosis activity of Indian grass KHUS (*Vetiveria zizanioides* L. Nash). *Complementary Theoretical Medicine* 20(6):434-6. doi.org/10.1016/j.ctim.2012.07.010.
45. Dos Santos DSD Oberger JV, Niero R, Wagner T, Delle Monache F, Cruz AB, Martin-Quintal Z, Cechinel Filho V. 2014. Seasonal, phytochemical study and antimicrobial potential of *Vetiveria zizanioides* roots. *Acta Pharma* 64(4): 495-501. doi.org/10.2478/acph-2014-0040.