

Phosphate Solubilizing Bacteria for a Sustainable Agriculture – A Review

RASHI SUBBA*¹

¹ Department of Botany, Vidyasagar College for Women, Kolkata - 700 006, West Bengal, India

Received: 05 Sep 2023; Revised accepted: 03 Nov 2023; Published online: 24 Nov 2023

Abstract

Phosphorus is a major, essential macronutrient required for plant growth and development. Most soils contain insoluble inorganic phosphates, but they are of no use to plants unless they are solubilized. Therefore, in order to maintain the amount of phosphorus available in soil for plant use, a large amount of phosphorus-based fertilizer is often added to soil, the bulk of which could also be converted to insoluble form. This makes continuous application necessary, which in turn pollutes the soil. To overcome this problem, bacterial inoculants are an important approach that increases plant production for sustainable development. Microorganisms like phosphate-solubilizing bacteria isolated from different plants are found to solubilize the insoluble phosphates. There are different mechanisms involved to solubilize the insoluble phosphate into the soluble form and make it available to the plants, either by lowering the pH, organic acid production, the involvement of enzymes, etc. Application of phosphate-solubilizing bacteria not only solubilizes the insoluble phosphates, but they are also found to increase plant growth. In this review, we have focused on the importance of phosphorus, types of PSB, mechanisms to solubilize insoluble phosphates, and the effect of PSB on plant growth.

Key words: Phosphorus, PSB, pH, Organic acid, Enzymes, Plant growth

Phosphorus is one of the major plant nutrients, second only to nitrogen in requirement and makes up about 0.2% of plant dry weight [1]. It plays an important role in all major metabolic processes in plant including photosynthesis, energy transfer, signal transduction, macromolecular biosynthesis and respiration [2] and nitrogen fixation in legumes [3]. A greater part of soil phosphorus, approximately 95-99% is present in the form of insoluble phosphates and hence cannot be utilized by the plants [4]. Up to 75% of the soluble phosphate fertilizers added to crops may be converted to sparingly soluble forms by reacting with the free Ca^{2+} ions in high pH soils or with Fe^{3+} and Al^{3+} in low pH soils [5-6]. To increase the availability of phosphorus for plants, large amounts of fertilizer are used on a regular basis. But after application, a large proportion of fertilizer phosphorus is quickly transferred to the insoluble form [7]. Therefore, very little percentage of the applied phosphorus is used, making continuous application necessary [8]. Plants can absorb phosphate in two soluble forms, the monobasic (H_2PO_4^-) and the dibasic (HPO_4^{2-}) ions [9]. Interest has been focused on the inoculation of phosphate-solubilizing micro-organisms into the soil so as to increase the availability of native fixed phosphate and to reduce the use of fertilizers [10].

Phosphate solubilising microorganisms are capable of solubilizing tricalcium, aluminium and iron phosphates, as well as rock phosphate making the phosphorus present in the soil available to the plants [11-12]. Soils also contain organic phosphorus, which can be used by crops only if it is mineralized. Organisms that cause increases in plant available phosphate in the soil system belong to a diversified group

including bacteria, actinomycetes and several groups of fungi. The composition and dynamics of this functional group was influenced greatly by vegetation type, soil texture, soil chemical elements, and pH in soil solution [13-15]. It was reported that about 20% of microorganisms in soil can solubilize insoluble inorganic phosphate and that phosphate solubilizing activity of PSM is related to the environmental conditions such as farming practices. Phosphate solubilizing bacteria are common in the rhizosphere and secretion of organic acids and phosphatases are common method of facilitating conversion of insoluble forms of phosphate to plant-available forms [16]. Phosphate solubilizing bacteria have been used to convert insoluble rock phosphate into soluble form and make it available for the plant growth [17-18]. This conversion is through acidification [19-20], chelation and exchange reactions [21-23] and produces, in the periplasm, strong organic acids [24], which have become indicators for routine isolation and selection procedures of phosphate solubilizing bacteria [25].

PSB also produce amino acids, vitamins and growth promoting substances [25-26], which promote plants growth. It was reported that IAA produced by bacteria improves plant growth by increasing the number of root hairs and lateral roots [27]. The production of Indole Acetic Acid (IAA), gibberellins and cytokinins by PSB has been reported earlier by several workers [28]. Many PSB are proved to be effective biofertilizers or bio-controlling agents and can be regarded as broad spectrum biofertilizers [29]. These findings have been further supported by many researchers. Likewise, Chakraborty *et al.* [30] reported that *Bacillus megaterium* promoted the

*Correspondence to: Rashi Subba, E-mail: rashis1402@gmail.com; Tel: +91 9593283019

growth of tea seedlings and two-year old plants significantly, as evidenced by increased plant height, number of branches and leaves.

Phosphate solubilizing bacteria

The immediate vicinity of root surface which constitute the rhizosphere is an extremely important habitat for microbes. Roots secrete a number of compounds into the soil which may either enhance or inhibit the growth of microorganisms [31]. Kobus [32] reported that the numbers of PS bacteria in a soil were influenced more by soil type and the manner of its cultivation than by the physical composition or content of humus, N or P in the soil.

Bacteria are more effective in phosphorus solubilization than fungi [33]. Among the whole microbial population in soil, PSB constitute 1 to 50%, while phosphorus solubilizing fungi (PSF) are only 0.1 to 0.5% in P solubilization potential [34]. Among the soil bacterial communities, ectorhizospheric strains from *Pseudomonas* and *Bacilli*, and endosymbiotic rhizobia have been described as effective phosphate solubilizers [35]. Strains from bacterial genera *Pseudomonas*, *Bacillus*, *Rhizobium* and *Enterobacter* are the most powerful P solubilizers [36] (Whitelaw, 2000). *Bacillus megaterium*, *B. circulans*, *B. subtilis*, *B. polymyxa*, *B. sircalmous*, *Pseudomonas striata*, and *Enterobacter* could be referred as the most

important strains[37-38]. The most important P solubilizing bacterial genera are *Pseudomonas*, *Bacillus*, *Rhizobium*, *Burkholderia*, *Achromobacter*, *Agrobacterium*, *Micrococcus*, *Aereobacter*, *Flavobacterium* and *Erwinia* [39]. There are reports of other bacteria as P solubilizers such as *Azotobacter* [40], *Xanthomonas* [41], *Kurthia* [42] *Rhodococcus*, *Arthrobacter*, *Serratia*, *Chryseobacterium*, *Phyllobacterium*, etc. [43], *Pantoea*, *Klebsiella* [44].

Isolation of phosphate solubilizing bacteria

Ten gram (10 g) of Rhizospheric soil sample was suspended in 90 ml of sterile distilled water and 10^{-1} dilution was obtained. Serial dilutions were prepared by mixing 1 ml of the suspension made into 9 ml sterile water blanks, until the 10^{-7} dilution was obtained. Pikovskaya's agar (10g Glucose, 5g tricalcium phosphate, 0.5g ammonium sulphate, 0.2g potassium sulphate, 0.1 g magnesium sulphate, 0.5 g yeast extract, trace amount of manganese sulphate and ferrous sulphate, 20 g agar, 1000 ml distilled water) medium was used for isolation and maintenance of PSB [45]. The serially diluted soil suspensions were spread-plated on Pikovskaya's agar plates and incubated at 37 °C for 7 days. Bacterial colonies causing clear zones by a turbid white background were considered as phosphate solubilizers. The diameter of PSB colony as well as halo zones were measured by using metric scale.



Fig 1 Bacterial colonies showing clear zones in Pikovskaya's Agar media [46]

Mechanism of phosphate solubilization

The main P solubilization mechanisms employed by soil microorganisms include: (1) release of complex or mineral dissolving compounds e.g., organic acid anions, siderophores, protons, hydroxyl ions, CO_2 ; (2) liberation of extracellular enzymes (biochemical P mineralization) and (3) the release of P during substrate degradation (biological P liberalization) [47]. Thus, microorganisms play an important role in the soil P cycle i.e., dissolution-precipitation, sorption-desorption and mineralization-immobilization.

Several theories exist explaining the mechanisms of microbial P solubilization: the sink theory [48], the organic acid theory [49], and the acidification by H^+ excretion theory [50].

Inorganic P is solubilized by the action of organic and inorganic acids secreted by PSB in which hydroxyl and carboxyl groups of acids chelate cations (Al, Fe, Ca) and decrease the pH in basic soils [51-52]. The PSB dissolve the soil P through production of low molecular weight organic acids mainly gluconic and keto gluconic acids [53-54], in addition to lowering the pH of rhizosphere. Other organic acids that have been involved in the phosphate solubilization are primarily citric, lactic, gluconic, 2-ketogluconic, oxalic, glycolic, acetic, malic, fumaric, succinic, tartaric, malonic, glutaric, pro-pionic, butyric, glyoxylic, and adipic acid [55-60].

Among these organic acids, gluconic acid ranks as the most important in this process and has been studied widely [61-62].

A direct correlation between drops in pH and increase in available P of the culture media has been observed in certain cases [63-64]. In few others, the degree of solubilization was not always proportional to the decline in pH [65-66].

The workers who believe in organic acid theory hardly observed any correlation between the amount of P solubilized and organic acid concentration in the culture medium. Hence it is doubtful as to whether the organic acids are directly and exclusively involved in solubilization [67-68]. Solubilization of calcium phosphate has been reported to occur even in the absence of organic acid [10]. Banik and Dey [69] and Asea *et al.* [67] detected organic acids in culture solutions of PSM but did not show any correlation between the solubilization of P and amount of organic acids produced by PSM. However, an HPLC analysis of the culture solution of *Pseudomonas*, solubilized unavailable forms of P without any organic acid production [25]. In each of these cases, acidification of the medium resulted and it was postulated that H^+ excretion originating from NH_4 assimilation [68] and respiratory H_2CO_3 production [70] as an alternate mechanism of mineral phosphate solubilization. It was hypothesized that the gluconic acid produced lead to the release of protons that finally solubilize the insoluble phosphates [71]. In a study of *Pseudomonas fluorescens*, the form of C supply (e.g., glucose versus fructose) rather than N supply (e.g., NH_4^+ versus NO_3^-) had the greatest effect on proton release [72].

Rudolph [73], reported that MPS activity occurs as a consequence of microbial sulphur oxidation, nitrate production

and CO₂ formation. These processes result in the formation of inorganic acids like sulphuric acid [22]. Further, Azam and Memon [74], supported their findings by saying that, bacteria like *Nitrosomonas* and *Thiobacillus* species can dissolve phosphate compounds by producing inorganic acids such as nitric and sulphuric acids. Inorganic acids e.g., hydrochloric acid can also solubilize phosphate but they are less effective compared to organic acids at the same pH [75]. However, the concept of involvement of inorganic acids in P solubilization are less effective than organic acids [75].

The major source of organic phosphorus in soil is the organic matter. Organic P may constitute 4-90% of the total soil P [76] and soil organic P is largely in the form of inositol phosphate (soil phytate). Other organic P compounds that have been reported are: phosphomonoesters, phosphodiester, phospholipids, nucleic acids and phosphor-triesters [77]. According to the sink theory of Halvorson *et al.* [78]; P solubilizing organisms stimulate the indirect dissolution of Ca-P compounds by continuous removal of P.

Such P can be released from organic compounds in soil by three groups of enzymes: (1) Non-specific acid phosphatases (NSAPs): the most studied among these NSAPs enzymes released by PSM, are the phosphor-monoesterases also referred as phosphatases [79].

(2) Phytases: Another enzyme produced by PSM which is responsible for the release of P from phytate degradation. Phytate in its basic form is the primary source of inositol and are stored in the plant seeds and pollen [80],

(3) Phosphonates and C-P lyases are able to release free P from recalcitrant organic P forms [81].

The overall results of the study indicate that acid production was not the only reason for phosphate solubilization. However, P-solubilization is a complex phenomenon, which depends on many factors such as nutritional, physiological and growth conditions of the culture [82] and in certain cases it is induced by phosphate starvation [83].

Effects of PSB in plant growth

Inappropriate application of mineral fertilizers in agriculture has resulted in pollution and salinization of agricultural lands and water resources. In particular, plant growth-promoting rhizobacteria (PGPR) have been reported to be key elements for plant establishment under nutrient-imbalance conditions. Their use in agriculture can favour a reduction in agro-chemical use and support ecofriendly crop production [84-86]. PGPR can help the improvement of plant growth, plant nutrition, root growth pattern, plant competitiveness, and responses to external stress factors. They can also inhibit soil borne plant pathogens by producing growth-promoting chemical substances and inducing plant resistance [87-89]. Different plant-growth promoting rhizosphere bacteria, including associative bacteria such as *Azospirillum*, *Bacillus*, *Pseudomonas*, *Enterobacter* group have been used for their beneficial effects on plant growth [90]. Several studies clearly showed the effect of plant growth-

promoting bacteria on growth of different crops at different climates, soils and temperatures [91-92].

Phosphate solubilizing microorganisms have an important contribution to overall plant P nutrition and growth, and have increased yields of many crops [93].

Many researchers have reported an increase in P uptake and seed yields, due to PSB inoculation of wheat, barley, mungbean, chickpea and maize genotypes [94-95]. Increased in the plant P uptake and production by 34% in Maize by the application of plant growth-promoting bacteria (PGPB) such as *Azospirillum brasilense*, *Bacillus subtilis*, and *Pseudomonas fluorescens* have been reported by Pereira *et al.* [96]. Phosphate Solubilizing *Pseudomonas* and *Bacillus* species were inoculated into wheat resulting in improved phosphorus uptake and grain production [97]. Similarly, increased plant growth and phosphate uptake have been reported in many crop species as a result of PSB inoculants, e.g., *Pseudomonas* sp. in rice [98], *Pseudomonas* in soya bean [99] and *Pseudomonas* sp. in wheat [100]. *Rhizobium leguminosarum* is of particular interest because of its dual function: its ability to fix N and to solubilize P [101-103]. Inoculation with two strains of P solubilizing *R. leguminosarum* improved root colonization and growth in lettuce and maize. Additionally, rhizobia exhibited an ability to promote plant growth in non-legumes [104]. Increased in plant height, green fodder yield and grain yield of sorghum were reported by the application of PSB based biofertilizer [105].

Indirect growth promotion by PSM is achieved by reducing pathogen infection via the antibiotic or siderophores which are synthesized and supplied by the bacteria [106-107]. A rhizospheric bacterium *Pseudomonas fluorescens*, solubilizes P, and produces antibiotics such as pyoluteorin [108]. Hydrogen cyanide produced by *Pseudomonas* was used as a biological control of black root rot of tobacco [106].

Similarly, a number of *Pseudomonas* strains are found to be well adapted to higher altitude soils and have exhibited antifungal, phosphate solubilizing, and plant growth promoting properties [109]. These species have been reported as efficient degraders of organic matter [110-111].

CONCLUSION

Out of nitrogen, phosphorus and potassium, phosphorus is the second essential macronutrient for plant growth and development. In spite of its presence in soil in large quantities, it is not easily available to the plants because of its fixation, which makes them insoluble. Due to the continuous application of chemical fertilizer to the soil, it has become necessary to find an alternative for the sustainable agriculture. Phosphate solubilizing bacteria present in the soil solubilize the insoluble phosphates and increase the plant yield. These properties of PSB are under consideration for use as a biofertilizer. However, further investigations are required to develop PSB as biofertilizers isolated from different crops, different geographical regions for better crop productivity and to reduce environmental pollution to promote sustainable agriculture.

LITERATURE CITED

- Schachtman DP, Reid RJ, Ayling SM. 1998. Phosphorus uptake by plants: From soil to cell. *Plant Physiology* 116: 447-453.
- Khan MS, Zaidi A, Ahemad M, Oves M, Wani PA. 2010. Plant growth promotion by phosphate solubilizing fungi – current perspective. *Arch. Agron. Soil Science* 56: 73-98.
- Saber K, Nahla LD, Ahmed D, Chedly A. 2005. Effect of P on nodule formation and N fixation in bean. *Agron. Sustain. Dev.* 25: 389-393.
- Vassileva M, Vassilev N, Azcon R. 1998. Rock phosphate solubilization by *Aspergillus niger* on olive cake-based medium and its further application in a soil-plant system. *World Journal of Microbiol. Biotech.* 14: 281-284.

5. Goldstein AH. 1986. Bacterial solubilization of microbial phosphates: Historical perspective and future prospects. *Am. Jr. Alter. Agric.* 1: 51-57.
6. Arpana N, Kumar SD, Prasad TN. 2002. Effect of seed inoculation, fertility and irrigation on uptake of major nutrients and soil fertility status after harvest of late sown lentil. *Jr. App. Bio.* 12: 23-26.
7. Omar SA. 1998. The role of rock-phosphate-solubilizing fungi and vesicular arbuscular mycorrhiza (VAM) in growth of wheat plants fertilized with rock phosphate. *World Journal of Microbiol. Biotech.* 14: 211-218.
8. Abd Alla MH. 1994. Phosphatases and the utilization of organic phosphorus by *Rhizobium leguminosarum* biovar *viceae*. *Lett. Appl. Microbiology* 18: 294-296.
9. Glass ADM. 1989. *Plant Nutrition: An Introduction to Current Concepts*. Jones and Barlett Publishers, Boston, MA, 1994. USA. pp 234.
10. Illmer P, Schinner F. 1992. Solubilization of inorganic phosphates by microorganisms isolated from forest soil. *Soil Biol. Biochemistry* 24: 389-395.
11. Katznelson H, Bose B. 1959. Metabolic activity and phosphate-dissolving capability of bacterial isolates from wheat roots, rhizosphere and non-rhizosphere soil. *Can. Jr. Microbiology* 5: 79-82.
12. Katznelson H, Peterson EA, Rovatt JW. 1962. Phosphate dissolving microorganism in seed and in the root zone of plants. *Can. Jr. Botany* 40: 1181-1186.
13. Curl EA, Bonner DF, Sabey BR. 1986. *The Rhizosphere*. Belin. Springer-Verlag. pp 167-175.
14. Kucey RMN. 1983. Phosphate-solubilizing bacteria and fungi in various cultivated and virgin Alberta soils. *Can. Jr. Soil Science* 63: 671-678.
15. Lin QM, Zhao XR, Sun YX, Yao J. 2000. Community characters of soil phosphobacteria in four ecosystems. *Soil Env. Science* 9: 34-37.
16. Kim KY, Jordan D, McDonald GA. 1998. Effect of phosphate solubilizing bacteria and vesicular-arbuscular mycorrhizae on tomato growth and soil microbial activity. *Biol. Fert. Soils* 26: 79-87.
17. Nahas E, Banzatto DA, Assis LC. 1990. Fluorapatite solubilization by *Aspergillus niger* in vinasse medium. *Soil Biol. Biochemistry* 22: 1097-1101.
18. Bojinova D, Velkova R, Grancharov I, Zhelev S. 1997. The bioconversion of Tunician phosphate using *Aspergillus niger*. *Nutr. Cyc. Agroecosyst.* 47: 227-232.
19. Muromtsev GS. 1958. The dissolving action of some root and soil microorganisms on calcium phosphates insoluble in water. *Agrobiolog.* 5: 9-14.
20. Louw HA, Webley DM. 1959. A study of soil bacteria dissolving certain phosphate fertilizers and related compounds. *Jr. Appl. Bacteriology* 22: 227-233.
21. Duff RB, Webley DM. 1959. 2-ketogluconic acid as a natural chelator produced by soil bacteria. *Chem. and Ind.* pp 1376-1377.
22. Sperber JL. 1958b. Solution of apatite by soil microorganisms producing organic acids. *Australian Jr. Agricultural Research* 9: 782-787.
23. Gerke J. 1992. Phosphate, aluminium, and iron in the soil solution of three different soils in relation to varying concentration of citric acid. *Jr. Soil Sci. Plant Nutrition* 155: 339-343.
24. Alexander M. 1977. *Introduction to Soil Microbiology*. John Wiley and Sons Inc., New York, USA.
25. Gonzalez J, Salmeron V, Moreno J, Cornmenzana AR. 1983. Amino acids and vitamins produced by *Azotobacter vinelandii* ATCC 12837 in chemically-defined media and dialyzed soil media. *Soil Biol. Biochemistry* 15: 711-713.
26. Zimmer W, Roeben K, Bothe H. 1988. An alternative explanation for plant growth promotion by bacteria of the genus *Azospirillum*. *Planta* 176: 333-342.
27. Okon Y, Kapulnik Y. 1986. Development and function of *Azospirillum*-inoculated roots. *Plant Soil* 90: 3-16.
28. Khalid A, Arshad M, Zahir ZA. 2004. Screening plant growth promoting rhizobacteria for improving growth and yield of wheat. *Jr. Appl Microbiology* 96: 473-480.
29. Gupta AK. 2004. *The Complete Technology Book on Biofertilizers and Organic Farming*. National Institute of Industrial Research Press. India
30. Chakraborty U, Chakraborty BN, Basnet M. 2009. Exploitation of tea rhizosphere microorganisms for improvement of plant health status. *Jr. Mycol. Pl. Pathology* 39: 1-13.
31. Chakraborty U, Chakraborty BN. 1997. Phyllosphere and rhizosphere microorganisms of *Camellia sinensis* grown in the Eastern Himalayan Regions. In: *Recent researches in Ecology, Environment and Pollution* (Vol.10). (Eds) S. C. Sati, J. Saxena and R.C. Dubey. Today and Tomorrow's Printers and Publishers, New Delhi, India. pp 189-203.
32. Kobus J. 1962. The distribution of microorganisms mobilizing phosphorus in different soils. *Acta Microbiologica Polonica* 11: 255-262.
33. Alam S, Khalil S, Ayub N, Rashid M. 2002. *In vitro* solubilization of inorganic phosphate by phosphate solubilizing microorganism (PSM) from maize rhizosphere. *Int. Jr. Agric. Biol.* 4: 454-458
34. Chen YP, Rekha PD, Arun AB, Shen FT, Lai WA, Young CC. 2006. Phosphate solubilizing bacteria from subtropical soil and their tricalcium phosphate solubilizing abilities. *Appl. Soil Ecol.* 34: 33-41.
35. Igual JM, Valverde A, Cervantes E, Velazquez E. 2001. Phosphate solubilizing bacteria as inoculants for agriculture use of updated molecular techniques in their study. *Agronomie.* 21: 561-568.
36. Whitelaw MA. 2000. Growth promotion of plants inoculated with phosphate-solubilizing fungi. *Advanced Agronomy* 69: 99-151.
37. Subbarao NS. 1988. Phosphate solubilizing micro-organism. In: *Biofertilizer in agriculture and forestry*. Regional Biofert. Dev. Centre, Hissar, India. pp 133-142.
38. Kucey RMN, Janzen HH, Leggett ME. 1989. Microbiologically mediated increases in plant-available-phosphorus. *Advanced Agronomy* 42: 199-228.

39. Rodriguez H, Fraga R. 1999. Phosphate solubilising bacteria and their role in plant growth promotion. *Biotech. Advance* 17: 319-339.
40. Kumar V, Behl RK, Narula N. 2001. Establishment of phosphate-solubilizing strains of *Azotobacter chroococcum* in the rhizosphere and their effect on wheat cultivars under greenhouse conditions. *Microbiol. Research* 156: 87-93.
41. DeFreitas JR, Banerjee MR, Germida JJ. 1997. Phosphate-solubilizing rhizobacteria enhance the growth and yield but not phosphorus uptake of canola (*Brassica napus* L.). *Biol. Fertil. Soils* 24: 358-364.
42. Sharma BC, Subba R, Saha A. 2012. *Kurthia* sp. a novel member of phosphate solubilizing bacteria from rhizospheric tea soil of Darjeeling Hills, India. *IOSR Jr. Pharm. Biol. Sci.* (IOSRJPBS) 2(3): 36-39.
43. Wani PA, Zaidi A, Khan AA, Khan MS. 2005. Effect of phorate on phosphate solubilization and indole acetic acid (IAA) releasing potentials of rhizospheric microorganisms. *Annals Plant Protection Science* 13: 139-144.
44. Chung H, Park M, Madhaiyan M, Seshadri S, Song J, Cho H, Sa T. 2005. Isolation and characterization of phosphate solubilizing bacteria from the rhizosphere of crop plants of Korea. *Soil Biol. Biochemistry* 37: 1970-1974.
45. Pikovskaya RI. 1948. Mobilization of phosphorus in soil connection with the vital activity of some microbial species. *Microbiology* 17: 362-370.
46. Subba R. 2013. Isolation and characterization of phosphate solubilizing microbes from Darjeeling soils for their use as potential inoculants in upland farming systems. *Ph. D. Thesis*, University of North Bengal.
47. McGill WB, Cole CV. 1981. Comparative aspects of cycling of organic C, N, S and P through soil organic matter. *Geoderma* 26: 267-286.
48. Halvorson HO, Keynan A, Kornberg HL. 1990. Utilization of calcium phosphates for microbial growth at alkaline pH. *Soil Biol. Biochemistry* 22: 887-890.
49. Cunningham JE, Kuiack C. 1992. Production of citric and oxalic acids and solubilization of calcium phosphate by *Penicillium bilaii*. *Appl. Environ. Microbiology* 58: 1451-1458.
50. Illmer P, Schinner F. 1995. Solubilisation of inorganic calcium phosphates – Solubilization mechanisms. *Soil Biol. Biochemistry* 27: 257-263.
51. Kpombrekou AK, Tabatabai MA. 1994. Effect of organic acids on release of phosphorus from phosphate rocks. *Soil Science* 158(6): 442-453.
52. Stevenson FJ. 2005. The Phosphorus Cycle. *In: Cycles of Soil: Carbon, Nitrogen, Phosphorus, Sulfur, Micronutrients*. John Wiley and Sons, New York. pp 231-284.
53. Goldstein AH. 1995. Recent progress in understanding the molecular genetics and biochemistry of calcium phosphate solubilization by Gram negative bacteria. *Biol. Agric. Hort.* 12: 185-193.
54. Deubel A, Gransee A, Merbach W. 2000. Transformation of organic rhizodeposits by rhizoplane bacteria and its influence on the availability of tertiary calcium phosphate. *Jr. Plant Nutr. Soil Science* 163: 387-392.
55. Kumar A, Kumar A, Patel H. 2018. Role of microbes in phosphorus availability and acquisition by plants. *Int. Jr. of Cur. Microbiol. Appl. Sci.* 7(5): 1344-1347.
56. Satyaprakash M, Nikitha T, Reddi EUB, Sadhana B, Vani SS. 2017. A review on “Phosphorous and phosphate solubilizing bacteria and their role in plant nutrition. *Int. Jr. Cur. Microbiol. Appl. Science* 6: 2133-2144.
57. Walpola BC, Yoon MH. 2012. Prospectus of phosphate solubilizing microorganisms and phosphorus availability in agricultural soils: A review. *Afr. Jr. Microbiol. Research* 6: 6600-6605.
58. Selvi KB, Paul JJA, Vijaya V, Saraswathi K. 2017. Analyzing the efficacy of phosphate solubilizing microorganisms by enrichment culture techniques. *Biochem. Mol. Biol. Journal* 3: 1-7.
59. Yousefi A, Khavazi K, Moezi A, Rejali F, Nadian H. 2011. Phosphate solubilizing bacteria and arbuscular mycorrhizal fungi impacts on inorganic phosphorus fractions and wheat growth. *World Appl. Sci. Jr.* 15: 1310-1318.
60. Ahmed N and Shahab S. 2011. Phosphate solubilization: Their mechanism, genetics and application. *Int. Jr. Microbiology* 9: 4408-4412.
61. Rodriguez H, Fraga R, Gonzalez T, Bashan Y. 2006. Genetics of phosphate solubilization and its potential applications for improving plant growth-promoting bacteria. *Plant Soil* 287: 15-21.
62. Alori ET, Glick BR, Babalola OO. 2017. Microbial phosphorus solubilization and its potential for use in sustainable agriculture. *Front Microbiology* 8: 971.
63. Sperber JJ. 1958. The incidence of apatite-solubilizing organisms in the rhizosphere and soil. *Australian Jr. Agric. Research* 9: 778-781.
64. Agnihotri VP. 1970. Solubilization of insoluble phosphates by soil fungi isolated from nursery seed beds. *Can. Jr. Microbiology* 16: 877-880.
65. Mehta YR, Bhide VP. 1970. Solubilization of tricalcium phosphate by some soil fungi. *Indian Jr. Exp. Boil.* 8: 228-229.
66. Krishnaraj PU. 1987. Studies on beneficial microorganisms in crop plants. *M. Sc. (Agri) Thesis*, UAS Bangalore.
67. Asea PEA, Kucey RMN, Stewart JWB. 1988. Inorganic phosphate solubilization by two *Penicillium* species in solution culture and soil. *Soil Biol. Biochemistry* 20: 459-464.
68. Parks EJ, Olson GJ, Brinckman FE, Baldi F. 1990. Characterization by high performance liquid chromatography (HPLC) of solubilization of phosphorus in iron ore by a fungus. *Jr. Ind. Microbiology* 5: 183-189.
69. Banik S, Dey BK. 1983. Alluvial soil microorganisms capable of utilizing insoluble aluminium phosphate as a sole source of phosphorus. *Zentralblatt Microbiology* 138: 437-442.
70. Jurinak JJ, Dudley LM, Allen MF, Knight WG. 1986. The role of calcium oxalate in the availability of phosphorus in soils of semiarid regions: A thermodynamic study. *Soil Science* 142(5): 255-261.
71. Goldstein AH. 1994. Involvement of the quinoprotein glucose dehydrogenase in the solubilization of exogenous phosphates by gram-negative bacteria. *In: Phosphate in Microorganisms: Cellular and molecular biology*. (Eds) Torriani-Gorini A Yagil E. Silver S. ASM Press, Washington, DC. pp 197-203.

72. Park KH, Lee CY, Son HJ. 2009. Mechanism of insoluble phosphate solubilization by *Pseudomonas fluorescens* RAF15 isolated from ginseng rhizosphere and its plant growth-promoting activities. *Letters Appl. Microbiology* 49: 222-228.
73. Rudolph W. 1922. Influence of S oxidation upon growth of soybeans and its effect on bacterial flora of soil. *Soil Science* 14: 247-263.
74. Azam F, Memon GH. 1996. *Soil Organisms*. In: (Eds) Bashir E, Bantel R. Soil science. National Book Foundation, Islamabad. pp 200-232.
75. Kim KY, McDonald GA, Jordan D. 1997. Solubilization of hydroxyapatite by *Enterobacter agglomerans* and cloned *Escherichia coli* in culture medium. *Biol. Fertil. Soils* 24: 347-352.
76. Khan AA, Jilani G, Akhtar MS, Naqvi SMS, Rasheed M. 2009. Phosphorus solubilizing bacteria, occurrence, mechanisms and their role in crop production. *Jr. Agric. Biol. Sci.* 1: 48-58.
79. Nannipieri P, Giagnoni L, Landi L, Renella G. 2011. Role of phosphatase enzymes in soil. In: Phosphorus in Action: Biological Processes in Soil Phosphorus Cycling. Soil Biology, eds E Bunemann, A. Oberson, And E. Frossard (Berlin: Springer). pp 215-243.
80. Richardson AE. 1994. Soil microorganisms and phosphorus availability. In: (Eds) Pankhurst CE, Doubeand BM, Gupta VVSR. Soil biota: management in sustainable farming systems. CSIRO, Victoria, Australia. pp 50-62.
81. Richardson AE, Simpson RJ. 2011. Soil microorganisms mediating phosphorus availability. *Plant Physiology* 156: 989-996.
83. Reyes I, Bernier L, Simard RR, Antoun H. 1999. Effect of nitrogen source on solubilization of different inorganic phosphate by an isolate of *Penicillium rugulosum* and two UV-induced mutants. *FEMS Microbiol. Ecol.* 28: 281-290.
83. Gyaneshwar P, Parekh LJ, Archana G, Poole PS, Collins MD, Hutson RA, Naresh KG. 1999. Involvement of a phosphate starvation inducible glucose dehydrogenase in soil phosphate solubilization by *Enterobacter asburiae*. *FEMS Microbiol. Letters* 171: 223-229.
84. Herrera MA, Salamanca CP, Barea JM. 1993. Inoculation of woody legumes with selected arbuscular mycorrhizal fungi and rhizobia to recover desertified mediterranean ecosystems. *Appl. Environ. Microbiology* 59: 129-133.
85. Requena N, Jimenez I, Toro M, Barea JM. 1997. Interactions between plant-growth promoting bacteria (PGPR), arbuscular mycorrhizal fungi and *Rhizobium* spp. In: The rhizosphere of *Anthyllis cytisoides*, a model legume for revegetation in Mediterranean semi-arid ecosystems. *New Phytology* 136: 667-677.
86. Glick BR. 1995. The enhancement of plant growth by free-living bacteria. *Can. Jr. Microbiology* 41: 109-117.
87. Lifshitz R, Kloepper JW, Kozlowski M, Simonson C, Carlson J, Tipping M, Zaleska I. 1987. Growth promotion of canola (rapeseed) seedlings by a strain of *Pseudomonas putida* under gnotobiotic conditions. *Can. Jr. Microbiology* 33: 390-395.
88. Bothe H, Korsgen H, Lehmacher T, Hundeshagen B. 1992. Differential effects of *Azospirillum*, auxin and combined nitrogen on growth of the roots of wheat. *Symbiosis* 13: 167-179.
89. Hoflich G, Wiehe W, Kuhn G. 1994. Plant growth stimulation with symbiotic and associative rhizosphere microorganisms. *Experientia* 50: 897-905.
90. Kloepper JW, Beauchamp CJ. 1992. A Review of issues related to measuring colonization of plant roots by bacteria. *Can. Jr. Microbiology* 38(12): 1219-1232.
91. Dobereiner J. 1992. Recent changes in concepts of plant bacteria interactions, endophytic N₂-fixing bacteria. *Ciencia e Cultura*. 44(5): 310-313.
92. Boelens J, Zoutmann D, Cambell J, Verstraete W, Paranchych W. 1993. The use of bioluminescence as a reporter to study the adherence of the plant growth promoting rhizospseudomonas 7NSK2 and ANP15 to canola roots. *Can. Jr. Microbiology* 39: 329-334.
93. Leggett ME, Gleddie SC, Holloway G. 2001. Phosphate-solubilizing microorganisms and their use. In: Plant nutrition acquisition: New perspectives (Eds) N. Ae, J. Arihara, K. Okada, and A. Srinivasan, Springer-Verlag, Tokyo. Lima. pp 299-318.
94. Singh S, Kapoor KK. 1999. Inoculation with phosphate solubilizing microorganisms and a vesicular-arbuscular mycorrhizal fungus improves dry matter yield and nutrient uptake by wheat grown in sandy soil. *Biol. Fertil. Soils*. 28: 139-144.
95. Ramirez R, Fernandez SM, Lizaso JI. 2001. Changes of pH and available phosphorus and calcium in rhizosphere of aluminium-tolerant maize germplasm fertilized with phosphate rock. *Commun. Soil Sci. Plant Anal.* 32: 1551-1565.
96. Pereira NCM, Galindo FS, Gazola RPD, Dupas E, Rosa PAL, Mortinho ES. 2020. Corn yield and phosphorus use efficiency response to phosphorus rates associated with plant growth promoting bacteria. *Front. Environ. Sci.* 8: 1-12.
97. Afzal A, Ashraf A, Saeed A, Asad A, Farooq M. 2005. Effect of phosphate solubilizing microorganisms on phosphorus uptake, yield and yield traits of wheat (*Triticum aestivum* L.) in rainfed area. *Int. Jr. Agric. Biol.* 7: 1560-8530.
98. Gusain YS, Kamal R, Mehta CM, Singh US, Sharma AK. 2015. Phosphate solubilizing and indole-3-acetic acid producing bacteria from the soil of Garhwal Himalaya aimed to improve the growth of rice. *Jr. Environ. Biology* 36: 301-30.
99. Fankem H, Tchakounte GVT, Nkot NL, Mafokoua HL, Dondjou DT, Simo C, Nwaga D, Etoa FX. 2015. Common bean (*Phaseolus vulgaris* L.) and soya bean (*Glycine max*) growth and nodulation as influenced by rock phosphate solubilizing bacteria under pot grown conditions. *Int. Jr. Agric. Policy Reseach* 3(5): 242-250.
100. Babana A, Antoun H. 2006. Effect of Tilemsi phosphate rock-solubilizing microorganisms on phosphorus uptake and yield of field-grown wheat (*Triticum aestivum* L.) in Mali. *Plant Soil* 287: 51-58.
101. Wood M, Cooper JE. 1984. Aluminium toxicity and multiplication of *Rhizobium trifolii* in a defined growth medium. *Soil Biol. Biochemistry* 16: 571-576.
102. Chabot R, Antoun H, Kloepper JW, Beauchamp CJ. 1996. Root colonization of maize and lettuce by bioluminescent *Rhizobium leguminosarum* biovar *phaseoli*. *Appl. Enviro. Microbiology* 62: 2767-2772.
103. Hara F, DeOliveira LA. 2004. Physiological and ecological characteristics of rhizobio isolated deriving of acid and alic soils of Presidente Figueiredo, Amazonas State. *Acta Amazonica* 34(3): 343-357.

104. Chabot R, Beauchamp CJ, Kloepper JW, Antoun H. 1998. Effect of phosphorus on root colonization and growth of maize by bioluminescent mutants of phosphate-solubilizing *Rhizobium leguminosarum* biovar *phaseoli*. *Soil Biol. Biochemistr* 30: 1615-161.
105. Akhtar S, Bashir S, Khan S, Iqbal J, Gulshan AB, Irshad S. 2020. Integrated usage of the synthetic and bio-fertilizers: an environment friendly approach to improve the productivity of sorghum. *Cereal Res. Communication* 48: 247-253.
106. Antoun H, Beauchamp CL, Goussard N, Chabot R, Roger L. 1998. Potential of *Rhizobium* and *Bradyrhizobium* species as plant growth promoting rhizobacteria on non-legumes: Effect on radishes (*Raphanus sativus* L.). *Plant and Soil* 204: 57-67.
107. Rosas SB, Andres JA, Rovera M, Correa N. 2006. Phosphate-solubilizing *Pseudomonas putida* can influence the rhizobia-legume symbiosis. *Soil Biol. Biochemistry* 38: 3502-3505.
108. Trujillo ME, Velazquez E, Miguelez S, Jimenez MS, Mateos PF, Martinez-Molina E. 2003. Characterization of a strain of *Pseudomonas fluorescens* that solubilize phosphates *in vitro* and produces high antibiotic activity against several microorganisms. In: E. Velazquez (Eds) First International Meeting on Microbial Phosphate Solubilization, 16-19 July 2002, Salamanca, Spain. pp 265-268.
109. Pandey A, Palni LMS. 1998. Microbes in Himalayan soils: Biodiversity and potential applications. *Journal of Science Ind. Research* 57: 668-673.
110. Singh M, Malik RK. 1993. Isolation of few lignocellulose degrading fungi. *Ind. Jr. Microbiology* 33: 265-267.
111. Bhardwaj KKR. 1995. Improvements in microbial compost technology: A special reference to microbiology of composting. In: Wealth from wastes (Eds) S. Khanna and Krishna Mohan). TERI Publications, New Delhi. pp 115-135.