

Impact of Mulberry Leaf Enriched with Leucine on Dehydrogenase Activities and Economic Characters of *Bombyx mori* L.

Abhishek B. S.¹, Anil Kumar M. N.*², Sandya B. Raju³ and Kiran Kumara D.⁴

¹⁻⁴ Laboratory of Silkworm Physiology and Biochemistry, Department of Studies in Sericulture Science, University of Mysore, Manasagangotri, Mysore - 570 006, Karnataka, India

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Abstract

The growth and development of silkworm significantly regulated by nutrition. Like any other heterotrophic insects, the silkworm, *Bombyx mori* L. obtain water and other chemical compounds from the diet which determine growth and silk production. An attempt has been made in the present investigation to record the effect of mulberry leaf fortified with leucine at varied concentrations viz., 0.5, 1.0 and 1.5% on succinate dehydrogenase (SDH) and lactate dehydrogenase (LDH) activity levels and economic parameters in bivoltine double hybrids namely FC₁ × FC₂ and FC₂ × FC₁. The results of the study inferred that, both the hybrids expressed maximum activities of succinate and lactate dehydrogenase at 1.5% supplementation over remaining concentrations as well as control batch. The highest enzyme activities were recorded in the fat body tissue during 5th instar 6th day followed by 3rd and 1st day larvae. Further, higher larval weight, cocoon weight, shell weight, shell percentage, filament weight, filament length and renditta were recorded at 1.5% supplementation as compared to control batch.

Key words: Silkworm hybrids, Leucine, Supplementation, Enzymes, Commercial parameters

The domesticated silkworm, *Bombyx mori* L derive nutrients from mulberry leaf for the consolidation of biomass in the larval stage and utilize energy reserve during non-feeding stage of metamorphosis [1]. By and large, the quality and quantity of silk produced by this sericigenous insect mainly decided by mulberry leaf nutrients, environmental and edaphic factors, agronomical practices besides its genetic endowment [2-3]. The nourishment of silkworm with quality dietary nutrients is most important component of nutrition which promotes metabolic process inturn enhance larval growth and silk production [4]. Perhaps, utilization of dietary nutrients is proportional to synthesis of silk proteins in the silk gland [5]. Hence it is obvious that, exogenous supplementation of nutrients through the diet often recommended to enhance economic parameters of the silkworm by using various fortifying agents with mulberry leaf viz., honey and lemon, folic acid, ferrous magnesium sulphate, ecdysteroids, alfalfa [6-10]. The *B.mori* derived 72-86% of amino acid from the mulberry and nearly about 60% of the absorbed amino acids are utilized for synthesis of silk protein [11]. Moreover, the dietary proteins and amino acids not only regulate growth and silk production, but also digestibility and protein concentration in the haemolymph [12]. It is well document that *B. mori* require ten essential amino acids and are mostly obtained by diet [13]. A quite a good number of reports are available on exogenous supplementation of amino acids such as phenyl alanine, valine, methionine, glycine, alanine, glutamine and threonine by the diet at varied concentrations which enhance larval growth,

economic parameters, enzyme activities and synthesis of biomolecules in the silkworm [14-18].

The Succinate dehydrogenase (SDH) found in mitochondria of different tissue of insects including *B. mori*. It establishes connecting link between citric acid cycle and electron transport chain oxidation and play a pivotal role in carbohydrate metabolism. On the other hand, lactate dehydrogenase (LDH) is a glycolytic enzyme responsible for reversible conversion of lactate to pyruvate with the aid of NAD found abundantly in the fat body tissue of insects. In *Bombyx mori* L, elevated levels of Succinate dehydrogenase (SDH) and LDH activities were noticed when larvae supplemented with tryptophan and riboflavin as compared to controls and their activities differ sharply among different silkworm breeds [19], [29]. Furthermore, Succinate dehydrogenase (SDH) expressed positive correlation with respect to commercial parameters of silkworm [20]. However, information on mulberry leaf fortified with leucine on dehydrogenase enzymes and economic parameters is meager. Hence, current investigation was undertaken to record changes in dehydrogenase activities as well as economic parameters during the supplementation of leucine with mulberry leaf.

MATERIALS AND METHODS

The bivoltine double hybrids namely FC₁ × FC₂ and FC₂ × FC₁ were selected for the present investigation and larvae were reared as per the method [21].

*Correspondence to: Anil Kumar M. N., E-mail: anilkumar2068@gmail.com; Tel: +91 9880917421

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Supplementation of leucine

The leucine at varied concentrations viz., 0.5, 1.0 and 1.5% were prepared with distilled water and sprayed on ventral surface of mulberry leaves and surface dried under shade before feeding to the silkworms. For the experimentation purpose, silkworm larvae divided into four batches viz., batch I (T₁), batch II (T₂) and batch III (T₃) were reared with mulberry leaf supplemented with leucine at 0.5, 1.0 and 1.5%, respectively. The control batch (T₄) was maintained parallelly and larvae were reared on mulberry leaves sprayed with distilled water. The treated leaves were fed to silkworms once in a day during IV and V instars. In each treatment, three replications were maintained each with 100 larvae along with control. The fat body tissue was collected by dissecting V instar 1st, 3rd and 5th day old larvae of respective treatments and control batch. The tissue homogenate of 1% was prepared with distilled water. After centrifugation of the supernatant was collected and used for estimation of dehydrogenase enzymes. In addition, larval weight, cocoon weight, shell weight, shell percentage, filament length, filament weight, denier and renditta were also recorded. A minimum of twenty larvae were used from each treatment to measure succinate and lactate dehydrogenase activities.

Estimation of succinate and lactate dehydrogenase enzymes

The SDH and LDH activities were estimated as suggested by [22]. One ml of tissue extract was incubated with 1ml of sodium succinate, 1ml of phosphate buffer and 1ml of INT at 37 °C for 1 h. For LDH, lithium lactate was used as a substrate. The enzyme activities were stopped by adding 6ml of glacial acetic acid and 6ml of toluene. Finally, the test tubes were kept in refrigerator for overnight and OD was measured at 495nm using spectrophotometer against prepared blank. The standard graph was used for calculation and enzyme activities were expressed in terms of μ moles of formazan /g protein/h.

The commercial parameters such as shell ratio, filament length, denier and renditta were calculated by using following formulae:

$$\text{Shell ratio (\%)} = \frac{\text{Shell weight (g)}}{\text{Cocoon weight (g)}} \times 100$$

$$\text{Filament Length (L)} = R \times 1.125$$

R = Number of revolutions recorded by an eprouvette

1.125 = Circumference of eprouvette in meter.

$$\text{Denier} = \frac{\text{Total weight of reeled silk (g)}}{\text{Total length of reeled silk (m)}} \times 9000$$

It denotes thickness of silk filament

$$\text{Renditta} = \frac{\text{Weight of cocoon reeled (g)}}{\text{Weight of raw silk (g)}}$$

Unit quantity of cocoons required to produced one unit of raw silk.

The obtained data were compiled and statistically analyze by standard methods.

RESULTS AND DISCUSSION

Influence of leucine supplementation on dehydrogenase enzymes

In the living organisms, various metabolic functions are carried out at cellular level by amino acids. The amino acids and proteins are the main sources of nitrogen which not only regulate growth but also silk production. The silkworm larvae

utilize about 65 per cent of nitrogen during fifth instar for the production of silk [23-24]. It is the diet determine the quantity of free amino acids in the haemolymph. Moreover, increased free amino acid contents in larval haemolymph during supplementation of amino acids by the diet (mulberry leaf) because of synthesis of amino acids by non-protein sources such as glucose and fatty acids [25]. In *B. mori*, the function of fat body is analogue to vertebrate liver in which various metabolic process take place as haemolymph is close contact with fat body, silk gland and other tissues which helps to mobilize amino acids and other metabolites [26]. The overall growth of silkworm and synthesis of silk proteins are in fine tune between anabolic and catabolic processes. The leucine (C₆H₁₃No₂) is most potent ketogenic essential amino acid is cleaved catabolically by transamination, decarboxylation and dehydrogenation to form acetoacetate and acetyl-CoA [27]. Perhaps, synthesis of acetylc-CoA not only by metabolism of leucine but also by oxidation of fatty acids, amino acids and pyruvate from carbohydrates. The acetylc-CoA often regarded as a focal point in metabolism and is formed from coenzyme A (CoA). It is widely utilized at cellular level such as oxidation of glucose in the citric acid cycle, synthesis of cholesterol, fatty acids and steroid hormones [28]. The data pertaining to enzymes clearly depicts that the silkworm larvae FC₂ × FC₁ reared on mulberry leaf fortified with leucine registered maximum activity levels of both SDH and LDH in fat body tissue at 1.5% concentration (5.45 μ m of formazan/ g protein /h) and (5.31 μ m of formazan / g protein /h) over respective controls (4.19 μ m of formazan) and (4.17 μ m of formazan) during 5th instar 5th day followed by 3rd and 1st day. Similar results were also observed in FC₁ × FC₂ at 1.5% concentration (Fig 1-2).

The SDH belongs to class oxidoreductases which metabolize succinate into fumarate and during this process one hydrogen ion and two electrons are transfer to FAD. In TCA cycle, SDH transfer electron to cytochrome-b of respiratory chain and such association facilities quick electron transfer for oxidation of glucose or glycogen. Similarly, LDH is a glycolytic enzyme which converts lactic acid into pyruvate in which one hydrogen ion and 2 electrons are transfer to NAD. The significant variations were noticed with respect to SDH and LDH activities in insects including silkworm breeds and even at different stages of its life cycle during supplementation of nutrients [19]. It is clear from the current investigation is that both the silkworm hybrids expressed higher levels of SDH and LDH activities with higher concentration (1.5%) of leucine supplementation than lower concentrations (0.5 and 1.0%) as compared to control. The increase in SDH activity in the leucine supplemented batches might be due enhanced synthesis of acetyl-CoA which diffused into inner membrane of mitochondria to the sight of TCA cycle which in turn accelerate biological oxidation. Similarly, it is presumed that elevation in LDH activity in the larvae supplemented with leucine has resulted in increased synthesis of acetyl-CoA from the pyruvate. These results are in conformity with the observations of previous investigations [29] where in silkworm hybrids reared on mulberry leaves fortified with tryptophan at varied concentrations expressed increase in levels of SDH and LDH activities over control. Likewise, FC₁ and FC₂ silkworm larvae administrated with pyridoxine at different concentrations viz., 100, 500 and 1000ppm has resulted enhanced SDH activity when compared to control. However, the hybrid FC₂ × FC₁ expressed maximum levels of SDH and LDH activities compared to FC₁ × FC₂ as it has a greater potential to utilize the additional supplementation of leucine. These results are in agreement with earlier studies [29] who has reported the

utilization of tryptophan varies among silkworm hybrids. Further, SDH and LDH activities gradually increased from 5th instar 1st day followed by 3rd and 5th day. It clearly depicts that mature larva require more amount leucine and hence

progressive increase in enzyme activities were noticed. The elevated levels of SDH and LDH were observed as the larval age proceeds during 5th instar 1st day to 5th day during supplementation of tryptophan [38].

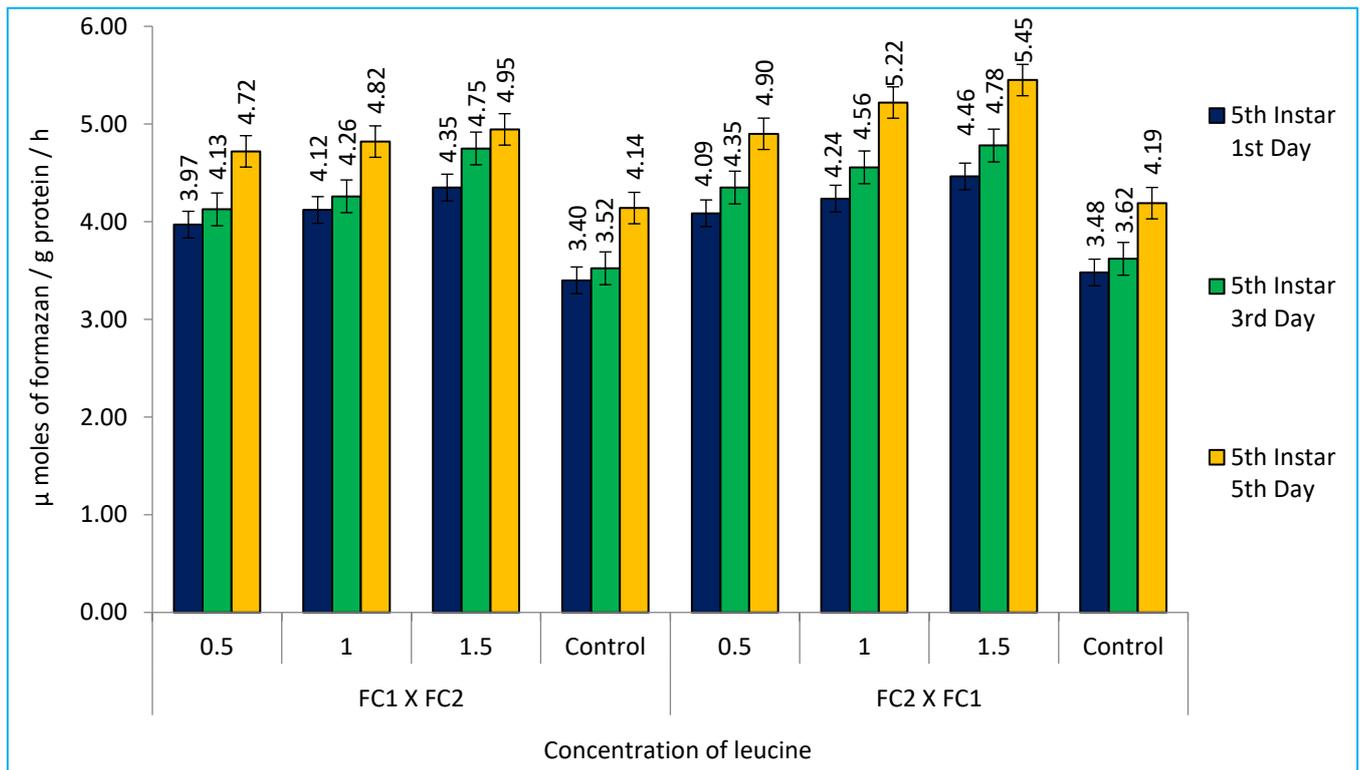


Fig 1 Effect of mulberry leaves supplemented with leucine at varied concentration on SDH activity in fat body of bivoltine double hybrids

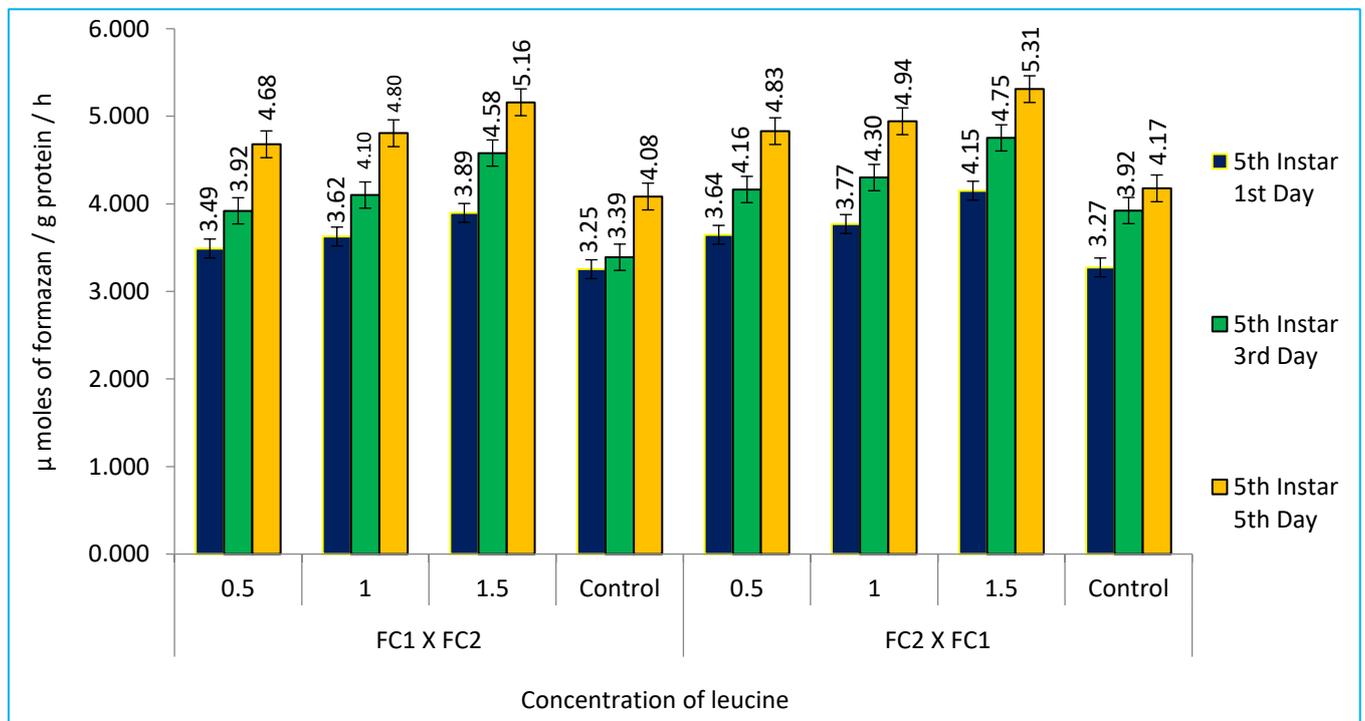


Fig 2 Effect of mulberry leaves supplemented with leucine at varied concentration on LDH activity in fat body of bivoltine double hybrids

Matured larval weight

Generally, proteins and amino acids supplements promotes growth and development which leads increased larval weight. Hence, gain in larval weight might be due to additional supplementation of leucine by the diet. Like other lepidopteran larvae, silkworm also exhibit intense feeding activity

particularly during fifth instar and obviously it is one of the reasons in gaining larval weight [30]. Silkworm hybrids reared on mulberry leaves fortified with leucine at different concentrations exhibited notable influence on larval weight. The FC₂ × FC₁ and FC₁ × FC₂ expressed more larval weight of 4.85 and 4.72g (Table 1) when worms supplemented with

leucine at 1.5% concentration against controls. The present findings are in harmony with the previous observations [31] who has reported that the silkworm hybrids supplemented with phenylalanine at 1.5% concentration exerted higher larval weight. Moreover, fat body and integument synthesis large

amount of proteins from amino acids and their accumulation leads to gain in larval weight [32]. Similar results were observed which improves larval weight by the supplementation of amino acids such as methionine, alanine, arginine, tryptophan and glycine [33-37].

Table 1 Impact of mulberry leaves supplemented with leucine at varied concentrations of larval and cocoon characters

Hybrid	Concentration (%)	Larval weight (g)	Cocoon weight (g)	Shell weight (g)	Silk percentage (%)	Filament length (m)	Filament weight (g)	Denier	Renditta
FC ₁ × FC ₂	0.5	4.24±0.02	2.05±0.01	0.33±0.03	16.09±0.05	985±2.08	0.32±0.01	2.92±0.09	6.41±0.09
	1	4.51±0.02	2.12±0.03	0.36±0.02	16.98±0.02	1050±2.00	0.34±0.01	2.91±0.08	6.23±0.08
	1.5	4.72±0.01	2.20±0.01	0.38±0.02	17.27±0.02	1150±5.00	0.36±0.01	2.81±0.03	6.15±0.04
	Control	3.95±0.05	1.85±0.03	0.29±0.01	15.67±0.04	880±5.00	0.27±0.01	3.10±0.01	6.82±0.08
FC ₂ × FC ₁	0.5	4.55±0.05	2.14±0.02	0.36±0.01	16.82±0.05	1002±4.00	0.33±0.01	2.90±0.03	6.33±0.08
	1	4.62±0.02	2.26±0.02	0.40±0.01	17.69±0.02	1125±3.00	0.37±0.01	2.80±0.01	6.10±0.03
	1.5	4.85±0.05	2.35±0.03	0.42±0.01	17.87±0.05	1225±1.00	0.39±0.01	2.79±0.03	6.02±0.05
	Control	4.10±0.01	1.95±0.01	0.31±0.01	15.89±0.05	895±3.00	0.29±0.02	2.91±0.01	6.72±0.07

Cocoon weight

The cocoon weight is an important economic parameter differ among voltine groups. It depends upon ability of silkworm breeds which converts mulberry nutrients into cocoon formation. Supplementation of leucine at varied concentrations in silkworms registered encouraging results on cocoon weight. The larvae reared on leucine at 1.5% expressed higher cocoon weight of 2.35 and 2.20g in FC₂ × FC₁ and FC₁ × FC₂, respectively (Table 1). The increase in cocoon weight in both the breeds might be due to absorption of leucine by the body cells and get assimilated into cellular structure. These results are on par with the earlier findings [33] who have noticed that increased cocoon weight by the silkworm BSRI 83/3 reared on mulberry leaves extra foliated with methionine and tryptophan at 500ppm over remaining concentrations viz., 250, 750 and 1000ppm. This type of trend was also noticed with lycine, alanine, cysteine and glycine at 2% registered highest cocoon weight of 2.835, 2.695, 2.636 and 2.548g, respectively against control (2.254g) [37]. Likewise, worm nourished with alanine and asparagine at 0.2% exerted higher cocoon weight.

Shell weight

Shell weight is an important commercial character which denotes actual silk content of cocoon. It is evident from the data is that silkworms treated with leucine at 1.5% recorded higher shell weight (0.42g) in FC₂ × FC₁ and (0.38g) FC₁ × FC₂ over respective controls (0.31 and 0.29g) (Table 1). It is presumed that increase in shell weight is due to utilization of leucine which might have enhance synthesis of silk proteins in turn reflects on shell weight. These results corroborate the earlier studies [38] who have opined that silkworm larva MU11 and MU303 reared on mulberry leaves enriched with methionine at 0.5% concentration recorded higher shell weight of 0.217 and 0.215mg over controls (0.160 and 0.162 mg), respectively. Similarly higher shell weight was obtained with phenylalanine supplementation [39-40]. Furthermore, worms treated with threonine at the rate of 2% exerted higher shell weight and with proline and amino acids mixture [41-42].

Shell ratio

It indicates silk productivity of the silkworm races/breeds and helpful in fixing cocoon price. A concentration of leucine at 1.5% has resulted in higher shell ratio of 17.87 and 17.27% in FC₂ × FC₁ and FC₁ × FC₂, respectively (Table 1) when compared to control batches. The current observations are

in conformity with the findings of previous investigations [14] where in silkworm hybrids FC₁ × FC₂, FC₂ and FC₁ were nourished with mulberry leaf fortified with phenylalanine at 1.5% registered higher shell percentage over respective controls. Similarly, larva fed on mulberry leaves enriched with lycine, alanine, cysteine and glycine recorded higher shell weight at 2.5% and with tryptophan at 550ppm concentration [33], [37].

Filament length

It is one of the quantitative parameters varies among silkworm breeds [43]. The silkworms reared on mulberry leaf extra foliated with leucine at varied concentrations registered positive impact on filament length. Both hybrids FC₂ X FC₁ and FC₁ X FC₂ treated with leucine at 1.5% registered longer filament length of 1225 and 1150m over control batches (895 and 880m) (Table 1). These results are on par with earlier studies [31] who has concluded that bivoltine hybrids FC₂ and FC₁ treated with phenylalanine at 1.5% expressed longest filament length as against respective controls. Similar result was observed with lycine, alanine, cysteine and glycine [37] and with amino acid mixture (proline) at 0.01% [44].

Filament weight

This character is mainly decided by quantum of mulberry leaf fed by silkworm. The worm reared on mulberry leaves fortified with leucine recorded marked improvement with respect to filament weight as compared control. The maximum filament weight of 0.39 g (FC₂ × FC₁) and 0.36 g (FC₁ × FC₂) were obtained with leucine at 1.5% as against respective controls (0.29 and 0.27g) (Table 1). The present findings are in agreement with the previous observations [33] who have opined that larva treated with methionine and tryptophan scored higher filament weight. Similar trend was also noticed when worms are nourished with methionine at 0.5% and with phenylalanine, valine and methionine at 1.5% [14], [38].

Denier

It is one of the fascinating quality parameters which denotes thickness of the cocoon silk filament and varies along its length [45]. Generally, finer or lower denier silk is preferred in weaving sector. It is evident from current investigation is that silkworms treated with leucine at 1.5% expressed lower denier of 2.79 and 2.81 in FC₂ × FC₁ and FC₁ × FC₂ hybrids, respectively (Table 1). The present findings are comparable

with those of earlier researchers [29], [33] who have observed that the larvae reared on mulberry leaves fortified with tryptophan at 0.5% expressed finer denier against control. This type of trend was also observed with amino acid mixture at 0.01% (2.42) over control (2.50) [44].

Renditta

This parameter denotes total available silk obtained from cocoons and lower value is preferred by the silk industry. The data pertaining to renditta registered encouraging results when silkworm larvae nourished on mulberry leaves enriched with leucine at different concentrations. The lowest renditta of 6.02 ($FC_2 \times FC_1$) and 6.15 ($FC_1 \times FC_2$) were obtained with leucine supplementation ment at 1.5% concentration (Table 1). The improvement for this parameter was noticed in the larvae treated with leucine. The current observations are in concurrence with the findings of previous investigations [29] who has obtained low renditta when the larvae treated with

tryptophan at 1.5% concentration. Similarly, FC_1 and FC_2 larvae reared on mulberry leaf fortified with phenylalanine at 1.5% exerted lowest renditta over respective controls [31]. Likewise, administration of silkworm larvae with phenylalanine, methionine and valine at 1.5% registered lowest renditta [14]. Overall, these observations suggest that amino acid supplementation, particularly at a concentration of 1.5%, can be a viable strategy to enhance the efficiency of silk production by reducing renditta.

CONCLUSION

The results of current investigation inferred that; the silkworm larvae nourished with mulberry leaves supplemented with leucine at 1.5% enhance dehydrogenase enzyme activities as well as commercial parameters. Our findings could be utilized for the formulation of foliar spray and even for preparation of synthetic diets.

LITERATURE CITED

- Harrison JF, Hadded GG. 2011. Effects of oxygen on growth and size; synthesis of molecular organism and evolutionary studies with *Drosophila melanogaster*. *Annu. Rev. Physiology* 73: 95-113.
- Anantha Raman KV, Magadam SB, Shivakumar GR, Giridhar K, Datta RK. 1995. Correlation studies on different economic and nutritional parameters in *Bombyx mori* L. hybrids. *Indian Jr. Sericulture* 34(2): 118-121.
- Bongale UD, Chaluvachari. 1993. Evaluation of four mulberry varieties by leaf biological analysis and bioassay with *Bombyx mori*. *Jr. Indian Botanical Society* 72: 59-62.
- Vijila NMK. 2018. Beneficial effects of *Bacillus licheniformis* and *Basillus niabensis* on growth and economic characteristics of silkworm, *Bombyx mori* L. *Int. Jr. Chem. Studies* 6: 750-1754.
- Fakuda T, Kamegama T, Matsuda MA. 1963. Correlation between the mulberry leaves consumed by the silkworm larvae in different ages of the larval growth and production of the cocoon fiber spun by the silkworm larvae and the eggs laid by the silkworm. *Bull. Seric. Exp. Stn.* 18: 165-171.
- Saritha D, Siva Prasad S. 2022. Synergistic effect of honey and lemon juice-enriched mulberry diets on the growth of silkworm, *Bombyx mori* L. during metamorphosis. *Global Jr. Res. Analysis* 11(10): 1-16.
- Banerjee SK, Khan AR. 1992. Effect of vitamin B6 on the fecundity of *Bacillus thuringensis*. Var. Kurstaki treated *Bombyx mori* L. *Bangladesh Jr. Zoology* 20: 361-362.
- Nirwani RB, Kaliwal BB. 1995. Effect of ferrous magnesium sulphate supplementation on some commercial characters of *B. mori*. *Bull. Ser. Research* 6: 21-27.
- Mondal M, Narayanswamy K, Tandon B. 2018. Seri more- An effective plant-based growth promoter for increasing an economic characters of silkworm *Bombyx mori* L. in large scale in different seasons. *Int. Jr. of Adv. Research* 4(1): 804-808.
- Thulasi N, Siva Prasad S. 2015. Determination of minimum effective concentration of alfalfa for optimal growth, metabolism and silkproduction in the silkworm, *B. mori*. *Jr. Bio. Innovation* 4(1): 18-27.
- Lu SL, Jiang ZD. 1988. Absorption and utilization of amino acids in mulberry leaves by *Bombyx mori* L. *Acta Sericologica Sin.* 14: 198-204.
- Seol GY. 1982. Studies on the effects of various levels of protein in the artificial diet on nutritional physiology of the silkworm, *Bombyx mori* L. *Seric. Jr. Korea* 23(2): 37-49.
- Devaiah MC, Narayanaswamy KC, Maribashetty VG. 1999. Advances in mulberry sericulture. 1st Ed. CVG Publications Bangalore, India. pp 320-344.
- Jayaraju P. 2022. Influence of mulberry leaf fortified with three essential amino acids on biochemical changes and economic characteristics of the silkworm, *Bombyx mori* L. *Ph. D. Thesis*, University of Mysore, Mysore.
- Jayaraju P, Anil Kumar MN. 2021. Variation in the aminotransferase enzymes in response to the dietary supplementation of amino acids in the silkworm hybrids. *Res. Jr. of Agril. Sciences* 12(6): 2175-2179.
- Sridhar P, Radha NV. 1987. Effect of supplementing glycine to the feed of silkworm, *Bombyx mori* L. *Proc. Seric. Symp. Semi.*, Coimbatore. pp 88-98.
- Rahman Khan A, Saha BN. 1995. Growth and development of the mulberry silkworm, *Bombyx mori* L. on feed supplement with alanine and glutamine. *Sericologia* 35(4): 657-663.
- Nicodemo D, Juliano E. 2014. Oliveira Impact of different silkworm dietary supplement on its silk performance. *Journal of Master Science* 49: 6302-6310.
- Anil Kumar MN, Jagadisha MC, Mahadevaswamy M. 2017. Impact of aqueous plant extract on the economic traits of the silkworm breeds CSR2. *International Journal of Academic Research* 2(7): 94-100.
- Mahesha HB, Farshid GK, Thejaswini PH. 2015. Studies on the correlation between protein, amylase, succinate dehydrogenase, esterase and alkaline phosphatase of silkworm, *Bombyx mori* L. *Int. Jr. Pure App. Bioscience* 3(2): 173-180.
- Dandin SB, Giridhar K. 2010. *Handbook of Sericulture Technologies*. Central Silk Board, Bangalore. pp 427.
- Nachals MM, Marguleis SL, Seligman AM. 1960. Sites of electron transfer to tetrazolium salts in the succinoxidase system. *Jr. Biol. Chemistry* 239(9): 2737-2743.
- Horie Y, Watanabe K. 1983. Effect of dietary pyridoxine on larval growth, free amino acid pattern in haemolymph and uric acid excretion in the silkworm, *Bombyx mori*. *Insect Biochemistry* 13: 205-212.

24. Unni BG, Das P, Gush AC. 2000. Silk gland and the biosynthesis of silk by silkworms; sericulture in India. (Eds). Agrawal A.O and Seth M.K. Bishen Singh Mahendra Pal Singh, Dehradun, India. pp 251-269.
25. Bose PC, Majumder SK, Sengupta K. 1989. Role of amino acids in silkworm, *Bombyx mori* L. nutrition and their occurrence in haemolymph, silk gland and silk cocoon. *Indian Jr. Sericulture* 28: 17-31.
26. Noguchi A, Takeshita H, Shigemastu H. 1974. Inter relationship between the silk gland and other tissue in protein metabolism in the latest larval stage of the silkworm, *Bombyx mori*. *Jr. Insect. Physiology* 20: 783-794.
27. Rama Rao AVSS. 2004. *A Textbook of Biochemistry*. UBS Publishers. Bangalore. India. pp 412-416.
28. Deb AC. 2011. *Fundamental of Biochemistry*. 10th Edition. New central book agency, Kolkata, India. pp 393-394.
29. Anil Kumar MN. 2018. Application of tryptophan as supplementary nutrient on dehydrogenase activities and economic parameters in *Bombyx mori* L. *Journal of Pharmaceutical, Chemical and Biological Science* 6(3): 209-217.
30. Dow JAT. 1986. Insect midgut function. *Adv. Insect Physiology* 19: 187-328.
31. Anil Kumar MN. 2018b. Effect of dietary supplementation of phenylalanine on aminotransferase enzymes and economic characters of *Bombyx mori* L. *Indian Jr. Sericulture* 57(1/2): 22-29.
32. Jadhav V, Kallapur VL. 1989. Contribution of proteins to the cocoon shell in the fifth instar silkworm, *Bombyx mori* L. *Entamon* 14: 21-24.
33. Laz R, Gani A, Saleh Reza AM. 2006. Effect of methionine and tryptophan on some economic characters in the mulberry silkworm, *Bombyx mori* L. *Univ. Jr. Zoology, Rajshahi University* 25: 57-62.
34. Radjaabi R. 2010. Effect of Mulberry leaves enrichment with amino acid supplementary nutrients on silkworm, *Bombyx mori* L. at North of Iran. *Academic Journal of Entomology* 3(1): 54-51.
35. Davis GRF. 1978. Growth response of larvae of *Tenebrio molitor* to concentration of dietary amino acids. *Jr. Stored Products Research* 14: 69-71
36. Krishnappa JB. 1987. Influence of amino acids supplementation on growth and development of mulberry silkworm, *Bombyx mori* L. *M. Sc., Thesis*, University of Agricultural Science, Bangalore, India.
37. Meeramaideen M, Rajasekar P, Sumathi K, Prabu G. 2017. Studies on the morphometric and economic parameters analysis of silkworm, *Bombyx mori* L. (Lepidoptera: *Bombycidae*) fed with amino acid (lysine) treated MR2 mulberry leaves. *Int. Jr. Modn. Res. Revs.* 5(1): 1468-1473.
38. Anil Kumar MN, Sunil Kumar B. 2018. Influence of fortified mulberry leaf with methionine on the economic traits and aminotransferase activity in *Bombyx mori* (Lepidoptera: *Bombycidae*). *Int. Jr. Curr. Adv. Research* 7(1): 9258-9262.
39. Shyamala MB, Visweswara Gowda BL. 1980. Effects of supplementing phenylalanine to Mysore-5 and local varieties of mulberry on silk production in *Bombyx mori* L. *Proc. Seri. Symp.*, Coimbatore. pp 88-92.
40. Radha NV, Sridhar P. 1983. Effect of supplementing amino acid to mulberry leaves on silk production. *Nat. Semi. Silk. Res. Dev.* 1983, Central Silk Board, Bangalore, Abst.125.
41. Nicodemo D, Oliveira JE, Sedano AA, Marconcini JM, Tonoli GHD. 2014. Impact of different silkworm dietary supplements on its silk performance. *Jr. Master Science* 49: 6302-6310.
42. Bhojne I, Naik RL, Kharbade SB. 2014. Effect of leaf supplementation with secondary metabolites on economic traits of mulberry silkworm. *Int. Jr. Entomol. Research* 2(1): 29-32.
43. Yokoyama T. 1979. Silkworm selection and hybridization. *In: Genetics in Relation to Insect Management*. Workshop Papers, The Rockefeller Foundation. pp 71-83.
44. Bhojne I, Naik RL. 2013. Effect of leaf supplementation with secondary metabolites on rearing parameters of mulberry silkworm. *Inr. Jr. Res. Scie. and Tech.* 2(6): 60-65
45. Premalatha VR, Raghevendra Rao D, Rama Mohan Rao P, Ahsan MM, Datta RK. 1997. Studies on the filament size variation in the single cocoon filament in the multi X Bi hybrids of the silkworm, *Bombyx mori* L. Current Technology Seminar, 18-19th September, Central Sericulture Research and Training institute, Mysore. Abstract. pp 23.