

Evaluation of Anti-hyperglycemic Potential of an Ethno Medicinal Plant *Grewia bracteata* B. Heyne ex Roth

K. Usha*¹, T. Francis Xavier² and R. Sabitha³

¹⁻³ Ethnopharmacological Research Unit, PG and Research Department of Botany, St. Joseph's College (Autonomous), (Affiliated to Bharathidasan University, Tiruchirappalli), Tiruchirappalli - 620 002, Tamil Nadu, India

Received: 24 Jul 2023; Revised accepted: 14 Jan 2024; Published online: 02 Feb 2024

Abstract

Grewia bracteata is an ethno medicinal shrub used by many traditional users for various illnesses. The phytochemicals present in *Grewia bracteata* ethyl acetate, ethanol and chloroform solvent extracts are screened by standard methods and its anti-hyperglycemic activity by enzymatic methods at various concentrations (59.37, 118.75, 237.5, 475, 950 µg/ml). The phytochemical analysis exhibited the various secondary metabolites like alkaloids, flavonoids, saponins, terpenoids, tannins, steroids and glycosides are present in *Grewia bracteata*. Both the enzymatic assays for anti-hyperglycemic assay exhibits a significant inhibitory effect at highest concentration of 950 µg/ml in all the solvents used. The ethanol extract exhibited the strongest activity in both the assays α -amylase and α -glucosidase with a percentage inhibition of 85.2 ± 0.27 and 55.46 ± 0.01 (IC₅₀, 116.24 \pm 1.16 and 400.03 \pm 1.09 µg/ml) respectively followed by ethyl acetate and chloroform. In Conclusion, this study has revealed that *Grewia bracteata* leaves possess noticeable in vitro α -amylase and α -glucosidase inhibitory activities due to the presence of its phytochemicals.

Key words: Acarbose, Anti-hyperglycemic activity, *Grewia bracteata*, Organic solvents, Phytochemicals

Plants are integral part of human civilization, which have one or more of its organs containing substances that can be used for the therapeutic purpose. The evolution of human civilization has been greatly influenced by medicinal plants [4]. Many contemporary medications are made directly from plants, which are also the basis of many ancient remedies [6]. The effects of medicinal plants have drawn more attention within the last ten years. Conventional herbal remedies originating from many ecosystems and regions may offer a fresh approach to wound care and infection prevention [5]. In recent times, focus on plant research has increased all over the world and a large body of evidence was collected to show immense potential of medicinal plants used in various traditional systems [7]. Because, the herbal preparations are complementary and alternative medicine (CAM) and the search for the discovery of novel compounds derived from natural sources like herbs or plants is growing mainly due to acquired resistance, side effects, and adverse events (AE) of allopathic medication [1], [15]. Diabetic mellitus (DM) is the condition arising by insulin deficiency, often combined with insulin resistance. It is because of the disruption of a metabolic system of carbohydrates, proteins and fats resulting in complications of the kidneys, eyes and cardiovascular system [13-14]. World Health Organization has also reported that about 171 million people worldwide were suffering from diabetes mellitus in the year 2000, while predicting that this figure will double come the year 2030. This report was corroborated by Wild et al. Diabetic type 2 (non-insulin-dependent diabetes mellitus) is most dominant; almost 90% of all diabetic cases belong to this type in the world [8].

The hypoglycemic effect of several plant extracts and herbal formulations have been confirmed which are being used as antidiabetic remedies and their therapeutic capabilities are investigated intensively. The utilization of traditional herbal medicine to treat diabetes has been accepted by the people who live in the rural area or in the urban/close to modern medical center. The use of medicinal plants reported to be cost effective anti-diabetic agents with fewer reported side effects. Especially, the price is lower for patients who cannot afford to buy medicine at higher price [2]. Medicinal plants have always been an exemplary source of drugs and which are particularly used by the traditional users since the ancient time but they do not have much scientific data [10]. Hence considering the mentioned points, the present study was therefore undertaken to investigate the antidiabetic activity of the ethnomedicinal plant *Grewia bracteata*.

MATERIALS AND METHODS

Grewia bracteata is Wight's Gross berry, a shrub and which is found in dry deciduous forests in Andhra Pradesh, Karnataka, Tamil Nadu, Kerala and Sri Lanka. The plant sample was collected and identified by Rapinat Herbarium, Center for molecular systematics, St. Joseph's College, Tiruchirappalli.

Extraction of plant material

The plant material was thoroughly washed and well dried in shade. Then the dried plant material was ground to the fine

*Correspondence to: K. Usha, E-mail: ushakanandan@gmail.com; Tel: +91 9791417299

powder, which is used to make extraction by using soxhlet apparatus. 20 g leaf powder has soaked separately in 250 mL of solvents such as ethyl acetate, ethanol, and chloroform for 24 h. Next to that, the extract of ethyl acetate, ethanol, and

chloroform was concentrated in a rotary vacuum evaporator (Evator) under reduced pressure at 40 °C until solid residues were obtained. Then the dried extracts were collected and stored at 20 °C use for in further studies.

Table 1 Phytochemicals present in *Grewia bracteata* leaf extracts

Phyto constituents	Chemical tests	Ethyl acetate	Ethanol	Chloroform
Carbohydrates	Molisch's test (2-3ml extract + Naphthol + Sulphuric acid → Violet colour ring)	+	+	+
Proteins	Million's test (2ml extract + Million's reagent → Red colour precipitation formed)	+	+	+
Amino acids	Ninhydrin test (Filterate + lead acetate solution + Spotted + ninhydrin spray → violet spots appeared)	+	+	+
Steroids	Salkowski test (Extract, chloroform and con. H ₂ SO ₄ (2 ml of each) → shaking for 5 minutes → stand for 5 minutes → Red colour layer formed)	+	+	-
Alkaloids	Mayer's test (Extract + Mayer's reagent → coloured creamy precipitate formed)	+	+	+
Phenolic compounds	Lead acetate test (Extract + few drops lead acetate solution → Formation of white precipitate)	+	+	-
Saponins	Foam test (2ml extract + diluted dist. H ₂ O 20ml equivalent + stirred for 20 minutes → formation of foam layer)	-	-	-
Glycosides	Keller-kilani test (2ml extract + 1ml glacial acetic acid + 1 drop 5% ferric chloride + 1ml Conc. HCl → Reddish brown colour appeared)	-	-	-
Tannins	Gelatin test (extract 2ml + gelatin solution + shake well → white precipitate formed)	+	-	+

Preliminary phytochemical analysis

The phytochemicals present in the yield of crude extracts were examined by using various biochemicals by standard techniques [16] (Table 1). Three solvent extracts viz. ethyl acetate ethanol and chloroform extracts of *G.bracteata* were used for the anti-hyperglycemic studies.

α -amylase inhibitory activity

The α -amylase with the *Grewia bracteata* leaf extracts at various concentrations (59. 37- 950 μ g/ mL) was premixed 0.5 % starch (substrate) was added to start the reaction. At 37 °C the reaction was incubated for 5 min and terminated by the addition of 2mL of DNS (3, 5- dinitrosalicylic acid) reagent and incubated. The reaction mixture was heated at 100 °C for 15 min and diluted with 10 ml of distilled water in an ice bath [18]. By measuring the spectrum at 540nm the α -amylase inhibitory activity was measured and the following formula was used for calculating the α -amylase inhibitory activity:

$$\% \text{ Inhibition} = \frac{[(\text{Abs Control} - \text{Abs Samples}) / \text{Abs Control}] \times 100}{100}$$

α -glucosidase inhibitory activity

The *Grewia bracteata* leaf extracts at various concentrations (50-200 μ g/ mL) was premixed with α -glucosidase. As a substrate 3mM p-nitrophenyl glucopyranoside was added (pNPG) to start the reaction [18]. Incubated the reaction mixture for 30 min at 37 °C and stop by added 2 mL of Na₂CO₃. P-nitrophenol released from pNPG was measured at 400 nm for determined the α -glucosidase activity and the inhibitory activity was calculated by the following formula:

$$\% \text{ Inhibition} = \frac{[(\text{Abs Control} - \text{Abs Samples}) / \text{Abs Control}] \times 100}{100}$$

RESULTS AND DISCUSSION

Preliminary phytochemical analysis

Phytochemicals are non-nutritive plant chemicals with protective or disease-prevention properties. Such compounds are found in plants to protect themselves, but current research suggests that they can also defend humans and animals from many diseases and infections [11]. The inhibitory activity of plant extract may vary according to the nature of active ingredients in the plant. Successful prediction of bioactive components from plant materials is largely dependent on the type of solvents used in the extraction procedure [9]. The phytochemicals present in *Grewia bracteata* leaf extracts (Ethyl acetate, ethanol and chloroform) were determined by various tests. The results revealed that the presence of carbohydrates, proteins, alkaloids, flavonoids, tannins, terpenoids, phenolic compounds and glycosides were present (Table 1).

Anti-diabetic activity

In vitro anti-diabetic activity of *G. bracteata* leaf extracts were assessed by the enzymatic method viz. α -amylase and α -glucosidase assays by the Xavier *et al.* [18] protocol.

α - Amylase inhibition activity

In vitro assay of α - Amylase inhibition activity was performed to determine the anti-diabetic activity of the *G. bracteata* (Fig 1). Here, α -amylase enzyme converts starch into glucose. When α -amylase, glucose and plant extract are taken together as a solution, the extract causes inhibition of enzyme activity [17]. The present study reveals that the inhibitory effect of α -amylase on *G. bracteata* ethanol extract showed the highest percentage of inhibition (85.2 \pm 0.27) at 950 μ g/ml concentration followed by ethyl acetate extract (82.92 \pm 0.15) at same concentration (Table 2). The IC₅₀ values of ethanol, ethyl acetate and chloroform are 116.24 \pm 1.16, 244.02 \pm 0.57 and

309.84±0.67 µg/ml. which is defined as the sample concentration required to triggering 50% inhibition in which the higher inhibition was noted as lower IC₅₀ value (Table 4). While (Fig 1) showed *Grewia bracteata* extracts exhibits comparatively similar level of inhibitory effect with standard

Acarbose. Whereas, Kumar *et al.* [12] showed that the best inhibition activity of *Solanum torvum* and *Solanum trilobatum* obtained using the water extract of these plants. When, the concentration increased the percentage of inhibition of α-amylase is also increased.

Table 2 α-amylase activity of *Grewia bracteata* in percentage

Concentration (ug/ml)	Ethanol	Ethyl acetate	Chloroform	Acarbose
59.37	39.17±0.27	36.25±0.41	30.95±0.27	44.47±0.41
118.75	52.69±0.41	44.38±0.27	38.08±0.27	51.59±0.15
237.5	60.82±0.27	57.07±0.15	49.4±0.15	57.80±0.27
475	69.22±0.15	63.92±0.31	66.11±0.31	70.95±0.27
950	85.2±0.27	82.92±0.15	81.55±0.15	83.01±0.27

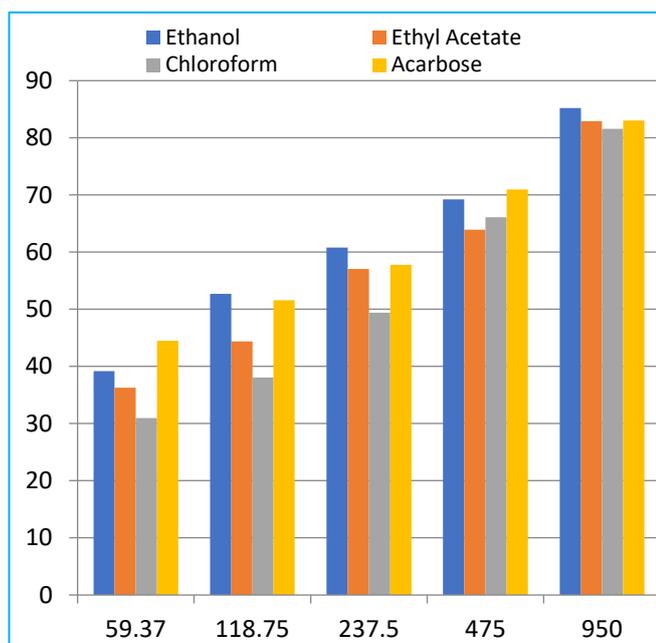


Fig 1 α-amylase activity of *Grewia bracteata* in percentage

α - Glucosidase activity

In vitro assay of α- glucosidase inhibition activity was performed to determine the anti-diabetic activity of the *Grewia bracteata*. The results of α - glucosidase inhibitory activity on ethyl acetate, ethanol and chloroform extract was mentioned with IC₅₀ of 400.03±1.09, 455.49±1.38 and 495.72±1.67 µg/ml respectively (Table 3). Here, α - glucosidase inhibitory effect is lower than the standard Acarbose with IC₅₀ of 88.90±1.42 µg/ml (Fig 3). Comparatively, Egharevba *et al.* (2019) reported the ability of *G. bracteolata* to inhibit α - glucosidase activity which is similar to the present findings. Likewise, the percentage of α - amylase activity of *G. bracteata* extracts were mentioned in (Table 3) [3]. The ethanol extract of the *G. bracteata* with the maximum percentage of inhibition (55.46±0.01) at 950µg/ml was shown in the (Fig 2), followed by chloroform and ethyl acetate at the same concentration (Table 2). Similarly, Selvi and Yogananth [15] documented their findings the concentration of the plant extract increases, the potential inhibitory action percentage also increased. When compared with both enzyme activities, α-amylase activity on an ethanol extract of *G. bracteata* leaf extract has the highest inhibitory activity and this is the first study for the evaluation of the enzymatic activity of *G. bracteata*.

Table 2 α - glucosidase activity of *Grewia bracteata* in percentage

Concentration (ug/ml)	Ethanol	Ethyl acetate	Chloroform	Acarbose
59.37	44.59±0.01	48.54±0.01	46.56±0.02	45.78±0.02
118.75	47.96±0.01	48.85±0.02	47.62±0.02	46.97±0.02
237.5	49.27±0.01	49.00±0.01	48.65±0.00	49.54±0.02
475	50.99±0.01	50.17±0.00	50.61±0.01	54.61±0.01
950	55.46±0.01	51.84±0.08	52.61±0.02	61.28±0.02

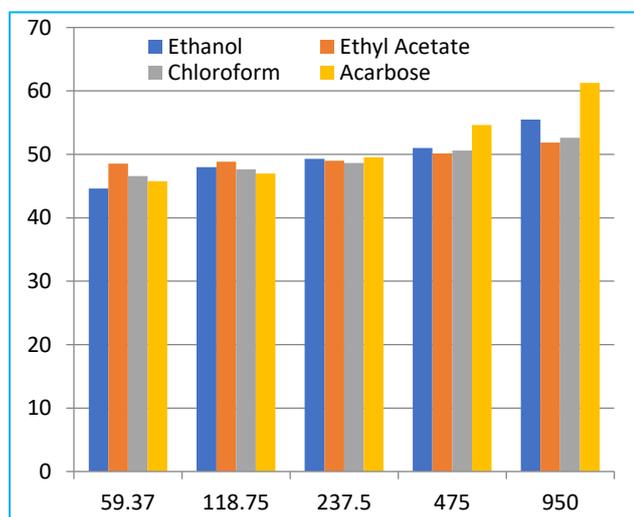


Fig 2 α - Glucosidase activity of *Grewia bracteata* in Percentage

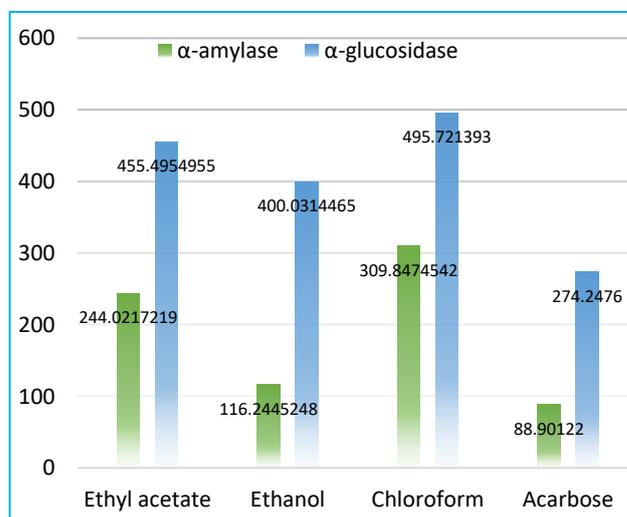


Fig 3 IC₅₀ value of anti-diabetic activity of *Grewia bracteata*

Table 3 IC₅₀ value of anti-diabetic activity of *Grewia bracteata*

Solvents used and control	IC ₅₀ Value (µg / ml)	
	α-amylase	α-glucosidase
Ethyl acetate	244.02±0.57	455.49±1.38
Ethanol	116.24±1.16	400.03±1.09
Chloroform	309.84±0.67	495.72±1.67
Acarbose	88.90±1.42	274.24±1.24

CONCLUSION

In conclusion with this study exhibited the highest polarity solvents like ethyl acetate, ethanol, and chloroform

extracts of *Grewia bracteata* leaves about the preliminary phytochemicals, in vitro anti-diabetic activities. The result of the determination of phytochemicals revealed that the secondary metabolites like proteins, alkaloids, flavonoids, tannins, carbohydrates, amino acids, phenolic compounds were present in both ethanol and ethyl acetate extracts. Saponins and glycosides were absent in chloroform extract. In enzymatic assays the best inhibition was noted on the α-amylase enzyme in all the extracts used when compare with α-glucosidase enzyme. So, the *Grewia bracteata* leaf extracts has the potential of high antidiabetic property due to the presence of various phytocompounds.

LITERATURE CITED

1. Aba PE, Asuzu IU. 2018. Mechanisms of actions of some bioactive anti-diabetic principles from phytochemicals of medicinal plants: A review. *Indian Journal of Natural Products and Resources* 9(2): 85-96.
2. Ablat A, Mohamad J, Awang K, Shilpi JA, Arya A. 2014. Evaluation of anti-diabetic and antioksidan properties of *Brucea javanica* seed. *The Scientific World Journal* 2014: 1-8.
3. Egharevba GO, Dosumu OO, Oguntoye SO, Njinga NS, Dahunsi SO, Hamid AA, Anand A, Amtul Z, Priyanka U. 2019. Antidiabetic, antioxidant and antimicrobial activities of extracts of *Tephrosia bracteolata* leaves. *Heliyon* 5(8): e02275.
4. Fahad FI, Barua N, Islam MS, Sayem SAJ, Barua K, Uddin MJ, Chy MNU, Adnan M, Islam MN, Sayeed MA, Emran TB, Simal-Gandara J, Pagano E, Capasso R. 2021. Investigation of the pharmacological properties of lepidagathis hyalina nees through experimental approaches. *Life (Basel)* 2021: 11(3): 180.
5. Rabizadeh F, Mirian MS, Doosti R, Kiani-Anbouhi R, Eftekhari E. 2022. Phytochemical classification of medicinal plants used in the treatment of kidney disease based on traditional Persian medicine. *Evidence-Based Complementary and Alternative Medicine* 2022: 1-13.
6. Fernández F, Guerrero RJ, Sánchez-Restrepo AF. 2021. Systematics and diversity of Neotropical ants. *Revista Colombiana de Entomología* 47(1): 1-20.
7. Gracz-Bernaciak J, Mazur O, Nawrot R. 2021. Functional studies of plant latex as a rich source of bioactive compounds: Focus on proteins and alkaloids. *International Journal of Molecular Sciences* 22(22): 124-127.
8. Hasan MR, Uddin N, Hossain MM, Hasan M, Yousuf ME, Lopa SS, Rahman T and Choudhuri MSK. 2014. Invitro α -amylase inhibitory activity and invivo hypoglycemic effect of ethyl acetate of *Mollatus repandus* (Willd) Muell. in stem in rat model. *Journal of Coastal Life Medicine* 2: 721-726.
9. Jayashree BS, Yusuf S, Kumar DV. 2009. Synthesis of some coumarinyl chalcones of pharmacological interest. *Asian Journal of Chemistry* 21(8): 5918.
10. Sinha J, Purwar S, Chuhan SK, Rai G. 2015. Nutritional and medicinal potential of *Grewia subinaequalis* DC. (syn. *G. asiatica*.) (Phalsa). *Journal of Medicinal Plants Research* 9(19): 594-612.
11. Kubmarawa D, Khan ME, Punah AM, Hassan M. 2008. Phytochemical screening and antimicrobial efficacy of extracts from *Khaya senegalensis* against human pathogenic bacteria. *African Journal of Biotechnology* 7(24).
12. Kumar RS, Raja NK, Vijay M, Raja CSG. 2016. Anti-oxidant, anti-diabetic, antimicrobial and hemolytic activity of *Solanum torvum* and *Solanum trilobatum*. *Jr. Pharm. Sci. Research* 8(8): 725-728.
13. Oyedemi SO, Adewusi EA, Aiyegoro OA, Akinpelu DA. 2011. Antidiabetic and haematological effect of aqueous extract of stem bark of *Azelia Africana* (Smith) on streptozotocin-induced diabetic Wistar rats. *Asian Pacific Journal of Tropical Biomedicine* 1: 353-358.
14. Patel DK, Kumar R, Laloo D, Hemalatha S. 2012. Natural medicines from plant source used for therapy of diabetes mellitus: an overview of its pharmacological aspects. *Asian Pacific Journal of Tropical Disease* 2: 239-250.
15. Selvi R, Yogananth N. 2016. In vitro evaluation of antidiabetic potential of leaf and stem extracts of *Solanum xanthocarpum* and *Solanum nigrum*. *Int. Jr. Adv. Res. Biol. Science* 3(12): 191-195.
16. Sindhu RK, Arora S. 2012. Evaluation of phenolic contents and antioxidant potential of *Murraya koenigii* (L) spreng roots. *Journal of Applied Pharmaceutical Science* 2(11): 120-122.
17. Suhashini R, Sindhu S, Sagadevan E. 2014. In vitro evaluation of anti-diabetic potential and phytochemical profile of *Psoralea corylifolia* seeds. *Int. Jr. Pharmacogn. Phytochem. Research* 6: 414-419.
18. Xavier TF, Sabitha R, Balavivekananthan S. 2022. In vitro pharmacological investigations of *Oxystelma esculentum* R.Br. and in silico molecular docking analysis of its leaf constituents on diabetic related target. *South African Journal of Botany* 149: 320-338.