

# Effect of Sodium Chloride and Boron Stress on Germination Percentage and Amylase Activity in Wheat (*Triticum aestivum* L.)

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## Abstract

Soil salinity and boron toxicity are significant abiotic stressors affecting agricultural performance. This study investigates the impact of these stresses on the germination and early seedling growth of the wheat variety HUW 234. Experiments were conducted during the rabi seasons of 2014-15 and 2015-16 using varying concentrations of sodium chloride (NaCl) and boron (B). Results indicated that increasing NaCl concentrations significantly decreased germination percentages and amylase activity, demonstrating the detrimental effects of soil salinity on wheat. Conversely, boron exhibited a more complex relationship: while high concentrations (4.5 and 5.0 mg B/kg soil) negatively affected germination and enzyme activity, an optimal concentration of 2.0 mg B/kg soil enhanced these parameters. These findings highlight the limited tolerance of HUW 234 to high soil salinity and its narrow optimal range for boron. Developing strategies to enhance salt and boron tolerance in wheat, such as genetic improvements and stress management approaches, is crucial for improving productivity in saline and boron-rich soils. Further research into the physiological and molecular mechanisms underlying these responses is essential for designing effective mitigation strategies.

**Key words:** *Triticum aestivum* L., Growth, High boron, Morphology, Saline conditions, Yield

Soil salinity is one of the most significant biotic stressors that negatively affects agricultural performance [1]. Salinity has a dual effect on crop growth. In the first phase, the decreased electrolyte content of the soil diminishes the water supply to plants. In saline soils, a variety of dissolved salts are present including NaCl, CaSO<sub>4</sub>, Na<sub>2</sub>SO<sub>4</sub>, MgCl<sub>2</sub>, Na<sub>2</sub>SO<sub>4</sub>, KCl and Na<sub>2</sub>CO<sub>3</sub> but NaCl is the most soluble and widely found [2-3]. Salinity has an impact on crop growth in two different stages. The first stage involves the reduction of water availability to plants due to the low osmotic potential of soil solution, further resulting in toxicity from ions like Na<sup>+</sup>, Cl<sup>-</sup> or B at excessive concentrations [4]. Additionally, affecting physiological processes in plants such as photosynthesis by reducing SC and transpiration while disrupting biosynthesis of photosynthetic pigments is salinity [5]. In the second phase, the accumulation of ions like Na<sup>+</sup>, Cl<sup>-</sup> or B that are over-concentrated causes ion toxicity [6]. In contrast to Na<sup>+</sup>, information about the transport of Cl ions is limited, even though it is one of the most common anions in many salinized soils [7]. Additionally, Cl<sup>-</sup> is a crucial micronutrient that regulates the activity of enzymes in the cytoplasm, it is a partner in photosynthesis, and it functions as an anti-ion to maintain the potential of the membrane and have a role in turgor and pH regulation [8-9].

Boron (B) stands as a micronutrient essential in its own right. It wields influence over major metabolic happenings and plant cellular function [10]. Actively growing plant regions that are teeming with cell division and rapid differentiation depend on this element highlighting its indispensable part in growth

dynamics. Yet the exact role of B within physiological processes like photosynthesis remains shrouded in mystery, even bordering on contradiction at times without any direct reports showcasing impact of boron on plant photosynthesis [11]; some researchers point towards an indirect link between B and photosynthesis based on findings in soybean crop plants [12]. Both excess [13] and deficiency [14] conditions of Boron paint pictures of reduced photosynthetic rates: the former hints at lowered CO<sub>2</sub> assimilation due to possible oxidative burdens coupled with diminished enzyme activities at photosystems along hampered electron transport [15].

The co-occurrence of salinity and boron toxicity is common, especially in arid or semi-arid regions [16]. However, the literature lacks a clear consensus on how these two stresses interact to affect plant growth and development. Studies have reported varying results, including additive effects [17-18], independence of the interaction [19], and even antagonistic effects [20-21]. Wheat varieties with enhanced salt tolerance are expected to accumulate lower levels of boron when exposed to boron toxicity [22]. Previous research has shown differences in salinity and boron accumulation among wheat cultivars [23]. As a result, selecting appropriate wheat genotypes is crucial for successful crop production with improved yield quality in saline, toxic, or boron-deficient conditions. This study investigates the morphological attributes responses of wheat HUW 234 genotypes, differing in salinity tolerance when exposed to externally applied soil salinity, varying boron levels, and there.

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## MATERIALS AND METHODS

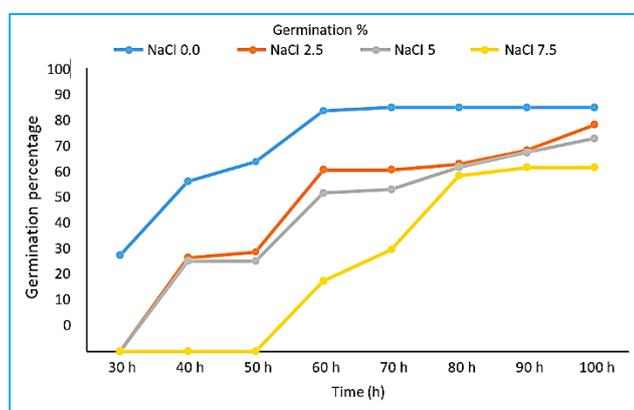
Experiments were conducted during the rabi seasons of 2014-15 and 2015-16 using the wheat variety HUW 234. The experiments were carried out on the experimental farm of the Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, Uttar Pradesh. Screening of Sodium Chloride (NaCl) Concentration: Various concentrations of NaCl (0, 2.5, 5.0, and 7.5 mg/kg soil) were employed to induce stress conditions in the wheat variety. Initially, the different Sodium Chloride (NaCl) concentrations were applied to sterilized soil, which was then saturated with the salt and allowed to dry. After the soil reached dryness, 30 seeds of the wheat variety HUW 234 were planted in each pot in replicates. The germination percentage of wheat was monitored up to 100 hours from the sowing time. Simultaneously, amylase activity during the wheat seedling stage was analyzed at 60 hours, 80 hours, and 100 hours. Based on the results of these experiments, two Sodium Chloride (NaCl) concentrations, 2.5 mg and 5.0 mg per kg of soil, were chosen for further investigation. Screening of Boron (B) Concentration: Various concentrations of boron (ranging from 0 to 5.0 mg B/kg soil in increments of 0.5 mg) were utilized to induce stress conditions in the wheat variety. The different boron concentrations were initially applied to sterilized soil, which was then saturated with the boron and allowed to dry. Following the soil's dryness, 30 seeds of the wheat variety HUW 234 were sown in each pot in replicates. The germination percentage of wheat was measured up to 100 hours from the sowing time. Simultaneously, amylase activity during the wheat seedling stage was analyzed at 60 hours, 80 hours, and 100 hours. Based on the results of these experiments, two boron

concentrations, 2.0 mg and 3.5 mg per kg of soil, were selected for further investigation.

## RESULTS AND DISCUSSION

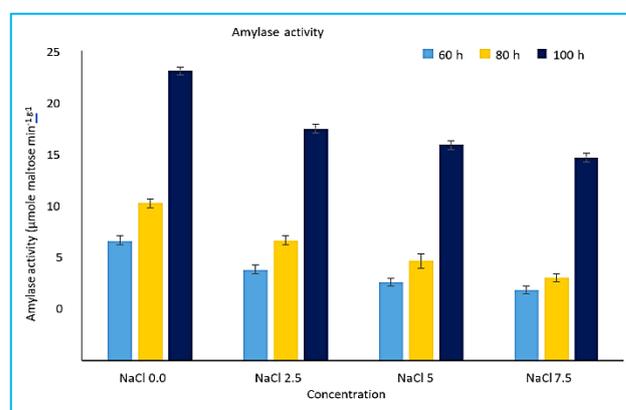
### Screening of sodium chloride concentration on germination percentage and amylase activity

The results indicate that increasing concentrations of sodium chloride (NaCl) in the soil had a significant negative impact on the germination percentage of the wheat variety HUW 234 over time. In the absence of NaCl, the germination percentage reached up to 86% after 100 hours. However, with increasing NaCl concentrations of 2.5 mg, 5.0 mg, and 7.5 mg per kg of soil, the germination percentages decreased to 80%, 75%, and 65%, respectively, at 100 hours. This demonstrates the detrimental effect of soil salinity on wheat seed germination. Simultaneously, the amylase activity in the wheat seedlings was analyzed during the early growth stages. Amylase activity increased with time, with the highest increment observed in the absence of Sodium Chloride (NaCl), reaching 23  $\mu\text{mole maltose min}^{-1} \text{g}^{-1}$  at 100 hours. In contrast, the lowest amylase activity of approximately 16  $\mu\text{mole maltose min}^{-1} \text{g}^{-1}$  was found in wheat seeds treated with 7.5 mg Sodium Chloride (NaCl) per kg of soil. This indicates that increased soil salinity negatively impacts the amylase activity, which is crucial for the breakdown of starch and mobilization of stored nutrients during germination and early seedling growth. Based on these findings, two NaCl concentrations, 2.5 mg and 5.0 mg per kg of soil, were selected for further investigation, as they represent moderate and high salinity levels that significantly impacted the germination and amylase activity of the wheat variety.



The figure shows the germination percentage during the germination stage in the wheat HUW 234 variety under different concentrations of Sodium Chloride (NaCl) at different time intervals

Fig 1 Screening of salt concentration against germination percentage in wheat crop



The figure shows amylase activity during the germination stage in wheat HUW 234 under different concentrations of Sodium Chloride (NaCl) at different time intervals. Data represented as mean and SE

Fig. 2 Effect of different NaCl concentration on amylase activity in germinating wheat seeds

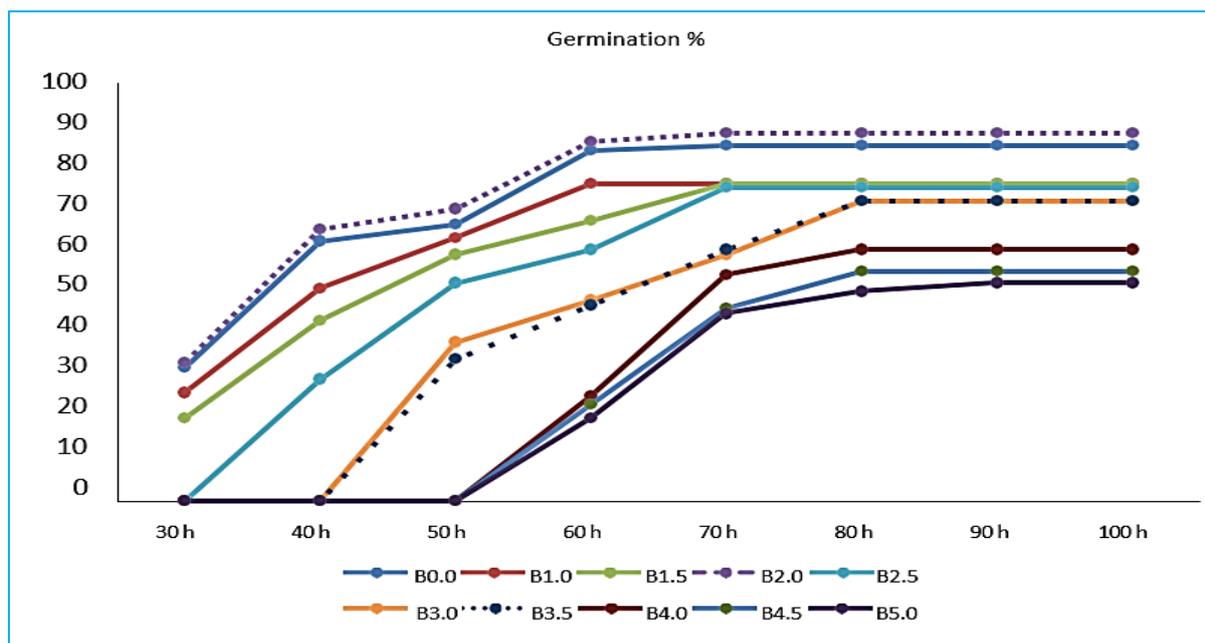
### Screening of boron concentration on germination percentage and amylase activity

The results showed that the germination percentage of the wheat variety HUW 234 exhibited a significant decline with increasing concentrations of boron in the soil, except at 2.0 mg B  $\text{kg}^{-1}$  soil, where an increase was observed over time. The maximum germination percentage of 88% was achieved at 2.0 mg B  $\text{kg}^{-1}$  soil after 100 hours. In contrast, at higher boron concentrations of 4.5 and 5.0 mg B  $\text{kg}^{-1}$  soil, the germination percentages were considerably lower, reaching only 52% and 52%, respectively, at 100 hours. The amylase activity during the wheat seedling stage also showed a similar trend [24]. The

maximum amylase activity of 26  $\mu\text{mole maltose min}^{-1} \text{g}^{-1}$  was observed at 2.0 mg B  $\text{kg}^{-1}$  soil at 100 hours, followed by the control treatment (0.0 mg B  $\text{kg}^{-1}$  soil). The lowest amylase activity of approximately 13.23  $\mu\text{mole maltose min}^{-1} \text{g}^{-1}$  was found in wheat seeds treated with 5.0 mg B  $\text{kg}^{-1}$  soil. Based on these results, two concentrations of boron, 2.0 mg and 3.5 mg per kg of soil, were selected for further investigation, as they represent the optimal and moderately toxic levels of boron for the wheat variety HUW 234 [25]. The findings from the screening experiments on the effects of sodium chloride (NaCl) and boron concentrations on the germination percentage and amylase activity of the wheat variety HUW 234 provide

valuable insights into the physiological responses of wheat to these abiotic stresses. The selected sodium chloride (NaCl) and boron concentrations will be used in subsequent experiments to elucidate the underlying mechanisms and potential strategies

for improving wheat tolerance to these stresses [26]. The selected concentrations will be used in subsequent experiments to further explore the underlying mechanisms and potential strategies for enhancing wheat tolerance to these stresses.



The figure shows the germination percentage during the germination stage in the wheat HUW 234 variety under different concentrations of Boron at different time intervals

Fig 3 Screening of boron concentrations against germination percentage in wheat

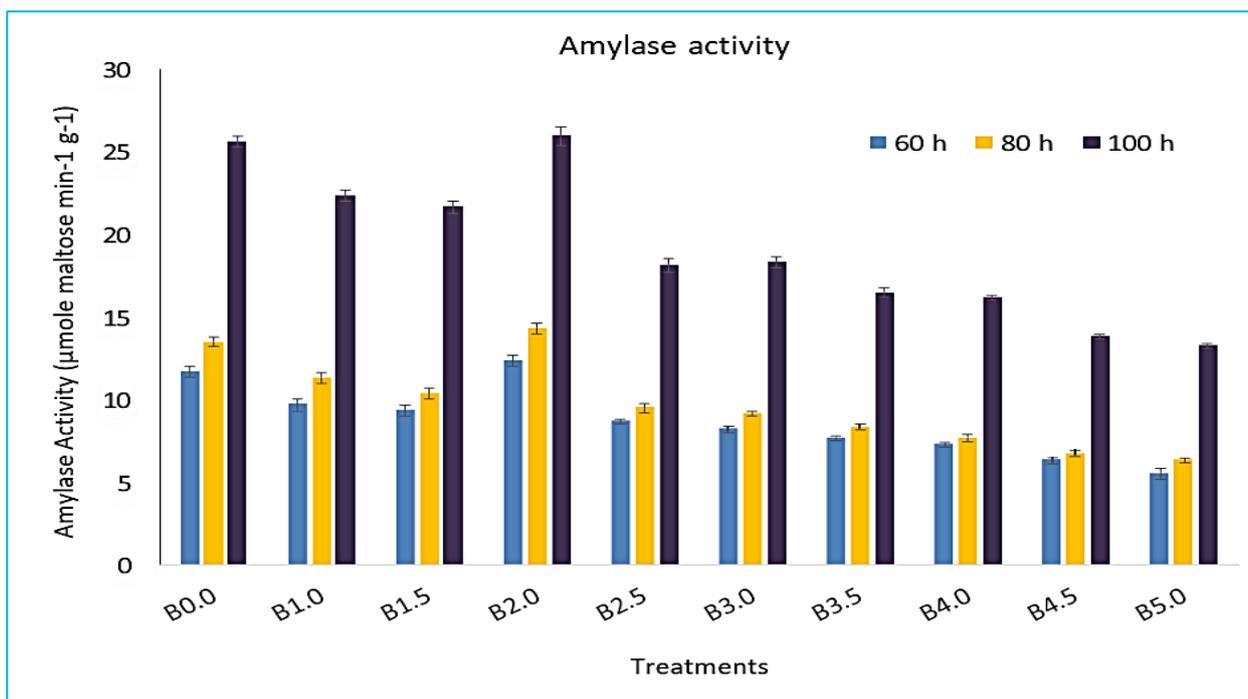


Figure shows amylase activity during the germination stage in wheat HUW 234 under different concentrations of Boron at different time intervals. Data represented as mean and SE

Fig 4 Effect of different boron concentrations on amylase activity in germinating wheat seeds

## CONCLUSION

This study investigated the impact of two key abiotic stresses, soil salinity (NaCl) and boron toxicity, on the seed germination and early seedling growth of the wheat variety HUW 234. The results clearly demonstrate that increasing soil salinity, as indicated by higher NaCl concentrations, has a

detrimental effect on the germination percentage and amylase activity of this wheat variety. The reduction in these key physiological parameters under salt stress can be attributed to the osmotic and ionic effects of NaCl, which disrupt water uptake and cellular homeostasis, ultimately impairing seed germination and early seedling establishment. In contrast, the impact of boron stress on the wheat variety HUW 234 exhibited

a more complex relationship. While excessive boron levels (4.5 and 5.0 mg B kg<sup>-1</sup> soil) had a negative effect on germination and amylase activity, the optimal boron concentration of 2.0 mg B kg<sup>-1</sup> soil actually enhanced these physiological parameters compared to the control. This suggests that a moderate level of boron can be beneficial for this wheat variety, likely by stimulating processes such as cell wall formation, membrane integrity, and carbohydrate metabolism. These findings indicate that the wheat variety HUW 234 has a limited tolerance to high soil salinity, but a relatively narrow optimal range for boron requirements. Developing strategies to improve its salt and boron tolerance, such as the use of salt-tolerant rootstocks,

application of Osmo protectants, or genetic improvements through breeding and biotechnology, could be crucial for enhancing the productivity and adaptability of this wheat variety in saline-affected and boron-rich regions. Further research is warranted to elucidate the underlying physiological and molecular mechanisms governing the responses of HUW 234 to these abiotic stresses, which could provide valuable insights for designing effective stress management approaches. Evaluating the performance of this wheat variety under field conditions and exploring strategies for mitigating salt and boron stress would be important next steps in ensuring its successful cultivation in challenging environmental conditions.

## LITERATURE CITED

1. Flowers TJ. 2004. Improving crop salt tolerance. *Journal of Experimental Botany* 55: 307-319.
2. Rengasamy P. 2002. Transient salinity and subsoil constraints to dryland farming in Australian sodic soils: An overview. *Aust. Jr. Exp. Agric.* 42: 351-361.
3. Munns R, Tester M. 2008. Mechanisms of salinity tolerance. *Ann. Rev. Plant Biology* 59: 651-681.
4. Yamaguchi T, Blumwald E. 2005. Developing salt-tolerant crop plants: challenges and opportunities. *Trends Plant Science* 10: 615-620.
5. Sairam RK, Srivastava GC. 2002. Changes in antioxidant activity in sub-cellular fractions of tolerant and susceptible wheat genotypes in response to long term salt stress. *Plant Science* 162: 897-904.
6. Yamaguchi T, Blumwald E. 2005. Developing salt-tolerant crop plants: challenges and opportunities. *Trends Plant Science* 10(12): 615-620.
7. Teakle NL, Tyerman SD. 2010. Mechanisms of Cl<sup>-</sup> transport contributing to salt tolerance. *Plant Cell Environ.* 33: 566-589.
8. Xu G, Magen H, Tarchitzky J, Kafkafi U. 2000. Advances in chloride nutrition of plants. *Adv. Agronomy* 68: 97-150.
9. White PJ, Broadley MR. 2001. Chloride in soils and its uptake and movement within the plant: a review. *Annals of Botany* 88: 967-988.
10. El-Hamdaoui A, Redondo-Nieto M, Rivilla R, Bonilla I, Bolaños L. 2003. Effects of boron and calcium nutrition on the establishment of the Rhizobium leguminosarum-pea (*Pisum sativum*) symbiosis and nodule development under salt stress. *Plant Cell Environ.* 26: 1003-1011.
11. Dell B, Huang LB. 1997. Physiological response of plants to low boron. *Plant Soil* 193: 103-120.
12. Liu P, Yang YS, Xu GD, Fang YH, Yang YA, Kalin RM. 2005. The effect of molybdenum and boron in soil on the growth and photosynthesis of three soybean varieties. *Plant Soil Environ.* 51: 197-205.
13. Chen LS, Han S, Qi YP, Yang LT. 2012. Boron stresses and tolerance in citrus. *Afr. Jr. Biotechnology* 11: 5961-5969.
14. Sheng O, Song SW, Peng SA, Deng XX. 2009. The effects of low boron on growth, gas exchange, boron concentration and distribution of 'Newhall' navel orange (*Citrus sinensis* Osb.) plants grafted on two rootstocks. *Sci. Hortic.* 121: 278-283.
15. Han S, Tang N, Jiang HX, Yang LT, Lee Y, Chen LS. 2009. CO<sub>2</sub> assimilation, photosystem II photochemistry, carbohydrate metabolism and antioxidant system of citrus leaves in response to boron stress. *Plant Science* 176: 143-153.
16. Grieve CM, Poss JA. 2000. Wheat response to interactive effects of boron and salinity. *Jr. Plant Nutrition* 23: 1217-1226.
17. Wimmer MA, Muñhling KH, Lauchli A, Brown PH, Goldbach HE. 2003. The interaction between salinity and B toxicity affects the subcellular distribution of ions and proteins in wheat leaves. *Plant Cell Environ.* 26: 1267-1274.
18. Masood S, Wimmer MA, Witzel K, Zorb C, Muhling KH. 2012. Interactive effects of high boron and NaCl stresses on subcellular localization of chloride and boron in wheat leaves. *Jr. Agron. Crop Science* 198: 227-235.
19. Edelstein M, Ben MH, Cohen R, Burger Y, Ravina I. 2005. Boron and salinity effects in grafted and non-grafted melon plants. *Plant Soil* 269: 273-284.
20. Bastias EI, Gonzalez-Moro MB, Gonzalez-Murua C. 2004. *Zea mays* L. amylacea from the Lluta Valley (Arica Chile) tolerates salinity stress when high levels of boron are available. *Plant Soil* 267: 73-84.
21. Yermiyahu U, Ben-Gal A, Keren R, Reid RJ. 2008. Combined effect of salinity and excess boron on plant growth and yield. *Plant Soil* 304: 73-87.
22. Nable RO, Bañuelos GS, Paull JG. 1997. Boron toxicity. *Plant Soil* 193: 181-198.
23. Wimmer MA, Goldbach HE. 2012. Boron-and-salt interactions in wheat are affected by boron supply. *Jr. Plant Nutr. Soil Science* 175: 171-179.
24. Singh G, Singh P, Dadrwal B, Srivastava J. 2021. Effects of boron on germination percentage and morphological parameters of wheat (*Triticum aestivum* L.). *International Journal of Ecology and Environmental Sciences* 3(2): 78-83.
25. Gholizadeh F, Mirzaghaderi G, Danish S, Farsi M, Marashi SH. 2021. Evaluation of morphological traits of wheat varieties at germination stage under salinity stress. *PLoS One* 16(11): e0258703.
26. Chakraborty P, Bose B. 2020. Effects of magnesium nitrate and boric acid on germination and seedling growth parameters of wheat (*Triticum aestivum* L.) var. HUW-468. *Journal of Pharmacognosy and Phytochemistry* 9(4): 804-808.