

Analysis of Seed Dormancy Breaking in Buffel Grass (*Cenchrus ciliaris*) via Experimental Procedure

Manal Ahmed^{*1-2} and Bakri Mohamed³

¹Central Laboratory, Seed Administration, Ministry of Agriculture and Forests, Khartoum, Sudan

²Wuhan University of Technology Nanhu Campus East Gate International Student Apartment 430 070, Hongshan District, Wuhan, Hubei Province

³Department of Agronomy, Faculty of Agriculture, Khartoum University, Sudan

Abstract

In this article, an experimental analysis of Buffel grass (*Cenchrus ciliaris*) was conducted. This experiment was conducted at the Central Laboratory, Seed Administration-Ministry of Agriculture and Forests-Khartoum-Sudan. The case study and experimental procedure are carried out on the different effects of dormancy-breaking methods on seeds of Buffel grass (*Cenchrus ciliaris*). In the experiment, three techniques were used for seed-breaking dormancy. The acid was added as an additive to increase the prevention and germination levels of seeds. These acids are sulfuric acid (H₂SO₄) with a concentration of 50, 70, and 90%, and Gibberellic acid with levels of 50, 100, 150, 200, 250, and 300 ppm respectively suggested in the experiment. Moreover, the hot water (H.W) was considered at 60 °C for 15, 30, 45, 60, and 75 minutes at different intervals. The experiment was carried out in a completely randomized design (CRD) with five replications. These experiments are based on random samples to check and test the germination rate and other related parameters of the seeds and validate the quality of the seed. This thing improves the satisfaction of the customer as germination rate. Germination percentage and rate of germination were recorded for the seeds to study the effects of treatments. The results demonstrate that, in comparison to the control and other treatments, the Gibberellic acid treatment with a concentration of 300 ppm provided the highest germination percentage (71.86), germination rate (62.20), and mean germination percentage (43.24). Following that, the germination percentage was determined using sulfuric acid at a 90% concentration (63.52).

Key words: *Cenchrus ciliaris*, Germination percentage, Gibberellic acid, Germination rate, Least square design (LSD), Mean germination percentage, Sulfuric acid

A grass known as Buffel grass, or *Cenchrus ciliaris*, is indigenous to southern Asia, as well as parts of Africa, Arabia, the Middle East, and India. It was brought to Australia in the late 19th century, and today it may be found all over tropical, subtropical, and warm temperate regions. In many locations, it is either produced or naturalized. It has been classified as poisonous in some places and invasive in dry and semi-arid habitats (northern America, Hawaii, Mexico, and northern Australia). In the majority of warm, dry, and semi-arid regions of the world, it has been cultivated for fodder and to manage erosion. It frequently eludes plantings, particularly in disturbed areas where it fosters a cycle of grass fires. Invaded ecosystems can change, changing ecosystem processes and endangering native plants and animals [1-4] due to increased fire frequency and intensity as well as thick Buffel grass growth.

Buffel grass is frequently associated with dispersed woody legumes like *Leucaena leucocephala*, *Acacia spp.*, and *Prosopis spp.* It may co-occur with other drought-tolerant grasses such as *Urochloa maxima*, *Panicum maximum*, *Chloris*

spp., and *Eragrostis spp.* Along seasonal dry riverbanks and drainage pathways in South Africa, a population of *Cenchrus ciliaris* and *Cyperus arginatus* has been identified [5]. On sandy (fast-draining) soils with annual rainfall as high as 1200 mm, buffelgrass has been documented to persist [6]. Magnaporthe (*Pyricularia grisea*) is a dangerous fungus that has led to the dieback of Buffel grass in several nations and regions. The research [7] depicts a few more insects. Around the world, the most crucial factors for feeding on seed heads are seed quality and affordability [8-9].

Seed dormancy

Dormancy can be broken by most ideal growing conditions. A seed is necessary for a little plant whose life activities are at their most basic. A seed that is physiologically given enough water, and oxygen for proper aerobic metabolism, and an environment with a temperature that does not exceed physiological limitations but does not germinate is said to be dormant [10-11].

Received: 08 Aug 2023; Revised accepted: 12 Oct 2023; Published online: 19 Oct 2023

Correspondence to: Manal Ahmed, Central Laboratory, Seed Administration, Ministry of Agriculture and Forests, Khartoum, Sudan, Tel: +8618607180576; E-mail: manalchina2020@gmail.com

Citation: Ahmed M, Mohamed B. 2023. Analysis of seed dormancy breaking in buffel grass via experimental procedure. *Res. Jr. Agril. Sci.* 14(5): 1608-1614.

Breaking the seed dormancy

Scarification Bonner is a general term for any procedure that eliminates or lowers the impermeability of the seed coat. The degree and type of seed dormancy have a significant impact on the effectiveness of treatment. Making a tiny hole in the seed coat before sowing is one of the simplest and most direct physical approaches [12-14]. According to a study indicating that the hardest-coated seeds become accessible to water when the seed coat is damaged or pierced by mechanical abrasion or chemical treatment, the percentage of *Acacia* seeds that germinate increased from 7% in control to 19% in scarified seeds [15]. Buffel grass (*Cenchrus ciliaris* L.) is a preferred species for erosion management of railway banks in the semi-arid regions of Central Queensland, Australia, due to its wide distribution, rapid regeneration, and simplicity of maintenance. The species has a deeper and larger root system that can provide higher strength against erosion when compared to other grasses like Rhodes (*Chloris gayana* L.) and green panic (*Panicum maximum* var. *tracheoles*) [16].

By storing carbohydrates at the base of the swelling stem, the plant can withstand periods of drought, grow again after burning [17], and start to receive rain [18]. Buffel grass is a great option for dry communities and the colonization of disturbed lands due to its ability to regenerate after burning and resistance to drought [19].

If rapid vegetation establishment is achieved without delaying germination when batter slopes are constructed, buffelgrass can effectively reduce soil erosion on railway banks. However, because of the inherent dormancy of seeds, low seed germination when planted on railway ties prevents the development of a good grass cover within a constrained timeframe and may have a detrimental effect on soil erosion control [20].

To determine the most efficient method of reawakening four *Adesmia* species from dormancy, Tedesco conducted germination studies. The outcomes showed that mechanical scarification was a successful method for obtaining a significant number of germinated seeds, as opposed to immersion in hot water at 60°C and undamaged seeds as a control. By scarifying the *Sporades corilifolia* using sandpaper, intact seeds obtained 90% germination. Physical scarification, as opposed to chemical scarification with hydrochloric acid, was found to be more efficient at releasing *Rhynchosia minima* L. seeds from dormancy by Shaukat and Burhan [21-22]. *Cassia moschata* and *Entada polyarchy* had the best germination rate and seedling emergence when mechanical scarification was performed on the radishes on the opposite side.

According to Ghadiri and Torshiz [23], mechanical scarification improved *Glycyrrhiza glabra* L. seed germination to 49–98%. As per Girase *et al.* [25], mechanical scarification can remove the dormancy that exists in the seeds of *Acacia auriculiformis* and *A. torilis*. Histochemical analysis of the seed coat of *Leucaena glauca* L. revealed that pectin served as a water barrier and that the only method for removing the seed coat's dormancy was mechanical scarification. According to the research of Baskin and Baskin [26], mechanical scarification can totally end the dormancy of *Dalea foliose* seed, which is caused by the seed coat's water-impermeable nature.

Many aborigines' seeds have responded well to hot water treatment [27]. Typically, boiling water is used to soak the seeds before the water is quickly removed from the heat source and allowed to cool gradually. For 12 hours, the seeds are submerged in water [28]. Both the modification or softening of the hard seed coat and the leaching out of chemical inhibitors are effects of wet treatment. Levitt [29] discovered that seeds that don't expand while being boiled can endure being boiled in

water for a number of hours. Using hot water to treat a variety of leguminous seeds has produced positive outcomes. The effectiveness of boiling water on the germination of three *Acacia* species was observed by Brown and Booysen [30]. With 3–5 minutes of soaking in hot water (80-88 °C).

A. Gregii's germination percentage increased considerably

In a similar manner, Clemens *et al.* [31] reported a substantial impact of hot water at three different temperatures for seven different time periods on five *Acacia* species. According to Wickens [32], the treatment of *Acacia albida* seeds with almost boiling water did not alter their characteristics from untreated seeds. The application of hot water therapy is very simple, safe, and successful.

Based on the literature review, the method suggested for that analysis is the least different design method to establish the model based on the data. The data were analyzed according to the scale and rules.

The objective of this study is to find the different effects/methods for breaking dormancy in the Buffel grass seeds in this study, the experimental procedure about the Buffel grass seed breaking dormancy with the help of the Ministry of Agriculture and Forests- Khartoum-Sudan was adopted. Furthermore, the germination percentage improvement and the period at which maximum germination rate is achieved.

This article is structured as follows: In Section 1, the literature review is discussed about Buffel grass and its production behavior. In Section 2, materials and methods are discussed. This section also describes the data collection methods and analysis methods. Section 3 covers the comprehensive procedure of results and a discussion of the proposed methods. In Section 4, the concluding remarks about the experimental analysis of germination are deliberated.

MATERIALS AND METHODS

A laboratory experiment was carried out to study the effect of different methods for breaking seed dormancy in Buffel grass (*Cenchrus ciliaris*). The study was conducted at the laboratory of the Seed Administration Ministry of Agriculture and Forestry, Khartoum, Sudan. This section is composed of two subsections. One is the materials about the research and the second is a method. The methodology is described to conduct the research.

Plant material

Buffel grass samples were taken from the fields such as Shabbat. The field locations were (latitude 15°40' N, longitude 32°32' E, and 380m above sea level). The climate of the site is semi-desert and the soil is alkaline cracking clay with low permeability. The data collected from the fields, farmers, and surveys of the fields at the different locations, the nature of land and sand are different to check and validate the germination process and their related parameters.

Reagents

There are different reagents are used to boost the germination process and control the insects during the germination.

Sulfuric acid (H_2SO_4)

Sulfuric acid was used during the experiment and acid was obtained from the National Center for Research, Khartoum, Sudan.

Gibberellic acid (GA_3)

The second reagent is gibberellic acid (300 ml/L). It was provided by the Department of Horticulture University of Khartoum. In concentration, 0.3 gram per liter other concentrations were obtained by diluting from the stock solution. The different dilutions are obtained separately.

In this section, the research methodology is used. The different properties of seeds are evaluated.

Treatments: The process of treatment plays a key role in the seed quality, and germination rate and also improves the germination rate. This treatment improves the quality of the seed to prevent the different insects which damage the seed quality. The seeds of Buffel grass were subjected to the following treatments:

Control: No treatment was applied to the seeds

Hot water: A sub-sample of the seeds was soaked in hot water (60 °C) for 15, 30, 45, 60 and 75 minutes. The different timeframes were adopted to see the effects on the seeds.

Sulfuric acid (H₂SO₄): The different concentration of sulfuric acid was used. Seed samples were soaked in 50, 70, and 90% concentration of the acid. Thereafter the seeds were thoroughly rinsed several times in distilled water to remove any trace of the acid.

Gibberellic acid (GA₃): Six different concentrations of 50, 100, 150, 200, 250, and 300 PPM of GA₃ were prepared by dilution using distilled water from the stock (0.3g/L), and sub-samples of the sterilized seeds were soaked in each of the concentrations in the dark at room temperature for two hours.

Germination test

This experiment was carried out to determine how the treatments affected the Buffel grass seeds' ability to germinate. A completely randomized design with five replications was used to conduct the test. Each replication included 25 seeds in it. The seeds are prepared using various dormancy-breaking techniques, then stored in sterilized Petri dishes with a 9 cm diameter on double-layered filter paper that has been soaked with 8 cc of distilled water. After that, the Petri plates were put in an incubator set to 30 °C. For 55 days, germination seeds were counted and taken out every 24 hours. When the radical tip had separated from the seed coat, a seed was said to have

germinated. The following parameters were then obtained using the collected data:

Germination percentage (%)

It is the ratio of germinated seeds to the total sprouted seeds. The number of germinated seeds was recorded daily and the germination percent was calculated as follows:

$$\text{Germination (\%)} = \frac{\text{No. of germinated seeds}}{\text{Total No. of seeds pe petri dish}}$$

Rate (speed) of germination (Gr)

The rate of germination was calculated according to Wiese and Binning [33] as follows:

$$\text{Germination rate (\%)} = \frac{1 + 2 + 3 + \dots + N}{\text{Total No. of seeds}}$$

Where; 1, 2, 3, N are counting days from day 1 to N. speed of germination expresses the rate of germination in terms of the total number of seeds that germinate in a time interval. Higher values indicate greater and faster germination.

Statistical analysis

According to the procedure outlined by Gomez and Gomez [34] for a fully randomized design, the gathered data were subjected to analysis of variance. After that, mean separation was carried out utilizing the least significant difference (LSD) technique. Stat analysis was used to perform the computations.

RESULTS AND DISCUSSION

A: Germination percentage of Buffel grass seeds

There are different treatment cases done for germination rate. These steps improve the quality of seeds and germination rate and also boost the optimization level of the germination. The obtained results can be summarized as follows:

Case 1: At 10 days

Buffel grass germination percentages in the various treatments varied significantly ($P \leq 0.01$) after ten days, according to an analysis of variance. The treatments were divided into three groups in (Table 1) based on the mean germination. The GA₃ 300ppm condition yielded the highest germination rate of 14.85.

Table 1 Mean germination percentage of Buffel grass seeds during the period 10 to 22 day after germination

Treatments	Days 10	Days 13	Days 16	Days 19	Days 22
Control	1.81	1.81	1.81	1.81	1.81
H ₂ SO ₄ 50	11.54	12.31	12.31	20.93	23.51
H ₂ SO ₄ 70	12.31	13.09	13.09	22.25	24.76
H ₂ SO ₄ 90	16.62	18.62	18.92	28.32	29.74
GA ₃ 50 PPM	13.09	15.62	15.62	21.59	25.92
GA ₃ 100 PPM	12.31	14.84	14.84	22.85	27.66
GA ₃ 150 PPM	16.39	18.33	18.33	27.07	30.90
GA ₃ 200 PPM	17.36	18.33	18.33	25.3	29.3
GA ₃ 250 PPM	16.39	17.36	17.36	28.69	30.80
GA ₃ 300 PPM	18.87	25.81	20.91	29.77	34.31
Hot water 60 °C / 15min	1.81	1.81	1.81	3.66	4.15
Hot water 60 °C / 30min	1.81	1.81	1.81	4.22	5.99
Hot water 60 °C / 45min	6.48	7.14	8.37	7.65	9.25
Hot water 60 °C / 60min	1.81	1.81	1.81	3.81	3.96
Hot water 60 °C / 75min	1.81	1.81	1.81	1.81	3.79
Mean	9.96	10.28	11.21	20.21	22.42
5% LSD	3.38	3.41	3.54	3.02	3.16

This germanium rate is not significantly greater than GA₃ 250ppm, GA₃ 200ppm, GA₃150ppm, and H₂SO₄ (70%), but was significantly higher than all other treatments as well as the control are shown in (Table 1, Fig 1-3). On the other hand, the lowest value which is 1.81 was recorded in all hot water treatments and the control.

Case 2: At 16 days

After sixteen days of treatment, an analysis of the data showed that there were major variations ($P \leq 0.01$) between the various treatments in the proportion of Buffel grass that germinated. The GA₃ 300ppm and GA₃ 250ppm concentrations yielded the highest germination percentages 25.85 and 25.81, respectively. They were not significantly greater than GA₃ 200 ppm, GA₃ 150 ppm, and H₂SO₄ (90%), but were significantly higher than all other treatments as well as the control are shown in (Table 1, Fig 1-3). On the other hand, the low values (3.14)

and (3.05) were recorded in H.W 60 °C/30min and 45min, the lowest values (1.81) were recorded in the remaining hot water treatments, and the control is also shown in (Table 1, Fig 1-3).

Case 3: At 31 days

Analysis of variance demonstrated highly significant deviations ($P \leq 0.01$) in the germination amount throughout the various treatments after thirty-one (31) day sessions. In GA₃ 300ppm, the highest germination percentage (44.65) was discovered.

It was not significantly greater than GA₃ 250ppm GA₃ 200ppm, GA₃ 150ppm, and H₂SO₄ 90%, but was significantly higher than all other treatments as well as the control are shown in (Table 2, Fig 1-3).

Moreover, all hot water treatments were not significantly different from the control and recorded lower germination percentage values in (Table 2).

Table 2 Mean germination percentage of Buffel grass during the period 25 to 37 days after germination

Treatments	Days 25	Days 28	Days 31	Days 34	Days 37
Control	1.81	1.81	1.81	1.81	1.81
H ₂ SO ₄ 50	25.92	28.74	30.87	32.42	35.87
H ₂ SO ₄ 70	27.07	29.32	31.92	32.75	35.89
H ₂ SO ₄ 90	35.92	38.48	41.28	43.21	46.84
GA ₃ 50 PPM	28.74	31.40	32.91	33.41	38.28
GA ₃ 100 PPM	29.85	33.41	35.40	36.35	40.15
GA ₃ 150 PPM	34.45	35.41	38.28	38.28	42.93
GA ₃ 200 PPM	32.95	36.85	39.56	40.50	44.08
GA ₃ 250 PPM	34.85	38.23	40.95	42.34	45.34
GA ₃ 300 PPM	36.32	39.88	43.26	44.65	49.31
Hot water 60 °C / 15min	3.22	3.33	4.52	4.55	5.25
Hot water 60 °C / 30min	6.08	6.25	6.96	7.16	7.82
Hot water 60 °C / 45min	10.25	11.58	13.58	14.56	15.23
Hot water 60 °C / 60min	3.97	3.99	4.16	4.89	5.16
Hot water 60 °C / 75min	3.11	3.19	3.35	3.66	4.12
Mean	24.89	27.41	29.82	30.32	34.19
5% LSD	3.42	3.74	3.88	3.67	3.74

Case 4: At 49 days

The analysis of variance revealed highly significant variations ($P \leq 0.01$) in the percentage of grass germination across the various treatments at day forty-nine. The treatment GA₃ 300ppm always gave the highest germination percentage

62.2 which is shown in (Table 3, Fig 1-3). All GA₃ 250ppm, GA₃ 200ppm, GA₃ 150ppm, and GA₃ 100ppm continued to give higher germination percentages, whereas all hot water treatments gave lower germination values than other treatments.

Table 3 Mean germination percentage of Buffel grass during the period 40 to 55 days after germination

Treatments	Days 40	Days 43	Days 46	Days 49	Days 52	Days 55
Control	1.81	1.81	1.81	1.81	1.81	1.81
H ₂ SO ₄ 50	39.09	42.35	44.88	50.67	48.54	52.67
H ₂ SO ₄ 70	40.06	42.93	45.92	48.91	52.18	56.05
H ₂ SO ₄ 90	48.45	51.25	54.26	57.38	59.93	63.52
GA ₃ 50 PPM	40.15	41.89	43.85	44.31	45.46	51.24
GA ₃ 100 PPM	44.07	45.69	48.45	49.39	50.77	53.05
GA ₃ 150 PPM	46.49	47.88	50.31	51.71	53.61	58.71
GA ₃ 200 PPM	47.53	50.32	52.61	54.11	55.36	61.95
GA ₃ 250 PPM	48.33	50.44	53.86	55.33	58.37	62.94
GA ₃ 300 PPM	53.31	55.83	58.98	62.23	63.51	71.86
Hot water 60 °C / 15min	9.22	10.35	11.11	12.03	15.44	15.81
Hot water 60 °C / 30min	9.98	10.66	11.95	12.48	15.78	16.79
Hot water 60 °C / 45min	16.23	17.58	17.91	18.24	18.65	19.39
Hot water 60 °C / 60min	7.66	8.96	9.45	9.81	10.87	12.37
Hot water 60 °C / 75min	6.22	7.33	8.24	9.29	10.41	11.47
Mean	37.24	37.79	38.64	38.64	41.84	43.24
5% LSD	3.88	3.85	3.75	3.757	3.68	4.13

Case 5: At 55 days:

The treatment GA₃ 300ppm continued to provide the best germination percentage at 55 days, which is 71.86. The highest germination values were provided by GA₃ 250 ppm (H₂SO₄

90%), GA₃ 200 ppm, GA₃ 150 ppm, and GA₃ 100 ppm, which were significantly greater than the control and other treatments. However, the mean separation was observed to rise in all treatments except for these four.

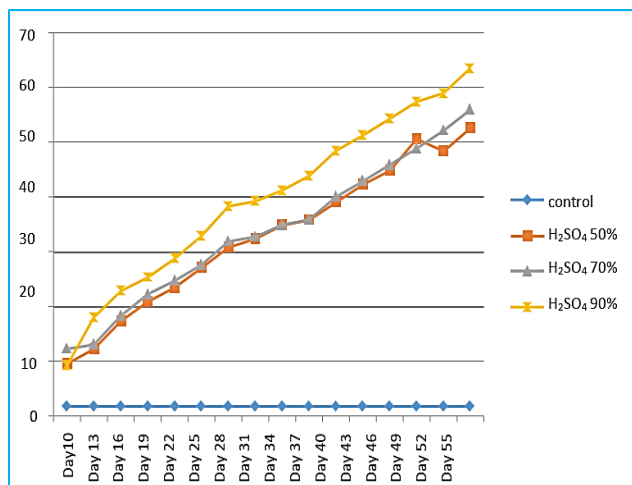


Fig 1 H₂SO₄ effects on pre-treatments on germination (10-55 days after treatments)

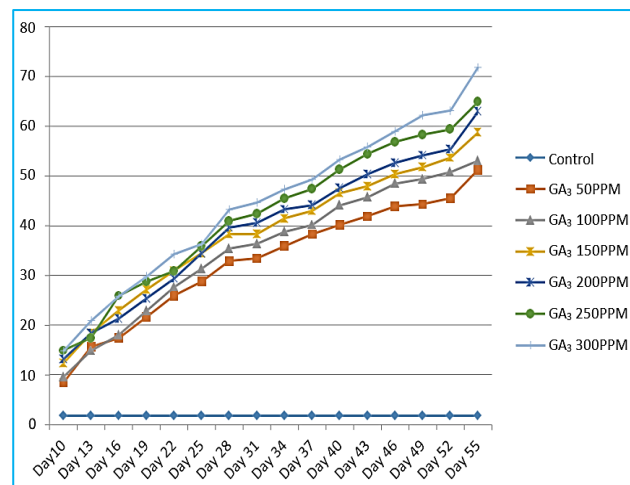


Fig 2 GA₃ effects on pre-treatments on germination (10-55 days after treatments)

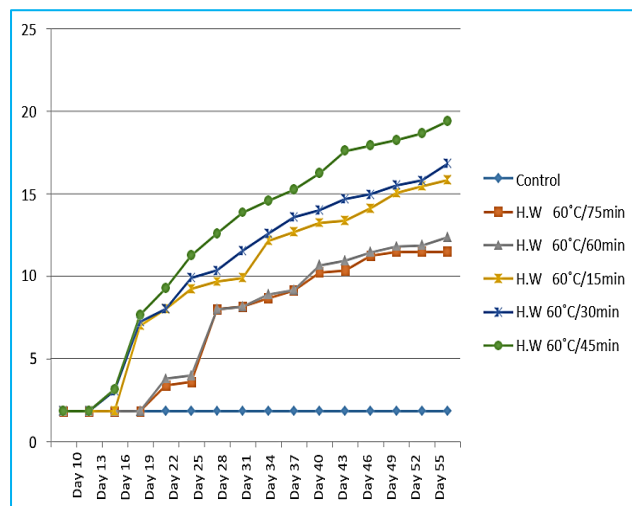


Fig 3 Hot water effects on pre-treatments of germination (10-55 days after treatments)

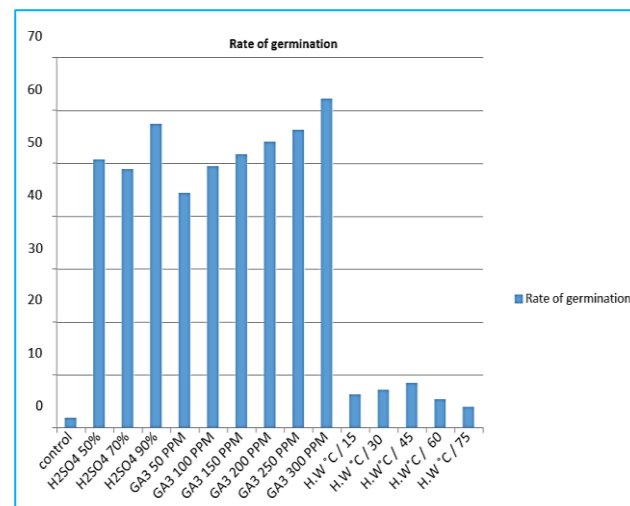


Fig 4 Germination rate effects after 55 days experiment

B: Germination rate

In this section, the germination rate of the seed is calculated through the least squared difference. The scale is set by the variance to estimate the level of germination rate. The germination rate of Buffel grass during the lab experiment of 55 days is shown in (Fig 4). This behavior shows a highly significant difference with respect to the scale $P \leq 0.01$. The germination treatments are shown in (Table 4). This data and facts of results show that the germination rate follows the linear rate of germination. The results show that the highest germination rate was recorded for GA₃ 300ppm, which is highly significant from all remaining treatments. In the proceeding of the results, the higher the concentration (level of sulfuric acid) is, the germination rate is higher than the previous dilution rate such as H₂SO₄ (90%) is a higher germination rate than H₂SO₄ (70%). but gibberellic acid (GA₃) effects are also considerable on the germination rate. The higher level of gibberellic acid (GA₃) increases the germination rate such as GA₃ (250) Higher germination was recorded than other dilutions. All hot water treatments recorded the lowest germination rate value, which is shown in (Table 4).

Table 4 Rate of germination in Buffel grass seeds after germination treatments for breaking dormancy under laboratory conditions, for 55 days

Treatments	Rate of germination
Control	1.81
H ₂ SO ₄ 50	50.67
H ₂ SO ₄ 70	48.91
H ₂ SO ₄ 90	57.38
GA ₃ 50 PPM	44.31
GA ₃ 100 PPM	49.39
GA ₃ 150 PPM	51.71
GA ₃ 200 PPM	54.11
GA ₃ 250 PPM	56.33
GA ₃ 300 PPM	62.20
Hot water 60 °C / 15min	6.26
Hot water 60 °C / 30min	7.13
Hot water 60 °C / 45min	8.42
Hot water 60 °C / 60min	5.34
Hot water 60 °C / 75min	3.89

The results of this study showed that gibberellic acid (GA₃) in different concentrations gave a significantly higher germination percentage than the other treatments. This demonstrated that GA₃ was effective in removing these seeds' dormancy. These outcomes indicate that GA₃ might be involved in the control of both protein and nucleic acid metabolism during dormancy. These results suggest that the enhancement of germination might occur due to the role played by GA₃ in breaking endogenous dormancy. These results agree with those reporting that the promotion of germination may be due to the fact that, GA₃ at increased concentration may act as a signal for amylase production, which plays an essential role in seed germination in rice seed, and in grass *Leymus arenarius*. The results indicated that seeds of *Cenchrus ciliaris* did not exhibit physical dormancy (no seed coat dormancy was found in the seeds), and the type of seed dormancy was endogenous, physiological, or morphophysiological.

CONCLUSION

The concluding remarks are written as:

1. The optimization is achieved at 300ppm because the breaking seed dormancy in Buffel grass seed is the application of GA₃. This is the best and most convenient method to achieve the level of optimization.
2. The reliable achievements are obtained by the experiment by using sulfuric acid. The different dilutions of acid are used to achieve the maximum breaking seed dormancy in Buffel grass. The optimum concentration is 90%.

The future recommendation based on the experiment is to emphasize on using seeds of different ages treated with GA₃ in different concentrations and H₂SO₄, to determine the appropriate concentration of GA₃ to be used for breaking seed dormancy in Buffel grass seeds.

LITERATURE CITED

1. Adam HS. 2002. *The Agriculture Climate*. Second Edition. (In Arabic) Gezira University Press. Wad Madani, Sudan. pp 119.
2. Abd El Dafai AER. 1977. Growth and vegetative propagation of some of the indigenous tree species of the Sudan (Doctoral dissertation, UOFK).
3. Aganga AA, Autlwtse MN. 2000. Utilization of sorghum forage, millet forage, veldt grass and Buffel grass by Tswana sheep and goats when fed *Lablab purpureus* L. as a protein supplement. *Asian-Australasian Journal of Animal Sciences* 13(8): 1127-1132.
4. Bhattarai SP, Fox J, Gyasi-Agyei Y. 2008. Enhancing Buffel grass seed germination by acid treatment for rapid vegetation establishment on railway batters. *Journal of Arid Environments* 72(3): 255-262.
5. Hacker JB, Ratcliff D. 1989. Seed dormancy and factors controlling dormancy breakdown in Buffel grass accession from contrasting provenances. *Journal of Applied Ecology* 26(1): 201-212.
6. Cox JR, Fourie JH, Rethman NFG, Wilcox DG. 1988. The influence of climate and soils on the distribution of four African grasses. *Journal of Range Management* 41(2): 127-139.
7. Díaz Franco A, Méndez Rodríguez A, Garza Cedillo R. 2007. Buffelgrass leaf blight: its presence in Tamaulipas, Mexico. *Agricultura Técnica en México* 33(3): 285-295.
8. Hartman GL, West ED, Herman TK. 2011. Crops that feed the World 2. Soybean— worldwide production, use, and constraints caused by pathogens and pests. *Food Security* 3: 5-17.
9. Bebawi FF, Mohamed SM. 1985. The pretreatment of seeds of six Sudanese Acacias to improve their germination response. *Seed Science and Technology* 13(1): 111-119.
10. Bradbeer JW. 2013. *Seed Dormancy and Germination*. Springer Science and Business Media.
11. Friedel MH, Puckey H, O'Malley C, Waycott M, Smyth A, Miller G. 2006. Buffel grass: Both friend and foe: An evaluation of the advantages and disadvantages of buffel grass use, and recommendations for future research: A report to the desert knowledge cooperative research centre on the dispersal, impact, and management of Buffel grass (*Cenchrus ciliaris*) in desert Australia.
12. Girase US, Jambhale ND, Suryawanshi YB. 2002. Germination and seed dormancy studies in *Acacia species*. *Seed Research* 30(1): 39-42.
13. Jackson J. 2004. Impacts and management of *Cenchrus ciliaris* (Buffel grass) as an invasive species in northern Queensland. Doctoral dissertation, James Cook University.
14. Kak A, Chitra Devi L, Gupta V, Singh N. 2006. Response of seed treatments on seed germination in wild *Crotalaria species*. In: 1st International Conference on Indigenous Vegetables and Legumes. *Prospectus for Fighting Poverty, Hunger and Malnutrition* 752: 261-265.
15. Owens MK, Wallace RB, Archer S. 1995. Seed dormancy and persistence of *Acacia berlandieri* and *Leucaena pulverulenta* in a semi-arid environment. *Journal of Arid Environments* 29(1): 15-23.
16. Harwood JL. 1999. Managing risk in farming: concepts, research, and analysis (No. 774). US Department of Agriculture, ERS.
17. Bhattarai SP, Fox J, Gyasi-Agyei Y. 2008. Enhancing Buffel grass seed germination by acid treatment for rapid vegetation establishment on railway batters. *Journal of Arid Environments* 72(3): 255-262.
18. Debouba M, Kbaier N, Talbi S, Gouia H, Ferchichi A. 2012. Seeds germination and shoot growth responses of a threatened poaceae (*Cenchrus ciliaris* L.) to increasing salt stress. *Rev. Ecology* 67(1): 19-27.
19. Stanisavljevic RS, Vuckovic SM, Simic AS, Markovic JP, Lakic ZP, Terzic DV, Dokic DJ. 2012. Acid and temperature treatments result in increased germination of seeds of three fescue species. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 40(2): 220-226. <https://doi.org/10.15835/nbha4027898>
20. Fox JL, Bhattarai SP, Gyasi-Agyei Y. 2011. Evaluation of different seed mixtures for grass establishment to mitigate soil erosion on steep slopes of railway batters. *Journal of Irrigation and Drainage Engineering* 137(9): 624-631.
21. Burhan N, Shaikat SS. 2000. Effects of atrazine and phenolic compounds on germination and seedling growth of some crop plants. *Pakistan Journal of Biological Sciences* 3(2): 269-274.
22. Shaikat SS, Burhan Nazia. 2000. Fecundity, seed characteristics and factors regulating germination of *Rhynchosia minima* (L.) DC. *Pakistan Journal of Botany* 32(1): 211-226.

23. Ghadiri H, Bagherani TN. 2000. Effects of scarification and temperature on germination of licorice (*Glycyrrhiza glabra* L.) seeds. *Journal of Agri. Sci. Tech.* 2: 257-262.
24. Jaiswal P. 2020. Improving germination of some medicinal and multipurpose species of arid lands through pre-sowing seed treatments. *International Research Journal of Modernization in Engineering Technology and Science* 2(11): 128-140.
25. El-Khalifa MAA. 2004. Effect of seed scarification methods on germination and seed vigor of Poinciana (*Delonix regia*). Doctoral dissertation, UOFK.
26. Baskin JM, Nan X, Baskin CC. 1998. A comparative study of seed dormancy and germination in an annual and a perennial species of Senna (Fabaceae). *Seed Science Research* 8(4): 501-512.
27. Teketay D. 1998. Germination of *Acacia origena*, *A. pilispinaand*, *Pterolobium stellatumin* response to different pre-sowing seed treatments, temperature, and light. *Journal of Arid Environments* 38(4): 551-560.
28. Tobe K, Zhang L, Omasa K. 2005. Seed germination and seedling emergence of three annuals growing on desert sand dunes in China. *Annals of Botany* 95(4): 649-659.
29. Hegarty TW. 1978. The physiology of seed hydration and dehydration, and the relation between water stress and the control of germination: A review. *Plant, Cell and Environment* 1(2): 101-119.
30. Brown NAC, Booysen PDV, De V. 1969. Seed coat impermeability in several *Acacia* species. *Agroplanta* 1: 51-60.
31. Clemens J, Jones PG, Gilbert NH. 1977. Effect of seed treatments on germination in *Acacia*. *Australian Journal of Botany* 25(3): 269-276.
32. Wickens GE. 1969. A Study of *Acacia albida* Del. (Mimosoideae). Kew Bulletin. Published By: Springer. 23(2): 181-202.
33. Wiese AM, Binning LK. 1987. Calculating the threshold temperature of development for weeds. *Weed Science* 35: 177-179.
34. Gomez KA, Gomez AA. 2010. *Statistical Procedures for Agricultural Research*. Wiley India Pvt. Ltd., New Delhi.